(19) World Intellectual Property Organization International Bureau



(43) International Publication Date 25 October 2001 (25.10.2001)

PCT

(10) International Publication Number WO 01/79555 A2

(51) International Patent Classification⁷: C1

C12Q 1/68

(21) International Application Number: PCT/US01/12131

(22) International Filing Date: 13 April 2001 (13.04.2001)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data: 09/549,654

14 April 2000 (14.04.2000) US

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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

 without international search report and to be republished upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: ROLES OF JAK/STAT FAMILY MEMBERS IN TOLERANCE INDUCTION

(57) Abstract: The present invention relates to methods and compositions for reducing immune rejection, for example, transplant or autoimmune disorder-related immune rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can reduce immune rejection. The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is suspected of, being affected by an autoimmune disorder). The present invention is further based, in part, on the discovery, demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue. In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present, e.g., present in an affected tissue.

Roles of Jak/Stat Family Members in Tolerance Induction

1. INTRODUCTION

The present invention relates to methods and compositions for reducing immune rejection, for example, transplant- or autoimmune disorder-related immune rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can reduce immune rejection.

The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is suspected of, being affected by an autoimmune disorder). The present invention is further based, in part, on the discovery, demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue. In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA, protein Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present, e.g., present in an affected tissue.

2. BACKGROUND OF THE INVENTION

Ongoing advances in transplantation, including new immunosuppressive agents and improvements in histocompatibility matching, organ procurement, and surgical techniques, are gradually improving the outcome of clinical transplantation (Hariharan et al, 2000. N Engl J Med 342:605-12). However, chronic allograft rejection remains the prime determinant of long-term graft survival (Paul. L.C., 1999, Kidney International 56:783-793).

Tissue transplantation between genetically nonidentical individuals results in immunological rejection of the tissue through T cell-dependent mechanisms. To prevent allograft rejection, immunosuppressive agents such as calcineurin phosphatase inhibitors and glucocorticosteroids which directly or indirectly interfere with IL-2 signaling are administered to transplant recipients (see, e.g., Borel, J.F., 1989, Pharmacol. Rev. 42:260-372; Morris, P.J., 1991, Curr. Opin. Immunol. 3:748-751; Sigal et al., 1992, Ann. Rev. Immunol. 10:519-560; and L'Azou et al., 1999, Arch. Toxicol. 73:337-345). The most

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commonly used immunosuppressive agents today are cyclosporin A, FK506, and rapamycin. These immunosuppressive agents act indiscriminately on all T cells by impairing T cell receptor ("TCR") signal transduction. Further, since the effect of the immunosuppressive agents is short-lasting, transplant recipients normally require life-long treatment of immunosuppressive agents to prevent transplant rejection. As a result of the long-term nonspecific immunosuppression, these immunosuppressive agents have many serious adverse effects. For example, the administration of cyclosporin A or FK506 to a transplant recipient results in degenerative changes in renal tubules. Transplant recipients receiving long-term immunosuppressive treatment have a high risk of developing infections 10 and tumors. For example, patients receiving immunotherapy are at higher risk of developing lymphomas, skin tumors and brain tumors (see, e.g., Fellstrom et al., 1993, Immunol. Rev. 134:83-98).

An alternative to immunosuppressive agents for the prevention of allograft rejection is the blockage of specific receptors involved in T cell costimulation. T cell activation 15 requires both TCR-mediated signal transduction and simultaneously delivered costimulatory signals. These costimulatory signals are contributed, in part, by the activation of the costimulatory molecule CD28, which is expressed on resting T cells, by CD80 (B7-1) or CD86 (B7-2) expressed on antigen presenting cells (APCs). The activation of the costimulatory molecule CD40, which is expression on antigen presenting cells (i.e., B cells, 20 dendritic cells, and macrophages), by CD40 ligand ("CD40L"), which is expressed on activated T cells, contributes to the upregulation of T cell activation by inducing the expression of B7-1 and B7-2 on antigen presenting cells and the production of certain chemokines and cytokines such as IL-8, MIP-1α, TNF-α, and IL-12 (Cella et al., 1996, J. Exp. Med. 184:747-752: and Caux et al., 1994, J. Exp. Med. 180:1263-1272). The 25 CD40/CD40L interaction also results in the differentiation of T cells to T helper ("TH") type 1 cells in part due to the expression of cytokines such as IL-12 by dendritic cells and macrophages.

CTLA-4 is normally expressed as a membrane-bound receptor on T cells and has been shown to downregulate T cell activation by competing with CD28 for B7-1 and B7-2. 30 The administration of soluble CTLA-4Ig is believed to prevent allograft rejection by competing with CD28 for B7-1 and B7-2. Soluble CTLA-4Ig has been administered to transplant recipients to disrupt the CD28/B7 interaction so that T cell costimulation is blocked and allograft rejection does not occur (Zheng et al., 1999, J. Immunol. 162:4983-4990; Lenschow et al., 1996, Ann. Rev. Immunol. 14:233-258). Unfortunately, CTLA-4Ig 35 has variable efficacy, and typically does not prevent development of chronic rejection.

Anti-CD40L (anti-CD154) monoclonal antibodies have also been administered to transplant recipients to prevent allograft rejection. These antibodies function by blocking the interaction of CD40 on antigen presenting cells (APC) and CD40L on activated T cells. It has recently been shown that graft survival achieved through the use of anti-CD40L monoclonal antibodies results in a significant inhibition of TH1 type cytokines (i.e., IL-2, 5 IL-12, TNF α , and IFN γ), and an increase in the levels of the TH2 type cytokines (i.e., IL-4, and IL-10) in the graft sections (Hancock et al., 1996, Proc. Natl. Acad. Sci. USA 93:13967-13972). Although the administration of anti-CD40L monoclonal antibodies has been shown to result in permanent graft survival when given to mice in combination with 10 donor-specific spleen cells, adverse side effects such as coagulation have also been shown to be associated with the administration of anti-CD40L monoclonal antibodies. Initial clinical trials in adult renal transplant recipients receiving anti-CD40L monoclonal antibody plus glucocorticoids were halted because of thromboembolic complications (Vincent, J., Biogen News, press release, November 2, 1999, www.prnewswire.com), though the extent 15 to which thromoboembolism was attributable to monoclonal antibodies versus non-specific factors in the antibody formulation is unclear (Kawai et al., 2000, Nature Med. 6:114; and Kirk et al., 2000, Nature Med. 6:114). Further, in the primate renal allograft study, concomitant use of mainstream immunosuppressive agents such as FK-506, methylprednisolone and mycophenolate mofetil diminished the efficacy of CD40L (CD154) 20 mAb, though the exact contribution of each of the individual drugs to this reduction in efficacy was not determined (Kirk, A.D., 1999, Nature Medicine 5:686-693.). The results presented herein demonstrate that some, but not all, combinations of CD154 mAb and immunosuppressive agents are antagonistic, and that strategies for design of clinical trials based on use of CD154 mAb can be logically developed by taking into account the extent to 25 which a given drug inhibits induction of CD154.

In addition, no satisfactory methods presently exist for monitoring whether a transplant graft is being accepted or rejected by a recipient. In general, signs of cellular damage within the transplant tissue can be assayed. Alternatively, for tissues such as kidney or liver, physiological function of the transplant tissue can be assayed. Often, however, by the time overt signs of either cellular damage or a decrease in physiological function are detected, the tissue graft is already beyond rescue. This is particularly true in the case of such organ transplants as heart transplants, with which the first overt signs of rejection are often complete failure of the heart's function.

Accordingly, there is a need for improved, safer immunomodulatory treatments that 35 have long-lasting effects for the prevention of transplant rejection. In particular, there is a need for treatments that are more specific and less toxic than the currently available

therapeutic agents. Further, there is also a great need for an improved method for monitoring acceptance of transplant tissue in subject mammals that have undergone a transplant.

2.1. <u>Jak/STAT Signal Transduction</u>

Signal transduction pathways represent molecular solutions to the fact that such molecules as polypeptide hormones, growth factors and cytokines cannot cross the cell membrane, but must activate intracellular signaling molecules to elicit a response in target cells. Among such signal transduction pathways is the Jak/Stat signal transduction pathway. See, e.g., Heim, M.H., 1999, J. Recept. & Sig. Trans. Res. 19:75-120; and Leonard, W.J. & O'Shea, J.J., 1998, Ann. Rev. Immunol. 16:293-322.

While the pathway was originally discovered as part of a study of interferon-induced intracellular signalling, to date, several dozen polypeptide ligands have been identified that activate the Jak/Stat pathway. Defects in the Jak/Stat pathway have been identified in a number of diseases, including leukemias, lymphomas, inherited immunodeficiency syndromes, breast cancer and a form of dwarfism caused by constitutively activation of a Stat by a mutant fibroblast growth factor-receptor.

Stats (Signal transducers and activators of transcription) are phosphoproteins that are transcription factors, and that are activated in response to cytokines, growth factors and interferons. Stats are activated by receptor-associated Janus kinases ("Jaks"), which include Jak1, Jak2, Tyk2, and Jak3. Specifically, a ligand-induced receptor aggregation results in the transphorphorylation and activation of the catalytic activity of the associated Jak. The activated Jak phosphorylates the receptors at multiple sites. Stats are recruited to the multimeric complex consisting of the phosphorylated receptor and catalytically active Jak.

25 The catalytically active Jak phosphorylates tyrosine residues in the carboxy-terminus of the Stats. The phosphorylated Stats form homodimers and heterodimers (Darnell, J.E., 1997, Science 277:1630-1635; and Leonard et al., 1998, Ann. Rev. Immunol. 16:293-322; and Darnell et al., 1994, Science 264:1415-1421). The dimerization of Stats is believed to trigger the dissociation of Stats from the receptor complex and their translocation to the nucleus. In the nucleus, Stat dimers bind to their cognate DNA regulatory elements, which binding results in increased transcription, i.e., transactivation. Thus, the Jak/Stat system provides a method of both signal amplification and transduction.

Seven Stat genes (Stat1, Stat2, Stat3, Stat4, Stat5A, Stat5B, and Stat6) and several Stat isoforms have been discovered, the isoforms resulting from alternative splicing or posttranslational processing (for review see, e.g., Leonard et al., 1998, Ann. Rev. Immunol. 16:293-322). Different Stats are activated in response to different cytokines and growth

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factors. For example, Stat4 has been shown be activated in response to IL-12 induced signal transduction (Thierfelder et al., 1996, Nature 382:171-174; and Kaplan et al., 1996, Nature 382:174-177). Stat6 has been shown to be activated in response to IL-4 and IL-13 induced signal transduction (Takeda et al., 1996, Nature 380:627-630). Certain transcription factors activated in response to a given cytokine have been shown to be important in TH1 and/or TH2 differentiation. Stat4 has been shown to be important in TH1 differentiation and Stat6 has been shown to be important in TH2 differentiation (see, e.g., Romagnani, S., 1997, Immunology Today 18:263-266; Ray, A. and Cohn, L., 1999, J. Clin. Invest. 104(8):985-993).

With respect to TH1 and TH2, the majority of mature T lymphocytes can be divided into two distinct phenotypes: CD8⁺ cytotoxic T lymphocytes (CTLs), which display the CD8 marker on their cell surface, and CD4⁺ helper T lymphocytes (T helper or TH cells), which display the CD4 marker on their cell surface. TH cells are involved in both humoral (i.e., antibody) and cell-mediated forms of immune response. TH cells have been further 15 categorized into two distinct subpopulations, termed TH1 and TH2 cell subpopulations. These two subpopulations of TH cells have been categorized on the basis of their restricted cytokine profiles and different functions. For example, TH1 cells are known to produce IL-2, IL-12, tumor necrosis factor β ("TNF- β "), and interferon- α ("IFN- α "). TH2 cells are known to produce IL-4, IL-5, IL-10 and IL-13. Inappropriate immune responses have been 20 shown to be associated with various diseases and disorders. For example, an inappropriate TH2-like response has shown to be associated with atopic conditions, such as asthma and allergy (see, e.g., Holgate, S.T., 1997, Lancet 350(suppl. II):5-9; Ray, A. and Cohn, L, supra; Oettgen, H.C. and Geha, R.S., 1999, J. Clin. Invest. 104(7):829-835). Further, an inappropriate TH1-like response has been shown to associated with the pathogenesis of 25 autoimmune diseases such multiple sclerosis, pancreases of insulin-dependent diabetes patients, thyroid glands of Hashimoto's thyroiditis, and gut of Crohn's disease patients.

2.2. NEGATIVE REGULATORS OF THE Jak/STAT SIGNALING PATHWAY

Three protein families have been discovered that negatively regulate cytokine-30 induced Jak/Stat signaling, tyrosine phosphatases SHP1 and SHP2, the suppressors of cytokine signaling ("SOCS"), and protein inhibitors of activated Stats (PIAS). SHP1 and SHP2 bind to phosphorylated tyrosine residues on receptors or Jaks, and inactivate signaling by dephosphorylating them (Haque et al., 1998, J. Biol. Chem. 273:33898-33896; and You et al., 1999, Mol. Cell. Biol. 19:2416-2424).

The SOCS family of proteins have been shown to inhibit the Jak/Stat pathway by 35 inhibiting the activity of the Jaks (Hilton et al., Proc. Natl. Acad. Sci. USA 95:114-119; and

Hilton, 1999, Cell. and Mol. Life Sci. 55:1658-1577). The nature of the interaction between the different receptors, Jaks, and the SOCS is unclear (Hilton, D. J., 1999, Cell. Mol. Sci. 55:1568-1577). SOCS1 have been shown to directly interact with all the Jaks and Tyk2. CIS (Cytokine inducible SH2 containing protein), a member of the SOCS family, on the other hand, was shown to interact with the EPO receptor or the β chain of the IL-3 receptor in a phosphorylation dependent manner, indicating it may act by competing with Stat molecules for binding to receptors (Yoshimura et al., 1995, EMBO J. 14:2816-2826). SOCS1 expression inhibits IL-6, LIF, oncostatin M, IFN-γ, IFN-β, IFN-α, thrombopoeitin, and growth hormone (GH) induced Jak/Stat signaling. SOCS3 expression inhibits IFN-v. 10 IFN-β, IFN-α, GH and leptin.

Four members of the PIAS family have been identified, PIAS1, PIAS3, PIASxa, and PIASx B. PIAS1 was found to bind only to activated Stat1, and PIAS3 to only activated Stat3 (Liu et al., 1998, Proc. Natl. Acad. Sci. USA 95:10626-10631; and Chung et al., 1997, Science 278:1803-1805). PIAS-mediated inhibition of the Jak/Stat signaling pathway. 15 unlike SOCS-mediated inhibition of the Jak/Stat signaling pathway, is very specific. However, unlike some of the SOCS which are elevated rapidly in response to cytokines, the PIAS levels in the cells are more or less constant.

3. SUMMARY OF THE INVENTION

The present invention relates to methods and compositions for reducing immune 20 rejection, for example, transplant- or autoimmune disorder-related immune injury or rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can 25 reduce immune injury.

The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is 30 suspected of, being affected by an autoimmune disorder). In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein, present in an affected tissue. The results presented herein also demonstrate that immune rejection can be monitored by determining the amount of Stat1 35 mRNA or protein, Stat2 mRNA or protein, or Stat3 mRNA or protein present, e.g., present in an affected tissue. The present invention is further based, in part, on the discovery,

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demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue.

Thus, in one aspect, the invention relates to methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, wherein said method comprises: determining the amount of at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or (iv) SOCS3 mRNA or SOCS3 protein, present in a transplant sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two, at 10 least three, or each of (i) to (iv) present in the transplant sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 15 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment, a method for monitoring acceptance of a transplant in a 20 subject mammal that has undergone a transplant comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a transplant sample from the subject. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

The methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant can further comprise assaying the transplant sample for evidence of 25 lymphocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized to evaluate internationally recognized and used diagnostic criteria for the evaluation of graft rejection, which include features specific for each organ involved. For example, immunohistologic evaluation of such tissues, via, e.g., use of labeled antibody techniques 30 to localize and quantitate gene expression. The evaluation of such criteria can, therefore, be enhanced by, for example, localization of Stat4, Stat6, SOCS1 and/or SOCS3 proteins, and/or detection of corresponding mRNAs via, e.g., in situ hybridization.

Such methods can also further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding pre-transplant subject 35 sample or a subject blood sample. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is greater than, or the amount of Stat6

mRNA or protein in the transplant sample is less than, that of the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is less than, or the amount of Stat6 mRNA or protein in the transplant sample is equal to or greater than that of the control sample, such a result indicates that acceptance of the transplant has occurred, is being induced or is being maintained. In instances wherein the ratio of Stat4 to Stat6 in the transplant sample is greater than or equal to that in the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. In 10 instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the control sample, such a result indicates that acceptance of the transplant has occurred, has been induced or is being maintained.

In another aspect, the invention relates to methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, wherein said method 15 comprises: determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, or (iii) Stat3 mRNA or Stat3 protein, present in a cell sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two or each of (i) to (iii) present in the sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined 20 via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a particular embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, 25 the cell sample is a transplant sample obtained within 2 to 3 days post-transplantation. In an alternative embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, the cell sample is a subject blood sample.

Such Stat 1-, Stat-2, and/or Stat 3-related methods can also further comprise 30 comparing the amount determined to that present in a control sample, for example, a corresponding pre-transplant subject sample or, in the case of embodiments wherein the cell sample is a transplant sample obtained within 2-3 days post-transplantation, a subject blood sample. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the cell sample is greater than that of the control sample, such a result indicates that acceptance 35 of the transplant has not occurred, has not been induced or is not being maintained. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the transplant

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sample is less than that of the control sample, such a result indicates that acceptance of the transplant has occurred, is being induced or is being maintained.

In another aspect, the invention relates to methods for monitoring an autoimmune disorder in a subject mammal, wherein said method comprises: determining the amount of at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or (iv) SOCS3 mRNA or SOCS3 protein, present in a sample from a subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected by the disorder. In alternate embodiments, such methods comprise determining the amount of at least two, at 10 least three, or each of (i) to (iv) present in the sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be 15 determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment, a method for monitoring an autoimmune disorder in a subject mammal comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and 20 Stat6 protein present in a sample from the subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected by the disorder. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

The methods for monitoring an autoimmune disorder in a subject mammal can 25 further comprise assaying the sample for evidence of leukocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized. Alternatively, standard techniques can be utilized to assay (e.g., in serum) for the presence of autoimmune antibodies associated with the particular autoimmune disorder of interest. There are internationally used diagnostic 30 criteria for evaluation of graft rejection, with features specific for each organ. The immunohistologic evaluation of such tissues, i.e., use of unlabeled-antibody techniques to localize and quantitate gene expression, can be enhanced by localization of Stat4 and Stat6 proteins, or detection of corresponding mRNAs by in situ hybridization.

Such methods for monitoring an autoimmune disorder in a subject mammal can 35 further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding tissue not affected by the disorder or a subject blood

sample. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is greater than, or the amount of Stat6 mRNA or protein in the sample is less than, that of the control sample, such a result indicates that the subject mammal exhibits or continues to exhibit the disorder. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is less than, or the amount of Stat6 mRNA or protein in the sample is equal to or greater than that of the control sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective. In instances wherein the ratio of Stat4 to Stat6 in the sample is greater than or equal to that in the control sample, such a result indicates that the subject mammal 10 exhibits or continues to exhibit the disorder. In instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective.

The methods for monitoring transplant acceptance or monitoring an autoimmune disorder can be performed with kits designed for carrying out such methods. As such, the 15 present invention also relates to kits for monitoring transplant acceptance and autoimmune disorders.

In yet another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount 20 of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control activated T cell sample that has not been contacted with the test 25 compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and 30 comparing the amounts to those present in the control sample.

In certain embodiments, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In 35 any such embodiment wherein a Stat protein amount is determined, the amount determined

can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount of Stat4 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (b) to those present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce ammune rejection is identified.

In another preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c) comparing the ratio in (b) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises:

(a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of at least one of the following:

(i) Stat1 mRNA or Stat1 protein,

(ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or 25 (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding resting T cell sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the amount of (i), (iii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to 30 reduce immune rejection is identified. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and comparing the amounts to those present in the control sample.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of

Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample. Further, in certain embodiments, the resting T cell is a primary T cell, and in other embodiments, the resting T cell is a T cell line.

In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of Stat4 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (b) to those present in a corresponding control resting T cell sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c) comparing the ratio in (b) to that present in a corresponding control resting T cell sample that has been contacted with a T cell activator, but has not been contacted with the test compound, so that if the ratio in (a) is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In preferred embodiments, the cytokine is IL-2, IL-14, IL-12, or IL-13.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine; (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and (c) comparing the ratio to in (a) to that present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In yet another aspect, the present invention relates to methods for reducing immune rejection in a subject mammal, said methods comprising: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to decrease the level of Stat4 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-κB activation in CD40L⁺ cells.

Alternatively, such methods for reducing immune rejection in a subject mammal can comprise: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to increase the level of Stat6 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said

compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

Such methods for reducing immune rejection in a subject mammal can also comprise: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to decrease the level of Stat4 mRNA or protein and maintain or increase the level of Stat6 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-kB activation in CD40L⁺ cells.

The methods of the present invention for reducing immune rejection can be utilized, e.g., for reducing immune rejection in a subject mammal that has undergone a transplant. For example, such methods can induce tolerance in a subject mammal that has undergone a transplant. The methods of the present invention for reducing immune rejection can also be utilized, e.g., for reducing immune rejection in a subject mammal exhibiting an autoimmune disorder.

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3.1. **DEFINITIONS**

As used herein, the term "transplant" includes any cell, organ, organ system or tissue which can elicit an immune response in a recipient subject mammal. In general, therefore, a transplant includes an allograft or a xenograft cell, organ, organ system or tissue. An allograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of the same species as the recipient. A xenograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of a different species as the recipient.

The term "immune rejection," as used herein, is intended to refer to immune responses involved in transplant rejection, as well as to the concomitant physiological result of such immune responses, such as for example, interstitial fibrosis, chronic graft artheriosclerosis, or vasculitis. The term "immune rejection," as used herein, is also intended to refer to immune responses involved in autoimmune disorders, and the concomitant physiological result of such immune responses, including T cell-dependent infiltration and direct tissue injury; T cell-dependent recruitment and activation of macrophages and other effector cells; and T cell-dependent B cell responses leading to autoantibody production.

The term "transplant rejection," as used herein, refers to T cell-mediated rejection of transplant cells, organs, organ systems or tissue. In general, such transplant rejection generally includes accelerated, acute and chronic rejection. It is intended that the term, as used herein, also refer to graft versus host disease, and the physiological results of such a disorder.

The term "reducing immune rejection," is meant to encompass prevention or inhibition of immune rejection, as well as delaying the onset or the progression of immune rejection. The term is also meant to encompass prolonging survival of a transplant in a subject mammal, or reversing failure of a transplant in a subject. Further, the term is meant to encompass ameliorating a symptom of an immune rejection, including, for example, ameliorating an immunological complication associated with immune rejection, such as for example, interstitial fibrosis, chronic graft atherosclerosis, or vasculitis. The term is also meant to encompass induction of tolerance in a subject mammal that has undergone a transplant.

The term "tolerance," as used herein, refers to a state wherein the immune system of a transplant recipient subject mammal is non-responsive to the transplant. This state is considered donor transplant-specific, and, as such, is distinguished from nonspecific immunosuppression. Operatively, the term as used herein, refers to permanent acceptance of a graft without ongoing immunosuppression, wherein, for example, challenge with a second 15 graft of donor origin (especially when the second graft is of the same tissue as the first graft) should be accepted, and challenge with a third party graft should be rejected.

The term "autoimmune rejection," as used herein, refers to immune responses involved in autoimmune disorders, and the concomitant physiological result of such immune responses.

The term "activated T cell," as used herein, refers to a T cell that expresses antigens indicative of T-cell activation (that is, T cell activation markers). Examples of T cell activation markers include, but are not limited to, CD25, CD26, CD30, CD38, CD69, CD70, CD71, ICOS, OX-40 and 4-1BB. The expression of activation markers can be measured by techniques known to those of skill in the art, including, for example, western 25 blot analysis, northern blot analysis, RT-PCR, immunofluorescence assays, and fluorescence activated cell sorter (FACS) analysis.

The term "resting T cell," as used herein, refers to a T cell that does not express Tcell activation markers. Resting T cells include, but are not limited to, T cells which are CD25, CD69, ICOS, SLAM, and 4-1BB. The expression of these markers can be 30 measured by techniques known to those of skill in the art, including, for example, western blot analysis, northern blot analysis, RT-PCR, immunofluorescence assays, and fluorescence activated cell sorter (FACS) analysis.

The term "T cell activator," as used herein, refers to any compound or factor that is a T cell receptor stimulatory factor, that is, induces T cell receptor signalling. Preferably, the 35 compound or factor also induces co-stimulatory pathways. Non-limiting examples of T cell activators include, but are not limited to, anti-CD3 antibodies (preferably monoclonal

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antibodies) either alone or in conjunction with anti-CD28 antibodies (preferably monoclonal antibodies), or mitogens such as, for example, phorbol 12-myristate 13-acetate (PMA), phytohemagglutinin (PHA) or concanavalin-A (Con-A).

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4. BRIEF DESCRIPTION OF THE DRAWINGS

- Figure 1. Human Stat1 nucleic acid and amino acid sequences (SEQ ID NOs:1, 2, respectively).
- Figure 2. Human Stat2 nucleic acid and amino acid sequences (SEQ ID NOs:3, 4, respectively).
 - Figure 3. Human Stat3 nucleic acid and amino acid sequences (SEQ ID NOs:5,6, respectively).

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- Figure 4. Human Stat4 nucleic acid and amino acid sequences (SEQ ID NOs:7, 8, respectively).
- Figure 5. Human Stat6 nucleic acid and amino acid sequences (SEQ ID NOs:9, 10, 20 respectively).
 - Figure 6. Human SOCS1 nucleic acid and amino acid sequences (SEQ ID NOs:11, 12, respectively).
- Figure 7. Human SOCS3 nucleic acid and amino acid sequences (SEQ ID NOs:13, 14, respectively).
 - Figure 8. Human Jak2 nucleic acid and amino acid sequences (SEQ ID NOs:15, 16 respectively).

- Figure 9. Human Tyk2 nucleic acid and amino acid sequences (SEQ ID NOs:17, 18 respectively).
- Figure 10. Effects of immunosuppressants on CD154 mAb-induced cardiac

 35 allograft survival. Murine recipients were followed for up to 100 days post-transplant, and mean (± SD) cardiac allograft survival are shown (n = 6/group). Statistical analysis (Mann-

Whitney U test) showed that CD154 mAb or combined CD154 and rapamycin (CD154RPM) induced highly significant prolongation of allograft survival (p<0.001) compared to recipients treated with IgG, combined CD154 and cyclosporin A (CD 154/CsA), or CD154 plus methylprednisolone (CD154/MP).

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Figure 11A-11B. Contrasting effects of immunosuppressive agents on activation-induced CD154 expression by CD4+ T cells, as determined by dual color flow cytometry after 7 hr of culture. Except for the control sample depicted in the uppermost section of Panel A, all culture plates were pre-coated with CD3 mAb.

Representative individual samples are shown in panel A, in which the histograms depict CD154 expression by gated CD4+ T cells. The percentage of CD154+ cells and the mean channel fluorescence of CD154 staining are indicated. Panel B shows the average activation-induced CD154 expression determined from 3 independent experiments. Percent CD154 expression (mean ± SD) was calculated relative to the percentage of CD4+ CD154+ cells observed in the absence of immunosuppressive drugs. Cyclosporin A (CsA) and methylprednisolone (MP) significantly inhibited activation-induced CD154 expression (p<0.01).

Figure 12A-12B. Critical role for NF-κB in activation-induced CD154
20 expression by CD4+ T cells in vitro. The experimental design and data presentation arc identical to that shown in Figure 10: splenic cells were derived from either NFκB/p50 KO or control wild-type (WT) mice: Genetic deletion of NFκB/p50 inhibited activation-induced CD154 expression (p<0.02). Inhibition of NF-κB p50 in WT mice by the proteasome antagonists lactacystin (LC) and MG-273 (MG) also significantly blocked activation-induced CD154 expression (p<0.04).

Figure 13. Permanent cardiac allograft survival using CD154 mAb is NF-κB-dependent. Mice were followed for up to 100 days post-transplant and mean (± SD) cardiac allograft survival are shown (n = 6/group). Statistical analysis (Mann-Whitney U test) showed that use of NF-κB/p50 KO mice as recipients, or administration of a lactacystin-derived (LC) proteasome inhibitor to wild-type mice, significantly impaired the efficacy of CD154 mAb therapy (p<0.001).

Figure 14A. Kinetics of Stat RNA expression in heart allografts. Post-35 transplantation expression of Stats at day 1, day 2, day 3, day 5 were studied using RNA obtained from transplanted (Balb/c), native (B6/129), and control (B6/129) hearts of 8-10

weeks old female mice. Equal amounts of heart RNA (25 μg) were loaded onto each lane of three 1.2% agarose-formaldehyde gels. Hybridizations were done with probes specific to the transcriptional activation domains and 3'-untranslated regions of the Stats. The Stat probes described in the Materials and Methods Section of Section 8, below, were used for all the hybridizations. The locations of probes are shown underneath the cDNA line drawings next to the hybridization patterns. Murine GAPDH cDNA fragment was used as a control. The Stats and SOCS listed on the right of the GAPDH hybridization patterns indicate the probes used with the particular membranes. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart. These data demonstrate that in the allografts Stats 1 and 2 increase progresssively, peaking at day 5, whereas Stat3 rises to a plateau level by day 1. All 3 Stats increase within native control hearts by day 5.

Figure 14B. Kinetics of Stat RNA expression in heart allografts. The Stat levels were normalized against the GAPDH values. Normalized Stat levels are graphed as relative optical density (relative to other Stats). Closed, open, and gray bars indicate Stat RNA levels in transplant, native, and control hearts, respectively. Designations: D1, day 1; D2, day2; D3, day3; D5, day5; NAT, native; TRA, transplant; CONT, control.

Figure 15. Kinetics of SOCSCIS RNA expression in heart allografts. Post-20 transplantation expression of SOCS/CIS RNA at day 1, day 2, day 3, and day 5 posttransplantation were studied by deprobing and rehybridizing the same membranes initially used for studying the Stat RNA expression studies described above in Figure 14A. Hybridizations were done with probes specific to the 3'-untranslated regions of the SOCS. The SOCS and CIS probes described in the Materials and Methods Section of Section 8, 25 below, were used for all the hybridizations. The locations of probes are shown underneath the cDNA line drawings next to the hybridization patterns. Murine GAPDH cDNA fragment was used as a control. The SOCS/CIS list on the right of the GAPDH hybridization patterns indicate the probes used with the individual membranes. Designations: N, native hearts; T, transplanted hearts; C, control hearts (B6/129). These 30 data demonstrate that SOCS1 is densely expressed in allografts at day 5, with only minor expression in native heart samples. SOCS3 is well-expressed from day 1 in allografts, with a further increase at day 5; no expression was seen in native hearts. SOCS5 is expressed predominantly as a 4.4 kb species, with a minor species at 3.8 kb, in control and native heart samples. However, in allografts, the 2 SOCS5 species are expressed in approximately equal 35 amounts. By contrast, CIS expression remained unchanged in native hearts but decreased in allografts from day 1 onwards.

Figure 16. The similarity in the expression patterns of Stat4 and SOCS3 RNA in heart allografts. Stat4 and SOCS3 RNA expression in the days that follow the surgery show a high degree of similarity between the patterns of Stat4 and SOCS3 RNAs, albeit differences in their abundance (using Kodak Biomax MR film at '80°C, with intensifying screen, the exposure time for the Stat4 blot was approximately 4 days, and for SOCS3 only 15 hrs).

Figure 17. Stat protein levels following cardiac transplantation. Total proteins from the native and transplant hearts of untreated mice were extracted following transplantation, electrophoresed, transferred onto Immobilon-P membranes and Stat levels analyzed by Western blots as described in Materials and Methods section of Section 8, below. To identify Stats correctly, cell extracts recommended as positive controls by the antibody suppliers were used in the Western blots, together with Precision Protein standards. Designations: N, native hearts; T, transplant hearts; C, control hearts (Balb/c or B6/129), HSC 70, constitutive heat-shock protein. Molecular weights of the Stats are shown on the right. The list on the right of the HSC70 patterns indicates which Stat antibody was used with the individual membranes. These data demonstrate a general agreement with the mRNA data, and show that all of the Stats except Stat5 recrease in allografts just prior to rejection (i.e. at day 5), with the greatest and most allograft-specific expression being found for Stat4.

Figure 18. Phosphorylated Stat levels following cardiac transplantation.

Proteins were prepared and Stat levels analyzed by Western blotting as described in

25 Materials and Methods section of Section 8, below. Designations: N, native hearts; T, transplant hearts; C, control hearts (Balb/c or B6/129), HSC 70, constitutive heat-shock protein. Molecular weights of the Stats are shown on the right. These data demonstrate an increase in pStat1 which parallels the rise in Stat1 levels.

Figure 19. Stat RNA expression in day 5 post-transplant heart allografts and the effect of anti-CD40L antibody MR-1 on Stat expression. Expression of Stat1, Stat2, Stat3, Stat4, Stat5A and Stat6 RNA was studied using RNA from transplanted (Balb/c) native (B6/129), and control (B6/129) hearts of 8-10 wks old female mice. Equal amounts of heart RNA (25 μg) were loaded onto each lane of four 1.2% agarose-formaldehyde gels, blotted and analyzed as explained in the Materials and Methods section presented in Section 8, below. The Stat probes described in the Materials and Methods section presented in

Section 8 were used for all the hybridizations. Murine GADPH cDNA fragment was used as a control. The size ofthe Stat RNAs are shown on the right. The Stat and SOCS list on the right of the GAPDH hybridization patterns indicate the probes used with the particular membranes. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart. These data demonstrate that CD40L mAb suppresses expression of each of the Stat mRNA.

Figure 20. SOCS/CIS RNA expression in day 5 post-transplant heart allografts and the effect of anti-CD40L antibody MR-1 on SOCS/CIS expression. Expression of SOCS1, SOCS2, SOCS3, and CIS RNA was studied by deprobing and rehybridizing the same membranes initially used for studying the Stat RNA expression. Hybridizations were done with probes specific 3'-untranslated regions of the SOCS. The locations of probes are shown on the right. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart. These data demonstrate that SOCS1 and SOCS3 levels increased during allograft rejection, but CD40L mAb prevented this induction. SOCS2 levels remained constant. CIS levels were decreased in transplants, regardless of CD40L mAb therapy.

Figure 21. The effect of treatment with the anti-CD40L monoclonal antibody MR-1 on the levels of Stats. Mice were given a single dose injection (DST plus IgG or DST plus MR-1) the day of the transplant surgery. Proteins were extracted from the native and transplanted hearts of both the IgG and MR-1 treated groups (2 mice/group) 5 days and 7 days following transplant surgery. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart. Molecular weights of the Stats are shown on the right. These data demonstrate that CD40L mAb suppresses intragraft levels of each of the Stat proteins except that of Stat6, which at day 7 in this series was increased post-CD40L mAb therapy.

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transplantation. Stat levels were studied in protein extracts from isografts (both donor and recipient mice were Balb/c) and allografts (donor mice were Balb/c and recipient mice were B6/129). Proteins were prepared and Stat, Bax, and HSC70 levels were analyzed by Western blotting as described in Materials and Methods section of Section 8, below. Designations: N, native hearts; T, transplant hearts; C, control hearts (Balb/c or B6/129), HSC 70, constitutive heat-shock protein. Molecular weights of the Stats are shown on the right. The list on the right of the HSC70 patterns indicates which Stat antibody was used with the individual membranes. These data demonstrate that isografts showed only low levels of Stats, whereas allografts had progressive increases in Stat proteins, peaking at day

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5. This pattern of induction was allo-specific and differed from that of an unrelated protein, Bax, which increased in both isografts and allografts.

5. DETAILED DESCRIPTION OF THE INVENTION

5.1. Methods and Compositions for Monitoring Transplant Acceptance and Autoimmune Disorders

As demonstrated below, immune rejection can be monitored by determining the 10 amount of particular members of the Jak/Stat signal transduction pathway present withm an Specifically, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present in an affected tissue. In particular, the results presented herein demonstrate that immune rejection 15 can be monitored by determining the amount of Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein, present in an affected tissue. The results presented herein also demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, or Stat3 mRNA or protein present, e.g., present in an affected tissue.

The term "affected tissue," as used herein, refers to a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system. For example, such an affected tissue can include, but is not limited to, heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, or intestine transplant cells, tissues, organs or organ system. The term "affected tissue," as used herein, also refers to a cell, tissue, organ or organ system that is, 25 or is suspected of, being affected by an autoimmune disorder. For example, such an affected tissue can include, but is not limited to, a cell, tissue, organ, or organ system involved in systemic lupus erythematosus, glomerulonephritis, rheumatoid arthritis, Wegener's granulomatosis, chronic active hepatitis, or vasculitis.

Thus, aspects of the present invention relate to methods and compositions for 30 monitoring such immune rejection. In particular, such methods and compositions can relate, for example, to methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, and can also relate, for example, to methods for monitoring an autoimmune disorder in a subject mammal being treated for or suspected of exhibiting an autoimmune disorder. Such methods and compositions are discussed in detail 35 herein.

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In one aspect, the invention relates to methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, wherein said method comprises: determining the amount of at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or (iv) SOCS3 mRNA or SOCS3 protein, present in a transplant sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, or each of (i) to (iv) present in the transplant sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the

In a preferred embodiment, a method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a transplant sample from the subject. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

Such methods can also further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding pre-transplant subject sample (e.g., a sample from a corresponding pre-transplant cell, tissue, organ, or organ system) or a subject blood sample.

In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is greater than, or the amount of Stat6 mRNA or protein in the transplant sample is less than, that of the control sample, such a result indicates that acceptance of the transplant has not been induced or is not being maintained. Likewise, in instances wherein the ratio of Stat4 to Stat6 in the transplant sample is greater than or equal to that in the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. Such results suggest a course of action that can include, for example, administration of a high dose of immunosuppressive drugs (e.g., a high dose of corticosteroids, in, for example, the form of a single bolus intravenous injection) and /or administration of compounds to effectuate T cell depletion, including but not limited to administration of anti-CD3 antibodies.

In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is less than, or the amount of Stat6 mRNA or protein in the transplant

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sample is equal to or greater than that of the control sample, such a result indicates that acceptance of the transplant has occurred, is being induced or is being maintained. Likewise, in instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the control sample, such a result indicates that acceptance of the transplant has occurred, has been induced or is being maintained.

The methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant can further comprise assaying the transplant sample for evidence of lymphocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized to evaluate internationally recognized and used diagnostic criteria for the evaluation of graft rejection, which include features specific for each organ involved. For example, for evaluation of heart allograft transplants see, e.g., Billingham, M.E., 1990, J. Heart Transplant. 9(3 Pt 2):272-6. For evaluation of renal allografts see, e.g., Racusen et al., 1999, Kidney Int. 55(2):713-23. In one non-limiting embodiment, immunohistologic evaluation of transplant tissues (such as heart or kidney) can be performed via, e.g., use of labeled antibody techniques to localize and quantitate gene expression. The evaluation of such criteria can, therefore, be enhanced by, for example, localization of Stat4, Stat6, SOCS1 and/or SOCS3 proteins, and/or detection of corresponding mRNAs via, e.g., in situ hybridization.

Additionally, methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant can comprise: determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, or (iii) Stat3 mRNA or Stat3 protein, present in a cell sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two or each of (i) to (iii) present in the sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a particular embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, the cell sample is a transplant sample obtained within 2 to 3 days post-transplantation. In an alternative embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, the cell sample is a subject blood sample.

Such Stat 1-, Stat-2, and/or Stat 3-related methods can also further comprise comparing the amount determined to that present in a control sample, for example, a corresponding pre-transplant subject sample or, in the case of embodiments wherein the cell sample is a transplant sample obtained within 2-3 days post-transplantation, a subject blood sample. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the cell sample is greater than that of the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the transplant sample is less than that of the control sample, such a result indicates that acceptance of the 10 transplant has occurred, is being induced or is being maintained. Such results suggest a course of action that can include, for example, administration of a high dose of immunosuppressive drugs (e.g., a high dose of corticosteroids, in, for example, the form of a single bolus intravenous injection) and /or administration of compounds to effectuate T cell depletion, including but not limited to administration of anti-CD3 antibodies.

Methods for monitoring acceptance of a transplant can be performed at any point post-transplantation. In a preferred embodiment, monitoring is performed daily during the first week post-transplant, followed by weekly monitoring until approximately one month post-transplant, followed by monthly monitoring until approximately one year posttransplant. It is understood, of course, that the frequency of monitoring can, at least in part, 20 depend upon the particular situation (e.g., the nature of the graft, overall health of the recipient subject mammal, the particular immunotherapeutic, immunosuppressive, or immunomodulatory treatment being administered, etc.).

In another aspect, the invention relates to methods for monitoring an autoimmune disorder in a subject mammal, wherein said method comprises: determining the amount of 25 at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or (iv) SOCS3 mRNA or SOCS3 protein, present in a sample from a subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected by the disorder. In alternate embodiments, such methods comprise determining the amount of at least two, at 30 least three, or each of (i) to (iv) present in the sample. In certain embodiments, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined.

In a preferred embodiment, a method for monitoring an autoimmune disorder in a subject mammal comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and 35 Stat6 protein present in a sample from the subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected

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by the disorder. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

Such methods for monitoring an autoimmune disorder in a subject mammal can further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding tissue not affected by the disorder or a subject blood sample.

In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is greater than, or the amount of Stat6 mRNA or protein in the sample is less than, that of the control sample, such a result indicates that the subject mammal exhibits or continues to exhibit the disorder. Likewise, in instances wherein the ratio of Stat4 to Stat6 in the sample is greater than or equal to that in the control sample, such a result indicates that the subject mammal exhibits or continues to exhibit the disorder. Such results suggest a course of action that can include, for example, the need to increase immunosuppression, as might be undertaken by bolus intravenous steroids (e.g., methylprednisolone) or use of a CD3 mAb such as OKT3.

In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is less than, or the amount of Stat6 mRNA or protein in the sample is equal to or greater than that of the control sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective. Likewise, in instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective.

The methods for monitoring an autoimmune disorder in a subject mammal can further comprise assaying the sample for evidence of leukocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized. Alternatively, standard techniques can be utilized to assay (e.g., in serum) for the presence of autoimmune antibodies associated with the particular autoimmune disorder of interest. For example, there are standard immunohistology methods for detection of autoantibodies directed against a particular tissue (e.g., anti-glomerular basement membrane, anti-parietal cell, anti-thyroid and anti-islet etc.), as well as assays for their detection in serum (e.g., rheumatoid factor assay and anti-double-stranded DNA antibodies). See, e.g., Manual of Clinical Laboratory Immunology (NR Rose, H Friedman, JL Fahey eds. 1986, Am Soc Microbiol, Washington, DC; Diagnostic Immunopathology (RB Colvin, et al., eds., 1995, Raven Press, New York).

The methods described herein can be performed using a sample from any subject mammal that has undergone a transplant or either exhibits or is suspected of exhibiting an

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autoimmune disorder. Preferably, the mammal is a human, however, such subject mammals can also include, but are not limited to, pigs, dogs, cats, horses, cattle, sheep, mice, rats, and rabbits.

It is noted that such methods for monitoring transplant acceptance and for monitoring autoimmunde disorders can be used to determine whether a subject can be effectively treated with a specific agent or class of agents intended to promote transplant acceptance or to treat the autoimmune disorder. Thus, in one embodiment, the present invention provides such methods for determining whether a subject can be effectively treated with an agent for an autoimmune disorder or for reducing immune rejection. 10 Monitoring the influence of agents (e.g., drugs and compounds) on the expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 mRNA or polypeptide can, therefore, be applied in basic drug screening, preclinical studies, clinical trials and during therapeutic treatment regimens designed to reduce immune rejection or to ameliorate a symptom of an autoimmune disorder.

The methods described herein comprise determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, and/or SOCS3 mRNA or protein present in a sample. Standard techniques, as described below, can routinely be utilized to determine these amounts. In general, such methods of the invention can routinely be performed using 20 standard techniques for detecting the presence or absence of a polypeptide or nucleic acid of the invention in a biological sample. This involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or an agent capable of detecting the polypeptide or mRNA such that the presence of a polypeptide or nucleic acid of the invention is detected in the biological sample.

When comparing levels, such comparisons can be either quantitative or qualitative. Thus, in qualitative instances, for example, in instances wherein a control sample is determined to contain none of a given molecule (that is, Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, SOCS3 mRNA or protein) and the molecule is determined to be present in the test sample (that is, either a transplant or autoimmune sample), the amount of the molecule in 30 the test sample is greater than that present in the control sample. In quantitative instances wherein both the control and test samples are determined to contain a given molecule, using standard techniques, the amount in the test sample can routinely be determined to be greater than, equal to, or less than that of the control sample. Likewise, using standard techniques, the ratio of Stat4 to Stat6 mRNA or protein present in test and control samples can routinely 35 be determined. In general, the amount of a given molecule in test and control samples will

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differ by at least 2-fold, and in certain instances, 2.5-fold, at least 3-fold, at least 4-fold, at least 5-fold, or at least 10-fold.

With respect to methods for monitoring acceptance of a transplant, such methods can be performed on any transplant from which a sample to be analyzed can be obtained. Such transplants include, but are not limited to, heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, or intestine transplants.

Likewise, with respect to methods for monitoring autoimmune disorders, such methods can be performed for any autoimmune disorder (or suspected autoimmune disorder) for which a sample of an affected tissue (or a tissue suspected of being affected) can obtained. Such autoimmune disorders include, but are not limited to, systemic lupus erythematosus, glomerulonephritis, rheumatoid arthritis, Wegener's granulomatosis, chronic active hepatitis, and vasculitis

Methods for obtaining samples from a recipient transplant subject mammal or from a subject mammal exhibiting or suspected of exhibiting an autoimmune disorder are well known to those of skill in the art. Such methods can include biopsy methods, such as, for example, standard needle or punch biopsy methods. In certain embodiments, a particular subset of the sample can be isolated for the analysis. For example, a particular subset of a transplant or autoimmune disorder sample containing cell types of interest (e.g., leukocyte cell types) can be isolated. Such isolation can performed utilizing standard techniques such as, for example, laser microdissection (see, e.g., Fend et al., 1999, Am. J. Pathol. 154(1):61-6; Schutze et al., 1998, Nat. Biotechnol. 16(8):737-42; and Simone et al., 1998, Trends Genet. 14(7):272-6).

Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and SOCS3 nucleic acid and amino acid sequences are well known to those of skill in the art. For Stat1, see, for example, Schindler et al., 1992, Proc. Natl. Acad. Sci. U.S.A. 89:7836-7839 and Zhong et al., 1994, Science 264:95-98. For Stat2, see, for example, Yan et al., 1995, Nucleic Acids Res. 23(3):459-463, Bluyssen and Levy, 1997, J. Biol. Chem. 272(7):4600-4605, and Paulson et al., 1999, J. Biol. Chem. 274(36):25343-25349. For Stat3, see, for example, Ripperger, 1995, J. Biol. Chem. 270(50):29998-30006, Akira et al., 1994, Cell 77(1):63-71, Zhong et al., 1994, Science 264:95-98, Zhong et al., 1994, Proc. Natl. Acad. Sci. U.S.A. 91:4806-4810. For Stat4, see, for example, Quelle et al., 1995, Mol. Cell. Biol. 15:3336-3343; and Hou et al., 1994, Science 265:1701-1706. For Stat6, see, for example, Yamamoto et al., 1994, Mol. Cell. Biol. 17:4342-4349; Zhang et al., 1994, 91:4806-4810; and Yamamoto et al., 1997, Cytogenet. Cell. 77:207-210. For SOCS1 and 3 sequences, see, for example, Starr et al., 1997, Nature, 387:917-921; Minamoto et al., 1997, Biochem., Biophys. Res. Commun. 237:79-83; Masuhara et al., 1997, Biochem, Biophys. Res. Commun. 239:439-446; Naka et

al., 1997, Nature <u>387</u>:924-929; and Endo et al., 1997, Nature <u>387</u>:921-924. Representative examples of human Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and SOCS3 sequences are shown in FIGS. 1-7 (SEQ ID NOs:1-14), respectively.

Further, additional forms, e.g., alleles or species homologs of such sequences can routinely be obtained and detected using the sequences described above in conjunction with standard cloning and hybridization techniques such as those find in Sambrook et al., eds., Molecular Cloning: A Laboratory Manual, 2nd. ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

The methods for monitoring transplant acceptance or monitoring an autoimmune disorder can be performed with kits designed for carrying out such methods. As such, the present invention also relates to kits for monitoring transplant acceptance and autoimmune disorders.

Such kits can be utilized for determining the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 mRNA present within a sample (e.g., a transplant sample or a sample obtained from an autoimmune tissue or a tissue suspected of being effected by an autoimmune disorder). Alternatively, such kits can be utilized for determining the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 polypeptide present within a sample (e.g., a transplant sample or a sample obtained from an autoimmune tissue or a tissue suspected of being effected by an autoimmune disorder). A kit can be capable of being used to determine the amount of any one, two, three, four, five, six, or seven of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 sequences (mRNA or polypeptide).

The kit, for example, can comprise a microarray for determining such amounts, wherein the microarray comprises one or more nucleic acid sequences immobilized onto a solid surface, said nucleic acid sequence or sequences exhibiting complementarity to at least one of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 mRNA. The kit can, in addition, comprise a labeled compound or agent capable of detecting the of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 polypeptide or mRNA encoding the polypeptide in a biological sample and means for determining the amount of the polypeptide or mRNA in the sample (e.g., an antibody which binds the polypeptide or an oligonucleotide probe which binds to DNA or mRNA encoding the polypeptide). Kits can also include instructions for use in determining the amount or amounts of mRNA or polypeptide, and can also include directions for monitoring and diagnosis.

For antibody-based kits, the kit can comprise, for example: (1) a first antibody (e.g., attached to a solid support) which binds to a of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 polypeptide; and, optionally, (2) a second, different antibody which binds to either the polypeptide or the first antibody and is conjugated to a detectable agent.

For oligonucleotide-based kits, the kit can comprise, for example: (1) an oligonucleotide, e.g., a detectably labeled oligonucleotide, which hybridizes to a of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 nucleic acid sequence; or (2) a pair of primers useful for amplifying a of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 nucleic acid molecule. The kit can also comprise, e.g., a buffering agent, a preservative, or a protein stabilizing agent. The kit can also comprise components necessary for detecting the detectable agent (e.g., an enzyme or a substrate).

For microarray-based kits, such kits can comprise a nucleotide sequence, e.g., an oligonucleotide sequence, immobilized onto the surface of a solid support (e.g., a glass or porous solid support).

The kits can also contain a control sample or a series of control samples (postive control, negative control, or both) which can be assayed and compared to the test sample contained.

Each component of the kit is usually enclosed within an individual container and all of the various containers are within a single package along with instructions for observing and monitoring transplant acceptance or monitoring an autoimmune disorder.

NUCLEIC ACID DETECTION

Preferred agents for detecting an mRNA of interest (that is Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 mRNA) are primers or labeled nucleic acid probes capable of hybridizing to the mRNA under stringent hybridization conditions. Nucleic acid probes can be, for example, full-length sequences, such as the nucleic acid sequences depicted in FIGS. 1-7 (SEQ ID NOs:1, 3, 5, 7, 9, 11 and 13), or complements thereof, or portions of such sequences (or complements thereof), such as oligonucleotides of at least about 12, 15, 25 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to the mRNA.

In certain embodiments, determination of the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 mRNA involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g., U.S. Patent Nos. 4,683,195 and 4,683,202), such as, for example, anchor PCR, RACE PCR or RT-PCR. Such methods can include the steps of collecting a cell sample, isolating mRNA from the cells of the sample, reverse transcribing the mRNA, contacting the sample with one or more primers which specifically hybridize to the selected sequence under conditions such that hybridization and amplification of the sequence (if present) occurs, and determining the amount of product that is present.

Alternative amplification methods can also routinely be utilized. Such methods can include, for example, self sustained sequence replication (Guatelli et al., 1990, Proc. Natl.

Acad. Sci. USA <u>87</u>:1874-1878), transcriptional amplification system (Kwoh et al., 1989, Proc. Natl. Acad. Sci. USA <u>86</u>:1173-1177), Q-Beta Replicase (Lizardi et al., 1988, Bio/Technology <u>6</u>:1197), or any other nucleic acid amplification method, followed by the detection/quantitation of the amplified molecules using techniques well known to those of skill in the art. These schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% (preferably at least 75%, more preferably at least 85%, most preferably at least 10 95%) identical to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, non-limiting example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50-65°C (preferably 65°C).

Probes can comprise any readily detectable label moiety. For example, probes utilized herein comprise a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor as a label moiety.

In alternate embodiments, the Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or 20 SOCS3 mRNA sequences can be detected "in situ" directly upon the sample, e.g., the biopsy sample. Techniques for such procedures are well known to those of skill in the art. See, e.g., Nuovo, G.J., 1992, "PCR In Situ Hybridization: Protocols and Applications," Raven Press, NY.

In other embodiments, the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 mRNA can be determined by hybridizing nucleic acid arrays, e.g., microarrays. In a specific embodiment of the invention, the expression of one or more of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and SOCS3 is measured or detected using a DNA microarray. A DNA microarray or chip is a microscopic array of DNA fragments or synthetic oligonucleotides, disposed in a defined pattern on a solid support, wherein they are amenable to analysis by standard hybridization methods (see, e.g., Schena, 1996, BioEssays 18: 427).

Microarrays share certain preferred characteristics: The arrays are reproducible, allowing multiple copies of a given array to be produced and easily compared with each other. Preferably the microarrays are small, usually smaller than 5 cm², and they are made from materials that are stable under binding (e.g., nucleic acid hybridization) conditions.

35 Microarrays contain a surface to which sequences corresponding to gene products (e.g., mRNA, cDNA, cRNA, or complements thereof), can be specifically hybridized or bound at

a known position. For practicing the methods of the present invention, the binding sites of the microarray are polynucleotides, preferably DNA polynucleotides, that specifically hybridize to at least a portion of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3 mRNA or cDNA, or any combination of such mRNA or cDNA molecules, produced by a subject mammal. That is, a given binding site or unique set of binding sites in the microarray will specifically bind the product (e.g., mRNA or cDNA) of a single gene, e.g., Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3.

Preferably, the nucleotide sequence of each of the different polynucleotide bound to the surface is in the range of about 15 to about 100 nucleotides in length. Polynucleotides can be synthesized using conventional methods, such as phosphoramidite-based synthesis methods. Alternatively, the binding site polynucleotide sequences can be derived from cDNA or genomic clones.

DNA microarrays can be probed using mRNA, extracted and, optionally, reversetranscribed and amplified from a sample (e.g., a transplant, autoimmune or control sample). 15 Nucleic acid hybridization and wash conditions are optimally chosen so that the probe "specifically binds" or "specifically hybridizes" to a specific array site, i.e., the probe hybridizes, duplexes or binds to a sequence array site with a complementary nucleic acid sequence but does not hybridize to a site with a non-complementary nucleic acid sequence. As used herein, one polynucleotide sequence is considered complementary to another when, 20 if the shorter of the polynucleotides is less than or equal to 25 bases, there are no mismatches using standard base-pairing rules or, if the shorter of the polynucleotides is longer than 25 bases, there is no more than a 5% mismatch. Preferably, the polynucleotides are perfectly complementary (no mismatches). It can easily be demonstrated that specific hybridization conditions result in specific hybridization by carrying out a hybridization 25 assay including negative controls (see, e.g., Shalon et al., 1996, Genome Research 6:639-645, and Chee et al., 1996, Science 274:610-614) or positive controls. Thus, in a preferred embodiment, a microarray of the invention further comprises a binding site designed to act as a negative control and/or a binding site designed to act as a positive control. For example, a positive control can relate to a constitutively expressed gene sequence, e.g., a 30 ubiquitin sequence, HSC70, or GADPH. A negative control can relate to a gene sequence not expressed in the test cell or tissue being assayed.

Exemplary, non-limiting examples of hybridization conditions that can be utilized with DNA microarrays are as follows: hybridization in 5 X SSC plus 0.2% SDS at 65° C for 4 hours followed by washes at 25° C in low stringency wash buffer (1 X SSC plus 0.2% SDS) followed by 10 minutes at 25° C in high stringency wash buffer (0.1 X SSC plus 0.2% SDS) (Shena et al., 1996, Proc. Natl. Acad. Sci. USA, 93:10614-19).

The use of a two-color fluorescence labeling and detection scheme to define alterations in gene expression has been described, e.g., in Shena et al., 1995, Science 270:467-470. An advantage of using mRNA, cRNA, or cDNA labeled with two different fluorophores is that a direct and internally controlled comparison of the mRNA levels corresponding to each arrayed gene in two cell states (e.g., control and transplant) can be made, and variations due to minor differences in experimental conditions (e.g., hybridization conditions) will not affect subsequent analyses. However, it will be recognized that it is also possible to use cDNA from a single cell, and compare, for example, the absolute amount of a particular mRNA in, e.g., a transplant or autoimmune 10 sample cell.

To facilitate detection the mRNA or cDNA are typically labeled with fluorescent dyes that emit at different wavelengths. Examples of fluorescent dyes include, but are not limited to, rhodamine, fluorescein, isothiocyanate, rhodamine, phycocythrin, phycocyanin, allophycocyanin, o-phthaldehyde and fluorescamine. The fluorescence emissions at each 15 site of a DNA array can be, preferably, detected by scanning confocal laser microscopy. In one embodiment, a separate scan, using the appropriate excitation line, is carried out for each of two fluorophores used. Alternatively, a laser can be used that allows simultaneous specimen illumination at wavelengths specific to the two fluorophores and emissions from the two fluorophores can be analyzed simultaneously (see, e.g., Shalon et al., 1996, Genome 20 Research 6:639-645).

Signals are recorded and, in a preferred embodiment, analyzed by computer, e.g., using a 12 bit analog to digital board. In one embodiment the scanned image is despeckled using a graphics program (e.g., Hijaak Graphics Suite) and then analyzed using an image gridding program that creates a spreadsheet of the average hybridization at each wavelength 25 at each site.

It will be appreciated that when mRNA or cRNA is hybridized to a microarray under suitable hybridization conditions, the level of hybridization to the site in the array corresponding to any particular gene will reflect the prevalence in the cell of mRNA transcribed from that gene. For example, when detectably labeled (e.g., with a fluorophore) 30 cRNA complementary to the total cellular mRNA is hybridized to a microarray, the site on the array corresponding to a gene (i.e., capable of specifically binding the product of the gene) that is not transcribed in the cell will have little or no signal (e.g., fluorescent signal), and a gene for which the encoded mRNA is prevalent will have a relatively strong signal.

Microarrays can be made in a number of ways well known to those of skill in the art. 35 With respect to the nucleic acids of the binding sites, the nucleic acid for the microarray can be generated by synthesis of synthetic polynucleotides or oligonucleotides, e.g., using N-

phosphonate or phosphoramidite chemistries (e.g., Froehler et al., 1986, Nucleic Acid Res 14:5399-5407). In some embodiments, synthetic nucleic acids include non-natural bases, e.g., inosine. Additionally, it is possible to vary the charge on the phosphate backbone of the oligonucleotide, for example, by thiolation or methylation, or even to use a peptide rather than a phosphate backbone. The making of such modifications is within the skill of one trained in the art. Further, nucleic acid analogues may be used as binding sites for hybridization. An example of a suitable nucleic acid analogue is peptide nucleic acid (see, e.g., Egholm et al., 1993, Nature 365:566-568; see also U.S. Patent No. 5,539,083, Cook et al., entitled "Peptide nucleic acid combinatorial libraries and improved methods of synthesis," issued July 23, 1996). In addition, binding (hybridization) sites can also be made from plasmid or phage clones of genes, cDNAs (e.g., expressed sequence tags), or inserts therefrom (Nguyen et al., 1995, Genomics 29:207-209). In yet another embodiment, the polynucleotide of the binding sites is RNA.

The nucleic acid or analogue is attached to a solid support to produce the binding site. Solid supports may be made from glass, silicon, plastic (e.g., polypropylene, nylon, polyester), polyacrylamide, nitrocellulose, cellulose acetate or other materials. In general, non-porous supports, and glass in particular, are preferred. The solid support may also be treated in such a way as to enhance binding of oligonucleotides thereto, or to reduce non-specific binding of unwanted substances thereto. Preferably, the glass support is treated with polylysine or silane to facilitate attachment of oligonucleotides to the slide.

Methods of immobilizing DNA on the solid support may include direct touch, micropipetting (Yershov et al., Proc. Natl. Acad. Sci. USA, 1996, 93:4913-4918), or the use of controlled electric fields to direct a given oligonucleotide to a specific spot in the array (U.S. Patent No. 5,605,662). In principal, any type of array, for example, dot blots on a 191 nylon hybridization membrane (see Sambrook et al., 1989, Molecular Cloning - A Laboratory Manual (2nd Ed.), Vols. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York), can used, although, as will be recognized by those of skill in the art, very small arrays are be preferred because hybridization volumes will be smaller. DNA can typically be immobilized at a density of 50, 75, 100, up to 10,000 oligonucleotides per cm² and preferably at a density of about 1000 oligonucleotides per cm².

In addition, nucleic acids can be attached to a surface by printing on glass plates (Schena et al., 1995, Science 270:467-470; DeRisi et al., 1996, Nature Genetics 14:457-460; Shalon et al., 1996, Genome Res. 6:639-645; and Schena et al., Proc. Natl. Acad. Sci. USA, 1996, 93(20):10614-19.) As an alternative to immobilizing pre-fabricated oligonucleotides onto a solid support, it is possible to synthesize oligonucleotides directly on the support (Maskos et al., 1993, Nucl. Acids Res. 21: 2269-70; Fodor et al., 1991,

Science 251: 767-73; Lipshutz et al., 1999, Nat. Genet. 21(1 Suppl):20-4; McGall et al., Proc. Natl. Acad. Sci. USA 93: 13555-60, 1996). Other methods for making microarrays, e.g., by masking (Maskos and Southern, 1992, Nuc. Acids Res. 20:1679-1684), may also be used.

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PROTEIN DETECTION

Standard techniques can also be utilized for determining the amount of the protein or proteins of interest (that is, Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 protein) present in a sample. It is to be understood, that such a determination of the amount of a protein present includes determining the total amount of a protein present, and also includes, especially with respect to determining the amount of a Stat protein present, determining the amount of a phosphorylated form of the protein present.

For example, standard techniques can be employed using, e.g., immunoassays such as, for example, Western blot, immunoprecipitation followed by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), immunocytochemistry, and the like to determine the amount of the protein or proteins of interest present in a sample. A preferred agent for detecting a protein of interest is an antibody capable of binding to a protein of interest, preferably an antibody with a detectable label.

With respect to determining the amount of a phosphorylated form of a protein of interest that is present in a sample, such a determination can also be performed using standard technques well known to those of skill in the art. For example, such a determination can include, first, immunoprecipitation with an antibody that is specific for a phosphorylated amino acid residue, e.g., an anti-phosphotyrosine antibody, such that all exhibiting such a phosphorylated residue in a sample will be immunoprecipitated. Second, the immunoprecipitated proteins can be contacted with a second antibody that is specific for the particular protein of interest, e.g., Stat1, Stat2, Stat3, Stat4, or Stat6. Alternatively, a phosphorylated protein of interest can be identified and quantitated using an antibody specific for the phosphorylated form of the particular protein itself, e.g, an antibody specific for phosphorylated Stat1 that does not recognize non-phosphorylated Stat1. Such antibodies exist, and are well known to those of skill in the art.

For such detection methods, protein from the sample to be analyzed can easily be isolated using techniques which are well known to those of skill in the art. Protein isolation methods can, for example, be such as those described in Harlow and Lane (Harlow, E. and Lane, D., 1988, "Antibodies: A Laboratory Manual", Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York).

Preferred methods for the detection of the protein or proteins of interest involve their detection via interaction with a protein-specific antibody. For example, antibodies directed a protein of interest can be utilized as described herein. Antibodies directed against Statl, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 protein are well known to those of skill in the art. For example, antibodies directed against Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be obtained from such companies as Zymed Laboratories, Inc. (South San Francisco, CA), Santa Cruz Biotechnology, Inc. (Santa Cruz, CA), and Research Diagnostics, Inc., (Flanders, NJ). Alternatively, such antibodies can be generated utilizing standard techniques well known to those of skill in the art. See, e.g., Section 5.3, below, for 10 a more detailed discussion of such antibody generation techniques. Briefly, such antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or F(ab')2) can, for example, be used.

For example, antibodies, or fragments of antibodies, specific for a protein of interest can be used to quantitatively or qualitatively detect the presence of the protein. This can be 15 accomplished, for example, by immunofluorescence techniques. Antibodies (or fragments thereof) can, additionally, be employed histologically, as in immunofluorescence or immunoelectron microscopy, for in situ detection of a protein of interest. In situ detection can be accomplished by removing a histological specimen (e.g., a biopsy specimen) from a patient, and applying thereto a labeled antibody thereto that is directed to a Stat1, Stat2, 20 Stat3, Stat4, Stat6, SOCS1 or SOCS3 protein. The antibody (or fragment) is preferably applied by overlaying the labeled antibody (or fragment) onto a biological sample. Through the use of such a procedure, it is possible to determine not only the presence of the protein of interest, but also its distribution, its presence in lymphocytes within the sample. A wide variety of well-known histological methods (such as staining procedures) can be utilized in 25 order to achieve such in situ detection.

Immunoassays for a protein of interest typically comprise incubating a biological sample, e.g., a biopsy or subject blood sample, of a detectably labeled antibody capable of identifying a protein of interest, and detecting the bound antibody by any of a number of techniques well-known in the art. As discussed in more detail, below, the term "labeled" 30 can refer to direct labeling of the antibody via, e.g., coupling (i.e., physically linking) a detectable substance to the antibody, and can also refer to indirect labeling of the antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody.

The biological sample can be brought in contact with and immobilized onto a solid 35 phase support or carrier such as nitrocellulose, or other solid support which is capable of immobilizing cells, cell particles or soluble proteins. The support can then be washed with

suitable buffers followed by treatment with the detectably labeled fingerprint gene-specific antibody. The solid phase support can then be washed with the buffer a second time to remove unbound antibody. The amount of bound label on solid support can then be detected by conventional means.

By "solid phase support or carrier" is intended any support capable of binding an antigen or an antibody. Well-known supports or carriers include glass, polystyrene, polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, gabbros, and magnetite. The nature of the carrier can be either soluble to some extent or insoluble for the purposes of the present invention. The support material can 10 have virtually any possible structural configuration so long as the coupled molecule is capable of binding to an antigen or antibody. Thus, the support configuration can be spherical, as in a bead, or cylindrical, as in the inside surface of a test tube, or the external surface of a rod. Alternatively, the surface can be flat such as a sheet, test strip, etc. Preferred supports include polystyrene beads. Those skilled in the art will know many other 15 suitable carriers for binding antibody or antigen, or will be able to ascertain the same by use of routine experimentation.

One of the ways in which a Stat1-, Stat2-, Stat3-, Stat4-, Stat6-, SOCS1- or SOCS3specific antibody can be detectably labeled is by linking the same to an enzyme and use in an enzyme immunoassay (EIA) (Voller, A., "The Enzyme Linked Immunosorbent Assay 20 (ELISA)", 1978, Diagnostic Horizons 2:1-7, Microbiological Associates Quarterly Publication, Walkersville, MD); Voller, A. et al., 1978, J. Clin. Pathol. 31:507-520; Butler, J.E., 1981, Meth. Enzymol. 73:482-523; Maggio, E. (ed.), 1980, ENZYME IMMUNOASSAY, CRC Press, Boca Raton, FL; Ishikawa, E. et al., (eds.), 1981, ENZYME IMMUNOASSAY, Kgaku Shoin, Tokyo). The enzyme which is bound to the antibody will 25 react with an appropriate substrate, preferably a chromogenic substrate, in such a manner as to produce a chemical moiety which can be detected, for example, by spectrophotometric, fluorimetric or by visual means. Enzymes which can be used to detectably label the antibody include, but are not limited to, malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate, 30 dehydrogenase, triose phosphate isomerase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6phosphate dehydrogenase, glucoamylase and acetylcholinesterase. The detection can be accomplished by colorimetric methods which employ a chromogenic substrate for the enzyme. Detection can also be accomplished by visual comparison of the extent of 35 enzymatic reaction of a substrate in comparison with similarly prepared standards.

Detection can also be accomplished using any of a variety of other immunoassays. For example, by radioactively labeling the antibodies or antibody fragments, it is possible to detect a protein of interest through the use of a radioimmunoassay (RIA) (see, for example, Weintraub, B., Principles of Radioimmunoassays, Seventh Training Course on Radioligand Assay Techniques, The Endocrine Society, March, 1986, which is incorporated by reference herein). The radioactive isotope (*e.g.*, ¹²⁵I, ¹³¹I, ³⁵S or ³H) can be detected by such means as the use of a gamma counter or a scintillation counter or by autoradiography.

It is also possible to label the antibody with a fluorescent compound. When the fluorescently labeled antibody is exposed to light of the proper wavelength, its presence can then be detected due to fluorescence. Among the most commonly used fluorescent labeling compounds are fluorescein isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthaldehyde and fluorescamine.

The antibody can also be detectably labeled using fluorescence emitting metals such as ¹⁵²Eu, or others of the lanthanide series. These metals can be attached to the antibody using such metal chelating groups as diethylenetriaminepentacetic acid (DTPA) or ethylenediaminetetraacetic acid (EDTA).

The antibody also can be detectably labeled by coupling it to a chemiluminescent compound. The presence of the chemiluminescent-tagged antibody is then determined by detecting the presence of luminescence that arises during the course of a chemical reaction.

Examples of particularly useful chemiluminescent labeling compounds are luminol, isoluminol, theromatic acridinium ester, imidazole, acridinium salt and oxalate ester.

Likewise, a bioluminescent compound can be used to label the antibody of the present invention. Bioluminescence is a type of chemiluminescence found in biological systems in, which a catalytic protein increases the efficiency of the chemiluminescent reaction. The presence of a bioluminescent protein is determined by detecting the presence of luminescence. Important bioluminescent compounds for purposes of labeling are luciferin, luciferase and aequorin.

5.2. Methods and Compositions for Identifying Compounds That Reduce Immune Rejection

As demonstrated below, immune rejection can be reduced and tolerance can be induced by modulating the amount of particular members of the Jak/Stat pathway present, expressed or active within an affected tissue. Specifically, the results presented herein demonstrate that modulation of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 levels can reduce immune rejection.

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The methods described herein identify compounds that modulate the expression and/or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 sequences in a manner that can reduce immune rejection (either, for example, in a transplant situation or in an autoimmune situation). The compounds identified via such methods are, therefore, useful as lead compounds in the development of the rapeutic compositions for the reduction of immune rejection. Such methods are particularly useful in that the effort and great expense involved in testing potential therapeutics in vivo is efficiently focused on those compounds identified via the in vitro and ex vivo methods described herein.

Thus, the present invention relates to a method for identifying a compound to be 10 tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, 15 present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In alternate embodiments, 20 such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and comparing the amounts to those present in the control sample.

In certain embodiments, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the 25 amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount of Stat4 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control activated T cell sample that has 35 not been contacted with the test compound, so that if the amount of Stat4 is decreased or the

amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c) comparing the ratio in (a) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the ratio in (a) is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is 10 identified.

In another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, 15 (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding resting T cell sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the 20 amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and comparing the amounts to 25 those present in the control sample.

Optionally, such methods can further include comparing the amount or amounts in (a) to a control resting T cell sample that has not been contacted with the T cell activator or with the test compound and/or with a control resting T cell sample that has been contacted with the test compound, but has not been contacted with a T cell activator (and, therefore, 30 remains in the resting state). Such controls provide evidence regarding the specificity and toxicity of the test compound.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of 35 Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined

can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample. Further, in certain embodiments, the resting T cell is a primary T cell, and in other embodiments, the resting T cell is a T cell line.

In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of Stat4 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control resting T cell 10 sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of a method for identifying a compound to be 15 tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c) comparing the ratio in (a) to that present in a corresponding control resting T cell sample that has been contacted with a T cell activator, but has not been contacted with the test 20 compound, so that if the ratio in the sample is decreased relative to that in the control samples, a compound to be tested for an ability to reduce immune rejection is identified.

In another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is 25 responsive to the cytokine; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control T cell 30 sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

Cytokines that can be used as part of these methods include, but are not limited to, 35 IL-2, IL-4, IL-12, or IL-13.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to State, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell 10 sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine; (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased on the amount of State is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the 20 cytokine; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and (c) comparing the ratio to in (a) to that present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce 25 immune rejection is identified.

Standard methods and compositions for determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, and SOCS3 mRNA or protein can be utilized. Such methods and compositions are described in detail, above, in Section 5.1.

In addition to the ability to modulate Stat and/or SOCS levels as described herein, it may be desirable, at least in certain instances, that compounds that reduce immune rejection also modulate the expression or activity of such molecules as IL-4, interferon-γ (IFN-γ), IL-12, or IL-13. Thus, the methods described herein for identifying compounds to be tested for an ability to reduce immune rejection can further comprise determining the level of IL-35 4, IFN-γ or IL-13 in the T cell sample that has been contacted with the test compound, and comparing this level with that of the control T cell sample that has not been contacted with

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the test compound. Preferred compounds are ones wherein: the level of IL-12 or IFN-y in the test sample is decreased relative to the corresponding level in the control sample, or wherein the level of IL-4 or IL-13 in the test sample is equal to or greater than the corresponding level in the control sample.

The present methods of identifying compounds that to be tested for an ability to reduce immune rejection, can comprise methods for identifying compounds that modulate the activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3. Thus, such methods can comprise: (a) contacting a T cell sample with a test compound; (b) determining the activity of at least one of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3; and (c) comparing the 10 activity level or levels to that/those in a corresponding control T cell sample that has not been contacted with the test compound, so that if the level of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 activity in (a) is decreased, or the level of Stat6 activity in (a) is increased, relative to the level of activity in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In one preferred embodiment, the activity of Stat4 and Stat6 is determined. Such a preferred embodiment can further include determining the ratio of Stat4 activity to Stat6 activity so that if the ratio in the test sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

The T cell sample utilized herein can be an activated T cell sample, a resting T cell 20 sample, or a cytokine-responsive T cell sample, as discussed above. In instances wherein the T cell sample is a resting T cell sample, the T cell sample is contacted with a T cell activator and the test compound. In instances wherein the T cell sample is a cytokineresponsive T cell sample, the T cell sample is contacted with the cytokine and the test compound.

Standard techniques can be utilized to determine the level of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 activity. For example, the activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be determined by detecting the binding of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 to its cognate DNA binding element, via, for example, an electromobility shift assay ("EMSA"), detecting the expression of a gene 30 whose expression is controlled by a promoter that is responsive to Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3, detecting the induction of a reporter gene that comprises a regulatory element that is responsive to Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3, wherein the element is operably linked to a nucleic acid encoding a detectable marker, e.g., luciferase.

Genes whose expression is controlled by a Stat 1-responsive promoter are well known, and include, for example GBP-1, inducible NO synthase (iNOS), ICAM, IRF-1,

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major histocompatibility complex (MHC) class II transactivator (CIITA). See, e.g., Lew et al., 1991, Mol. Cell. Biol. 11:182-191, Gao et al., 1997, J. Biol. Chem. 272:1226-1230, Caldenhoven et al., 1994, J. Biol. Chem. 269:21146-21154, Sims et al., 1993, Mol. Cell. Biol. 13:690-702, Pine et al., 1994, EMBO J. 13:158-167, Harada et al., 1994, Mol. Cell.

Biol. <u>14</u>:1500-1509, and Piskurich et al., 1999, Mol. Cell. Biol. <u>19</u>(1):431-40. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 1-responsive promoters are well known (see, *e.g.*, Sims et al., 1993, Mol. Cell. Biol. <u>13</u>:690-702, Pine et al., 1994, EMBO J. <u>13</u>:158-167, and Piskurich et al., 1999, Mol. Cell.

Biol. 19(1):431-40), making the construction and assay of Stat 1-reporter genes routine. Genes whose expression is controlled by a Stat 2-responsive promoter are well known, and include, for example IRF-1. See, e.g., Li et al., 1996, J. Biol. Chem. 271(10):5790-5794. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques.

Likewise, the structure of Stat 2-responsive promoters are well known (see, e.g., Ghislain et al., 1996, J. Biol. Chem. 271(21):12408-12413 and Li et al., 1996, J. Biol. Chem. 271(10):5790-5794), making the construction and assay of Stat 2-reporter genes routine.

Genes whose expression is controlled by a Stat 3-responsive promoter are well known, and include, for example alpha-2-macroglobulin, fibrinogen, junB, haptoglobin, 20 matrix metalloproteinase (MMP-1), TIMP-1, and p21^{WAF/CIP1}. See, e.g., Wegenka et al., 1993, Mol. Cell. Biol. 13:276-288, Fujitani et al., 1994, Biochem. Bioph. Res. Co. 202:1181-1187, Coffer et al., 1995, Oncogene 10:985-994, Akira et al., 1994, Cell 77:63-71, and Chin et al., 1996, Science 272:719-722. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 3-responsive promoters are well known (see, e.g., Wegenka et al., 1993, Mol. Cell. Biol. 13:276-288 and Chin et al., 1996, Science 272:719-722), making the construction and assay of Stat 3-reporter genes routine.

Genes whose expression is controlled by a Stat 4-responsive promoter are well known, and include, for example interferon-γ and IL-12. See, e.g., Grigorieva et al., 2000, 30 J Biol. Chem. 275(10):7343-7350 and Naeger, L.K. et al., 1999, J. Biol. Chem. 274:1875-1878). Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 4-responsive promoters are well known (see, e.g., Grigorieva et al., 2000, J Biol. Chem. 275(10):7343-7350 and Naeger, L.K. et al., 1999, J. Biol. Chem. 274:1875-

35 1878), making the construction and assay of Stat 4-reporter genes routine.

Genes whose expression is controlled by a Stat 6-responsive promoter are well known, and include, for example IL-4, CD23, IL-4 receptor, MHC class II. See, e.g., Tinnell et al., 1998, Int. Immunol. 10(10):1529-38, Linehan et al., 1998, J. Immunol. 161(1):302-10, and Kotanides et al., 1996, J. Biol. Chem. 271(41):25555-25561. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 6-responsive promoters are well known (see, e.g., Curiel, R.E. et al., 1997, Eur. J. Imm. 27:1982-1987, Linehan et al., 1998, J. Immunol. 161(1):302-10, and Kotanides et al., 1996, J. Biol. Chem. 271(41):25555-25561), making the construction and assay of Stat 6-reporter genes routine.

EMSAs can also routinely be utilized to assess Stat1, Stat2, Stat3, Stat4 or Stat6 activity. Such techniques are well known to those of skill in the art. See, e.g., Amici et al., 1995, Cancer Research 55: 14452-4457. Briefly, in a representative, non-limiting example, extracts of cells treated with a test compound are mixed with ³²P-Stat 4 (or Stat1, 2, 3, or 6) element or a control oligonucleotide and poly(dI-dC) (Pharmacia Biotech Inc.) in binding buffer (e.g., Tris-Cl, pH 7.8, 50 mM NaCl, 1 mM EDTA, 0.5 mM dithiothreitol, 5% glycerol). After an approximately 20 minute incubation at room temperature, Stat4 (or Stat1, 2, 3, or 6)-DNA-complexes or control oligonucleotide complexes are analyzed by nondenaturing 4% polyacrylamide gel electrophoresis and autoradiography. The amount of shifted Stat4 (or Stat1, 2, 3, or 6) probe, an indicator Stat4 (or Stat1, 2, 3, or 6) activity, respectively, can be quantitated by Molecular Dynamics PhosphoImager (MDP) analysis.

The activity of SOCS1 or SOCS3 activity can be determined by, e.g., detecting the expression of a gene whose expression is controlled by SOCS1 or SOCS3. For example, SOCS1 expression inhibits IL-6, LIF, oncostatin M, IFN-γ, IFN-β, IFN-α, thrombopoeitin, and growth hormone (GH) induced Jak/Stat signaling. SOCS3 expression inhibits IFN-γ, IFN-β, IFN-α, GH and leptin. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques.

The activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can also be
assessed by detecting the proliferation of the T cell sample, detecting the effector function
of the sample or detecting differentiation of the sample. Techniques known to those of skill
in the art can be used for measuring these activities. For example, cellular proliferation can
be assayed by ³H-thymidine incorporation assays and trypan blue cell counts. The effector
function of T-cells can be measured, for example, by a ⁵¹Cr-release assay (see, *e.g.*,
Palladino et al., 1987, Cancer Res. <u>47</u>:5074--5079 and Blachere et al., 1993, J.
Immunotherapy <u>14</u>:352-356).

As set forth above, the methods described herein for identifying compounds to be tested for an ability to reduce immune rejection assay whether a test compound has an effect on the expression and/or activity of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, and/or SOCS3 mRNA or protein produced by a T cell, in particular, an activated T cell, or at a minimum, a T cell that has the ability to respond to exogenous cytokines.

The T cell used as part of the methods can be one that is constitutively activated (e.g., a constitutively activated T cell line), one that has or has gained the ability to respond to cytokines, one that is activated prior to performing the method, or one that is activated concurrently with the method. A T cell to be used as part of the methods described herein can be activated either prior to or simultaneously with contacting the cell with a test compound. With respect to activated, including constitutively activated T cells, activation of such T cells can, in certain instances be further enhanced by addition and contact with a T cell activator.

An activated T cell is one that expresses antigens indicative of T-cell activation (that is, T cell activation markers). Examples of T cell activation markers include, but are not limited to, CD25, CD26, CD30, CD38, CD69, CD70, CD71, ICOS, OX-40 and 4-1BB. The expression of activation markers can be measured by techniques known to those of skill in the art, including, for example, western blot analysis, northern blot analysis, RT-PCR, immunofluorescence assays, and fluorescence activated cell sorter (FACS) analysis. The activated T cell used as part of the present methods can be an activated T cell line or can be a primary cell that has been activated:

Activated T cell lines are well known to those of skill in the art. Examples of activated T cell lines include TH1 cell lines such as AE7, PL17, and OF6, and TH2 cell lines such as D10 and CDC35.

T cell lines that do not express the T cell activation markers required to constitute activation, but nonetheless have the ability to respond to cytokines are also well known to those of skill in the art. Examples of such T cell lines include CTLL-2 and HT-2.

Alternatively, primary T cells can be isolated, the majority of which will be in a resting state, and activated using standard techniques. For example, immune cells can be collected or isolated from blood, or secondary lymphoid organs of the subject, such as but not limited to lymph nodes, tonsils, the spleen, Peyer's patch of the intestine, and bone marrow, by any of the methods known in the art. Immune cells obtained from such sources typically comprise predominantly recirculating lymphocytes and macrophages at various stages of differentiation and maturation. Optionally, standard techniques, such as morphological observation and immunochemical staining, can be used, if desired, to verify

the presence of the desired cells, that is, T cells. In a preferred aspect, the immune cells used in the methods of the invention described herein are human peripheral blood compositions lacking red blood cells, e.g., whole blood leukocytes (whole peripheral blood from which the red blood cells and serum have been substantially removed), which can be collected from a human subject by standard techniques, such as by use of a syringe to withdraw the blood, followed by subjecting the blood to Ficoll-Hypaque (Pharmacia) gradient centrifugation. Blood, anticoagulated with preservative-free heparin, usually yields 0.5 to 1 x 10⁶ lymphocytes/ml. Separated blood cells (e.g., leukocytes) may be frozen by standard techniques prior to use in the present methods. In a specific embodiment, the immune cells used are purified white blood cells comprising lymphocytes and macrophages.

In one embodiment wherein further purification of T cells is desired, antibodies against specific surface markers can be directly labeled by conjugation of a detectable compound to such antibodies to facilitate detection and separation of T cells. Alternatively, in another embodiment, the primary antibody is detected by detecting binding of a secondary antibody or reagent to the primary antibody. In a further embodiment, the secondary antibody is labeled. Examples of detectable compounds include, but are not limited to, biotin, photobiotin, fluorescein isothiocyanate (FITC), or phycoerythrin (PE), or other compounds known in the art. Cells retaining labeled antibodies are then separated from cells that do not bind such antibodies by techniques known in the art such as, but not limited to, various cell sorting methods (e.g., FACS), affinity chromatography, and panning.

In another embodiment wherein further purification of T cells is desired, T cells are sorted using a fluorescence activated cell sorter (FACS). Fluorescence activated cell sorting (FACS) is a well-known method for separating particles, including cells, based on the fluorescent properties of the particles (Kamarch, 1987, Methods Enzymol, 151:150-165). Laser excitation of fluorescent moieties in the individual particles results in a small electrical charge allowing electromagnetic separation of positive and negative particles from a mixture.

In another embodiment wherein further purification of T cells is desired, magnetic 30 beads can be used to separate T cells. T cells may be sorted using a magnetic activated cell sorting (MACS) technique, a method for separating particles based on their ability to bind magnetic beads (0.5-100 μ diameter; Dynal, Inc., Lake Success, NY) as undertaken according to the manufacturer's instructions. A variety of useful modifications can be performed on the magnetic microspheres, including covalent addition of antibody which specifically recognizes a cell-solid phase surface molecule or hapten. A magnetic field is then applied, to physically manipulate the selected beads. The beads are then mixed with

the immune cells to allow binding. Cells are then passed through a magnetic field to separate out cells having T cell surface markers.

The isolated resting T cells can then be activated by contacting with a T cell activator. Any T cell activator can be utilized for this purpose. For example, any compound or factor that is a T cell receptor stimulatory factor, that is, induces T cell receptor signalling can be used. Preferably, the compound or factor also induces co-stimulatory pathways. Representative, non-limiting examples of T cell activators include, but are not limited to, anti-CD3 antibodies (preferably monoclonal antibodies) either alone or in conjunction with anti-CD28 antibodies (preferably monoclonal antibodies), or mitogens such as, for example, 10 phorbol 12-myristate 13-acetate (PMA), phytohemagglutinin (PHA), or concanavalin-A (ConA).

Compounds that can be tested and identified methods described herein can include, but are not limited to, compounds obtained from any commercial source, including Aldrich (1001 West St. Paul Ave., Milwaukee, WI 53233), Sigma Chemical (P.O. Box 14508, St. 15 Louis, MO 63178), Fluka Chemie AG (Industriestrasse 25, CH-9471 Buchs, Switzerland (Fluka Chemical Corp. 980 South 2nd Street, Ronkonkoma, NY 11779)), Eastman Chemical Company, Fine Chemicals (P.O Box 431, Kingsport, TN 37662), Boehringer Mannheim GmbH (Sandhofer Strasse 116, D-68298 Mannheim), Takasago (4 Volvo Drive, Rockleigh, NJ 07647), SST Corporation (635 Brighton Road, Clifton, NJ 07012), Ferro 20 (111 West Irene Road, Zachary, LA 70791), Riedel-deHaen Aktiengesellschaft (P.O. Box D-30918, Seelze, Germany), PPG Industries Inc., Fine Chemicals (One PPG Place, 34th Floor, Pittsburgh, PA 15272). Further any kind of natural products may be screened using the methods of the invention, including microbial, fungal, plant or animal extracts.

Furthermore, diversity libraries of test compounds, including small molecule test 25 compounds, may be utilized. For example, libraries may be commercially obtained from Specs and BioSpecs B.V. (Rijswijk, The Netherlands), Chembridge Corporation (San Diego, CA), Contract Service Company (Dolgoprudny, Moscow Region, Russia), Comgenex USA Inc. (Princeton, NJ), Maybridge Chemicals Ltd. (Cornwall PL34 OHW, United Kingdom), and Asinex (Moscow, Russia).

Still further, combinatorial library methods known in the art, can be utilize, including, but not limited to: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the "one-bead one-compound" library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, 35 while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, 1997, Anticancer Drug Des. 12:145). combinatorial

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libraries of test compounds, including small molecule test compounds, can be utilized, and may, for example, be generated as disclosed in Eichler & Houghten, 1995, Mol. Med. Today 1:174-180; Dolle, 1997, Mol. Divers. 2:223-236; and Lam, 1997, Anticancer Drug Des. <u>12</u>:145-167.

Examples of methods for the synthesis of molecular libraries can be found in the art. for example in: DeWitt et al., 1993, Proc. Natl. Acad. Sci. USA 90:6909; Erb et al., 1994, Proc. Natl. Acad. Sci. USA 91:11422; Zuckermann et al., 1994. J. Med. Chem. 37:2678; Cho et al., 1993, Science 261:1303; Carrell et al., 1994, Angew. Chem. Int. Ed. Engl. 33:2059; Carell et al., 1994, Angew. Chem. Int. Ed. Engl. 33:2061; and Gallop et al., 1994. 10 J. Med. Chem. <u>37</u>:1233.

Libraries of compounds may be presented in solution (e.g., Houghten, 1992, Bio/Techniques 13:412-421), or on beads (Lam, 1991, Nature 354:82-84), chips (Fodor, 1993, Nature 364:555-556), bacteria (U.S. Patent No. 5,223,409), spores (Patent Nos. 5,571,698; 5,403,484; and 5,223,409), plasmids (Cull et al., 1992, Proc. Natl. Acad. Sci. 15 USA <u>89</u>:1865-1869) or phage (Scott and Smith, 1990, Science <u>249</u>:386-390; Devlin, 1990, Science 249:404-406; Cwirla et al., 1990, Proc. Natl. Acad. Sci. USA 87:6378-6382; and Felici, 1991, J. Mol. Biol. 222:301-310).

Among the test compounds that can be tested are compounds, including small organic molecule compounds that act as protein tyrosine kinase inhibitors, including, but 20 not limited to, the typhostin class of protein tyrosine kinase inhibitors (see, e.g., Gazit et al. 1989, J. Medicinal Chem. 32:2344-2352; and Levitski, 1992, FASEB J. 6:3275). Among such tyrphostin compounds that can be tested are A25 (available, e.g., from CalBiochem) or AG-490 (B42) (Meydan, N. et al., 1996, Nature 379:645-648) or derivatives thereof. Further, among the compounds that can be tested are compounds that interfere with SH2 25 domain interactions (see, e.g., U.S. Patent No. 5,710,129; 5,776,902; and 5,580,97, or derivatives of compounds therein), e.g., SH2-mediated Stat 4/IL-12 receptor-β, interactions. Still further, among the compounds that can be tested are compounds that interfere with Jak 2/IL-12 receptor-β₂ interactions and/or ones that interfere with Tyk 2/IL-12 receptor- β_1 interactions.

30 Upon identification of compounds to be tested for an ability to reduce immune rejection, the compounds can be further investigated. In particular, for example, the compounds identified via the present methods can be further tested in vivo in accepted animal models of transplant or autoimmune disorders. Further, the compounds identified can also be analyzed with respect to their specificity. In particular, the compounds can be 35 tested for an effect on platelet aggregation and/or on NF-κB activation. Techniques for such additional compound investigation are described below.

Accepted animal models can be utilized to determine whether the compounds identified via the methods described herein. Such models can include both transplantrelated models as well as autoimmune disorder models.

For example, the ability of a compound to reduce immune rejection, including the ability of the compound to induce tolerance in a subject mammal that has undergone a transplant can include, but are not limited to, a murine allograft model in which an allogeneic heart is transplanted into a subject mouse recipient (Hancock et al., 1998, Nature Medicine 4:1392-1396). In addition, primate models can also be tested. Such models include, for example, a primate renal allograft model (Kirk et al., 1997, Proc. Natl. Acad. 10 Sci. USA 94:8789-8794). In addition, a graft versus host disease (GVHD) model can be used (see, e.g., Guillen et al., 1986, Laboratory Investigation 55:35-42). In such models, chronic and acute GVHD is made to result from introduction of donor cells into a host exhibiting disparate MHC alleles. The GVHD results, therefore, from the donor cells' response to such the host's disparate MHC alleles.

The ability of a compound to reduce immune rejection can also be tested in such autoimmune disorder models as, first, an experimental allergic encephalomyelitis (EAE) model. EAE is an experimental autoimmune disease of the central nervous system (CNS) (Zamvil et al, 1990, Ann. Rev, Immunol. 8:579) and is a disease model for the human autoimmune condition, multiple sclerosis (MS). EAE is an example of a cell-mediated 20 autoimmune disorder that is mediated via T cells. No direct evidence exists for an autoantibody requirement in disease progression. EAE is readily induced in mammalian species by immunizations of myelin basic protein purified from the CNS or an encephalitogenic proteolipid (PLP). SJL/J mice are a susceptible strain of mice (H-2') and, upon induction of EAE, these mice develop an acute paralytic disease and an acute cellular 25 infiltrate is identifiable within the CNS.

In addition, a collagen-induced arthritis (CIA) model can be utilized to determine whether the compound of interest reduce immune rejection. CIA is an animal model for the human autoimmune disease rheumatoid arthritis (RA) (Trenthorn et al., 1977, J. Exp. Med., 146:857). This disease can be induced in many species by the administration of 30 heterologous type II collagen (Courtenay et al., 1980, Nature 283:665; Cathcart et at, 1986, Lab. Invest., 54:26). With respect to animal models of arthritis see, in addition, e.g., Holmdahl, R., 1999, Curr. Biol. 15:R528-530.

Still further, animal models for type 1 diabetes, thyroid autoimmunity or systemic lupus erythematosus, including glomerulonephritis can be utilized to determine whether the 35 compound of interest reduces immune rejection (see, e.g., Flanders et al., 1999,

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Autoimmunity 29:235-246; Krogh et al., 1999, Biochimie 81:511-515; and Foster, N.H., 1999, Semin. Nephrol. <u>19</u>:12-24, respectively).

In addition, it is preferred that compounds to be utilized as therapeutic according to the methods described herein not induce platelet aggregation. Therefore, it is preferable that compounds identified via the methods described herein that are to be tested for an ability to reduce immune rejection be further tested for an ability to induce platelet aggregation. In vitro and ex vivo assays for platelet aggregation are well known and compounds of interest can easily be tested via such assays.

Specifically, such assays include, but are not limited to the turbidometric method, in 10 which aggregation is measured as an increase in transmission of visible light through a stirred or agitated platelet suspension. See, e.g., Chanarin, L., 1989, Laboratory Haematology, Chapter 30, Churchill, Livingstone, London; and Schmidt, R.M. (ed), 1979, CRC Handbook Series in Clinical Laboratory Science, CRC Press, Inc.: Boca Raton, Florida.

Platelet aggregation can also be assayed via methods such as those described in U.S. Patent 5,976,532. For example, in a non-limiting example of such a method, the platelet concentration in platelet-rich plasma obtained (PRP) obtained from blood samples is adjusted to 200,000 to 300,000/mm³. In an in vitro assay, the PRP is aliquoted and incubated in the presence or absence of a compound of interest for a period of time (e.g., 15 20 minutes at 37° C) prior to the addition of a platelet inducing agonist (e.g., ADP, thrombin, collagen, epinephrine, and ristocetin). In an ex vivo assay, the PRP obtained from individuals treated with the compound of interest or a placebo is aliquoted and incubated in the presence of a platelet inducing agonist (e.g., ADP, thrombin, collagen, epinephrine, and ristocetin). Platelet aggregation is measured by assessing an increase in the transmission of 25 visible light through a platelet suspension using a spectrophotometer.

It is also preferred that compounds to be utilized as therapeutic according to the methods described herein not affect NF-κB activation, in particular, NF-κB activation in CD40L⁺ cells. Therefore, it is preferable that compounds identified via the methods described herein that are to be tested for an ability to reduce immune rejection be further 30 tested for possible effect on NF-κB activation in CD40L+ cells. In such tests, a CD40L+ cell is contacted with the compound of interest, and its effect on NF-kB activation, if any is assayed, and compared to the level of NF-kB activation in a corresponding control CD40L+ cell that has not been contacted with the compound.

Standard techniques can be utilized to test for NF-kB activation. For example, the 35 activity of NF-κB can be assessed by detecting the binding of NF-κB to its cognate DNA binding element in an electromobility shift assay (EMSA), detecting the expression of a

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gene whose expression is controlled by a promoter that is responsive to NF-kB, detecting the induction of the expression of a reporter gene construct that comprises a regulatory element that is responsive to NF-kB is operably linked to a nucleic acid encoding a detectable marker, e.g., luciferase.

Genes whose expression is controlled by an NF-kB-responsive promoter are well known, and include, for example granulocyte/macrophage colony-stimulating factor (GM-CSF), granulocyte colony-stimulating factor (G-CSF), IL-2, IL-6, IL-8, tumor necrosis factor- α (TNF- α), and intercellular cell adhesion molecule 1 (ICAM-1). See, e.g., Baeuerle and Henkel, 1994, Ann. Rev. Immunol. 12:141-179. Thus, expression of such genes in 10 CD40L⁺ cells in the presence and absence of a compound of interest can routinely be determined using standard techniques. Expression can be determined using standard techniques. Preferably, the compound being tested will not increase such expression and, most preferably, will have no effect on such expression. Likewise, the structure of NF-kBresponsive promoters are well known (see, e.g., Baeuerle and Henkel, 1994, Ann. Rev. 15 Immunol. 12:141-179; and Thanos et al., 1995, Cell 80:529-532), making the construction and assay of NF-kB reporter genes routine. Thus, the induction and expression of such reporter genes in CD40L+ cells in the presence and absence of a compound of interest can routinely be determined using standard techniques. Preferably, the compound being tested will not induce expression of the reporter gene.

EMSAs can also routinely be utilized to assess NF-kB activity. Such techniques are 20 well known to those of skill in the art. See, e.g., Amici et al., 1995, Cancer Research 55: 14452-4457. Briefly, in a representative, non-limiting example, extracts of cells treated with a test composition or control composition are mixed with ³²P-NF-κB element or a control oligonucleotide and poly(dI-dC) (Pharmacia Biotech Inc.) in binding buffer (e.g., 25 Tris-Cl, pH 7.8, 50 mM NaCl, 1 mM EDTA, 0.5 mM dithiothreitol, 5% glycerol). After an approximately 20 minute incubation at room temperature, NF-kB-DNA-complexes or control oligonucleotide complexes are analyzed by non-denaturing 4% polyacrylamide gel electrophoresis and autoradiography. The amount of shifted NF-kB probe, an indicator NFκB activity, respectively, can be quantitated by Molecular Dynamics PhosphoImager 30 (MDP) analysis.

Further, the effect, if any, of the compound of interest on NF-kB activation can also be tested by assaying for a cellular response, for example, cellular differentiation, or cell proliferation. Cellular proliferation can be assayed by, for example, ³H-thymidine incorporation assays and trypan blue cell counts. Preferably, the compound will have no 35 effect on such cellular responses.

5.3. Methods and Compositions for Reducing Immune Rejection

Described herein are methods and compositions for reducing immune rejection in a subject mammal. In particular, such methods comprise administration of compounds that serve to reduce the amount or activity of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat4 mRNA or protein, SOCS1 mRNA or protein or SOCS3 mRNA or protein, and/or increase the amount or activity of Stat6 mRNA or protein. The compounds utilized herein are ones that neither induce platelet aggregation nor affect NF-κB activation.

Immune rejection in any subject mammal that has undergone a transplant, or that exhibits or is suspected of exhibiting an autoimmune disorder can be reduced using the methods presented herein. Preferably, the mammal is a human, however, such subject mammals can also include, but are not limited to, other primates, including monkeys, as well as pigs, dogs, cats, horses, cattle, sheep, mice, rats, and rabbits.

The term "reducing immune rejection," is meant to encompass prevention or inhibition of immune rejection, as well as delaying the onset or the progression of immune rejection. The term is also meant to encompass prolonging survival of a transplant in a subject mammal, or reversing failure of a transplant in a subject. Further, the term is meant to encompass ameliorating a symptom of an immune rejection, including, for example, ameliorating an immunological complication associated with immune rejection, such as for example, interstitial fibrosis, chronic graft atherosclerosis, or vasculitis. The term is also meant to encompass induction of tolerance in a subject mammal that has undergone a transplant.

Specifically, the present invention relates to methods for reducing immune rejection in a subject mammal, said methods comprising: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to reduce the level or activity of Stat4 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-κB activation in CD40L⁺ cells.

The present invention also relates to methods for reducing immune rejection in a subject mammal, said methods comprising: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to reduce the level or activity of Stat1 mRNA or protein, Stat2 mRNA or protein, or Stat3 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-kB activation in CD40L⁺ cells.

Such methods can also include methods for reducing immune rejection in a subject mammal, comprising administering to the subject mammal in need of such a reduction a

concentration of a compound sufficient to decrease the level or activity of Stat4 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, and wherein the level or activity of Stat6 mRNA or protein in the subject is maintained or increased relative to that observed in the subject in the absence of the compound. Further, the compound administered is one that does not induce platelet aggregation or affect NF-kB activation in CD-40L⁺ cells.

Alternatively, such methods for reducing immune rejection in a subject mammal can comprise: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to increase the level or activity of Stat6 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-kB activation in CD40L⁺ cells.

Such methods for reducing immune rejection in a subject mammal can also comprise: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to decrease the level or activity of Stat4 mRNA or protein and maintain or increase the level or activity of Stat6 mRNA or protein in the subject subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-kB activation in CD40L⁺ cells.

Generally, practice of these methods does not solely entail administration of compositions that are considered signal 2-type blockers (see, e.g., Gummert J.F., et al., 1999, J. Am. Soc. Nephrol. 10: 1366), that is, compounds (e.g., CD40L antibodies) that act to inhibit CD40/CD40L (CD154) interactions or B7/CD28 interactions.

It is noted, however, that embodiments of the present invention further include combinatorial immune reduction therapy utilizing compositions as taught herein in conjunction with immunosuppressive or immunomodulatory drug therapies, as described in detail, below.

The methods of the present invention for reducing immune rejection can be utilized, e.g., for reducing immune rejection in a subject mammal that has undergone a transplant.

30 For example, such methods can induce tolerance in a subject mammal that has undergone a transplant. Such methods can be used to reduce immune reject in a transplant situation involving any cell, organ, organ system or tissue which can elicit an immune response in a recipient subject mammal. In general, therefore, a transplant includes an allograft, or a xenograft cell, organ, organ system or tissue. An allograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of the same species as the recipient. A xenograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of a

different species as the recipient. In particular, the transplant can, for example, be an allograft heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, intestine or cornea transplant.

The methods of the present invention for reducing immune rejection can also be utilized, e.g., for reducing immune rejection in a subject mammal exhibiting an autoimmune disorder. Thus, the present invention can treat an autoimmune disorder affecting any body cell, tissue, organ or organ system, including but not limited to cutaneous, cardiac, pericardial, endocardial, vascular lining or wall, blood, blood-forming (e.g., marrow or spleen), endocrine (e.g., pancreatic or thyroid), gastrointestinal (e.g., bowel), respiratory 10 (e.g., lung), renal, central nervous system, peripheral nervous system, muscular or skeletal joint (e.g., articular cartilage or synovial) tissue. The methods and compositions of the present invention can, therefore, be utilized to treat any autoimmune disorder including, but not limited to atopic dermatitis, contact dermatitis, eczematous dermatitides, seborrheic dermatitis, Lichen planus, Pemphilgus, bullous pemphigus, Epidermolysis bullosa, 15 Alopecia areata, urticaria, angioedemas, erythema, eosinophilias, migraine, lupus, including cutaneous lupus (discoid lupus erythematosus), extracutaneous lupus, including systemic lupus erythematosus, acute lupus, lupus annularis, lupus discretus, lupus lymphaticus, lupus papillomatis, lupus psoriasis, lupus vulgaris, lupus sclerosis, neonatal lupus erythematosus, and drug-induced lupus; anti-phospholipid syndrome (APS), hemolytic anemia (HA), 20 idiopathic thrombocytopenia (ITP), thyroiditis, diabetes mellitus (DM), inflammatory bowel disease, e.g., Crohn's disease or ulcerative cholitis, rhinitis, uveitis, nephrotic syndrome, demyelinating diseases such as multiple sclerosis (MS), myasthenia gravis (MG), and arthritis, e.g., rheumatoid arthritis, non-rheumatoid inflammatory arthritis, arthritis associated with Lyme disease, or osteoarthritis.

The compounds utilized as part of these methods include, but are not limited to, 25 ones identified via the methods described above. A number of different points along the Jak/Stat pathway can be targeted by the compounds utilized as part of the methods for reducing immune rejection described herein. Administration methods, including gene therapy methods, and pharmaceutical preparations by which such compounds can routinely 30 be utilized as part of methods for reducing immune rejection are taught below.

For example, compounds that specifically downregulate Stat4 mRNA or protein levels or activity, while not affecting NF-kB activation or platelet aggregation can be utilized as part of these methods. In addition, compounds that specifically downregulate Stat1 mRNA or protein levels or activity, while not affecting NF-kB activation or platelet 35 aggregation can be utilized as part of these methods. Also, compounds that specifically downregulate Stat2 mRNA or protein levels or activity, while not affecting NF-kB

activation or platelet aggregation can be utilized as part of these methods. In addition, compounds that specifically downregulate Stat3 mRNA or protein levels or activity, while not affecting NF-kB activation or platelet aggregation can be utilized as part of these methods. In addition, compounds that specifically downregulate SOCS1 mRNA or protein levels or activity, while not affecting NF-kB activation or platelet aggregation can be utilized as part of these methods. Likewise, compounds that specifically downregulate SOCS3 mRNA or protein levels or activity, while not affecting NF-kB activation or platelet aggregation can be utilized as part of these methods. In addition, compounds or methods that specifically increase Stat 6 mRNA or protein levels or activity, while not affecting NF-10 kB activation or platelet aggregation can be utilized as part of these methods. Representative, non-limiting examples of such compounds are described in detail below.

First, such compounds can include, for example, antisense, ribozyme, or triple helix compounds that can downregulate the expression or Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3. Such compounds are described in detail in the subsection below.

Second, such compounds can include, for example, antibody compositions that can downregulate the expression or activity of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3. Such compounds are described in detail in the subsection below.

Further, among such compounds are ones, including ones administered via gene therapy techniques, that serve to upregulate Stat6 expression or activity, and compounds 20 that act in maintaining Stat6 expression or activity levels as Stat4 activity or expression levels are decreased.

Compositions can include, for example ones can be utilized that compete with Stat4 for binding to the IL-12 receptor β_2 ("IL-12R β_2 "). Examples of such compounds include, but are note limited to limited to, peptide compositions as in Naeger, L.K. et al., 1999, J.

- 25 Biol. Chem. <u>274</u>:1875-1878. Additional examples of compounds that can be utilized include compounds, such as small organic compounds that act as inhibitors of SH2 domainmediated interactions such as SH2-mediated Stat4/IL-12Rβ₂ interactions. One example of such an embodiment involves a composition of the invention comprises one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 from binding to the IL-
- 30 $12R\beta_2$, or that comprise one or more dominant-negative Stat4 polypeptides (e.g., a Stat4 polypeptide lacking its SH2 domain or a Stat4 polypeptide lacking its DNA binding domain). Examples of such polypeptides include, but are not limited to, (using the standard one-letter amino acid code) phospho-YLPSNID peptides (Naeger, L.K. et al., 1999, J. Biol. Chem. 274:1875-1878).
- In specific embodiments, Stat1 antisense oligonucleotides, Stat2 antisense 35 oligonucleotides, Stat3 antisense oligonucleotides, or any combination thereof, are

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administered to reduce immune rejection. In other embodiments one or more anti-Stat1 antibodies, anti-Stat2 antibodies, or anti-Stat3 antibodies are administered to reduce immune rejection. In other embodiments, one or more peptides that compete with Jak1 or Tyk2 for binding to the IFN-α receptor or IFN-β receptor are administered to reduce immune rejection. In yet another embodiment, one or more peptides that compete with Jak1 or Jak2 for binding to the IFN-y receptor are administered to reduce immune rejection. In other embodiments, one or more peptides that compete with Stat1 or Stat2 for binding to the IFN-α receptor or IFN-β receptor are administered to reduce immune rejection. In other embodiments, one or more peptides that compete with Stat1 for binding to the IFN-y 10 receptor are administered to reduce immune rejection. In yet other embodiments, one or more dominant-negative Stat1 polypeptides (e.g., a Stat1 polypeptide lacking its SH2 domain or a Stat1 polypeptide lacking its DNA binding domain), dominant-negative Stat2 polypeptides (e.g., a Stat2 polypeptide lacking its SH2 domain or a Stat2 polypeptide lacking its DNA binding domain), or dominant-negative Stat3 polypeptides (e.g., a Stat3 15 polypeptide lacking its SH2 domain or a Stat3 polypeptide lacking its DNA binding domain) are administered to reduce immune rejection.

Jak2 is involved in activation of Stat4 protein. In view of this, another composition that can be utilized as part of the methods of the invention comprises a composition that reduces the expression or activity of Jak2, while not affecting NF-κB activation or platelet aggregation. In one embodiment, therefore, one or more peptides that compete with Jak2 for binding to the IL-12Rβ₂ can be utilized. In other embodiments, such compounds include Jak2 antisense molecules, triple helix molecules or ribozyme molecules that serve to downregulate the expression of Jak2. Representative antisense compositions are described in detail below. Such compounds also include antibodies or fragments thereof that specifically bind to and inhibit the activity of Jak2.

Tyk2 is also involved in activation of Stat4 protein. In view of this, another composition that can be utilized as part of the methods of the invention comprises a composition that reduces the expression or activity of Tyk2, while not affecting NF-κB activation or platelet aggregation. In another embodiment, a composition of the invention comprises one or more peptides that compete with Tyk2 for binding to the IL-12Rβ₁. In other embodiments, such compounds include Tyk2 antisense molecules, triple helix molecules or ribozyme molecules that serve to downregulate the expression of Tyk2. Representative antisense compositions are described in detail below.

In yet another embodiment, a composition that can be utilized as part of these
35 methods comprises one or more small molecules that decrease or downregulate Stat4
expression or activity, while not affecting NF-kB activation or platelet aggregation. For

example, among the compounds that can be utilized as part of these methods are protein tyrosine kinase inhibitors, including, but not limited to the tyrphostin class of protein tyrosine kinase inhibitors. Preferable tyrphostin compositions are ones that inhibit or downregulate Stat4 activity by (without wishing to be bound by any particular mechanism) inhibiting Jak2 or Tyk2 protein tyrosine kinase activity without deleterious effects on normal hematopoiesis. In specific embodiments, the tyrphostin is AG-490 (B42), although it is preferred that this particular tyrphostin not be utilized for treatment of autoimmune disorders, specifically multiple sclerosis (MS).

ANTISENSE, RIBOZYME, TRIPLE-HELIX COMPOSITIONS

Representative, non-limiting examples of Stat1 antisense molecules include the following: 5'- GCT GAA GCT CGA ACC ACT GTG ACA TCC - 3' (SEQ ID NO:19); and 5'-AAG TTC GTA CCA CTG AGA CAT CCT GCC (SEQ ID NO:20).

Representative, non-limiting examples of Stat2 antisense molecules include the following: 5'- CAT CTC CCA CTG CGC CAT TTG GAC TCT TCA -3' (SEQ ID NO:21); and 5'-CAG CAT TTC CCA CTG CGC CAT TTG GGC-3' (SEQ ID NO:22).

Representative, non-limiting examples of Stat3 antisense molecules include the following: 5'- CTG GTT CCA CTG AGC CAT CCT GCT GCA TCAG - 3' (SEQ ID NO:-23); and 5'-CTG TAG CTG ATT CCA TTG GGC CAT CCT-3' (SEQ ID NO:24).

Representative, non-limiting examples of Stat4 antisense molecules include the following: 5'- GAT TCC ACT GAG ACA TGC TGC TCT CTC TCT C-3' (SEQ ID NO:25); and 5'-GAC TTG ATT CCA CTG AGA CAT GCT AGC-3' (SEQ ID NO:26).

Representative, non-limiting examples of Jak2 antisense molecules include the following: 5'- GCC AGG CCA TTC CCA TCT AGA GCT TTT TTC -3' (SEQ ID NO:27); and 5'-CGT AAG GCA GGC CAT TCC CAT GCA GAG-3' (SEQ ID NO:28).

Representative, non-limiting examples of Tyk2 antisense molecules include the following: 5'- CCC ACA CAG AGG CAT GGT CCC CAC CAT TCA -3' (SEQ ID NO:29); and 5'-GGC CAT CCC CCA GTG GCG CAG AGG CAT GCT CCC-3' (SEQ ID NO:30).

Representative, non-limiting examples of SOCS1 antisense molecules include the following: 5'- CCT GGT TGC GTG CTA CCA TCC TAC TCG AGG GGC -3' (SEQ ID NO:31); and 5'-CAC CTG GTT GTG TGC TAC CAT CCT ACT-3' (SEQ ID NO:32).

Representative, non-limiting examples of SOCS3 antisense molecules include the following: 5'- GCT GTG GGT GAC CAT GGC GCA CGG AGC CAG CG -3' (SEQ ID NO:33); and 5'- GGC GGG AAA CTT GCT GTG GGT GAC CAT-3' (SEQ ID NO:34).

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In addition, standard techniques can be utilized to produce antisense, triple helix, or ribozyme molecules for use as part of the methods described herein.

First, standard techniques can be utilized for the production of antisense nucleic acid molecules, *i.e.*, molecules which are complementary to a sense nucleic acid encoding a polypeptide of interest (*e.g.*, Stat1, Stat2, Stat3, Stat4, Jak2, Tyk2, SOCS1, or SOCS3), *e.g.*, complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire coding strand, or to only a portion thereof, *e.g.*, all or part of the protein coding region (or open reading frame). An antisense nucleic acid molecule can be antisense to all or part of a non-coding region of the coding strand of a nucleotide sequence encoding a polypeptide of interest. The non-coding regions ("5' and 3' untranslated regions") are the 5' and 3' sequences which flank the coding region and are not translated into amino acids.

An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides or more in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxylmethyl) uracil,

- 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine,
 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine,
 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine,
 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil,
- 30 beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and
- 35 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense

orientation (i.e., RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest).

Antisense nucleic acid molecules administered to a subject or generated in situ such that they hybridize with or bind to cellular mRNA encoding the polypeptide of interest to thereby inhibit expression, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention includes 10 direct injection at a tissue, e.g., transplant or autoimmune lesion, site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell, e.g., T cell, surface, e.g., by linking the antisense nucleic acid molecules to 15 peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using vectors, e.g., gene therapy vectors, described below. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

An antisense nucleic acid molecule of interest can be an α-anomeric nucleic acid molecule. An α-anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β-units, the strands run parallel to each other (Gaultier et al. (1987) Nucleic Acids Res. 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue et al. (1987) Nucleic 25 Acids Res. 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue et al. (1987) FEBS Lett. 215:327-330).

Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region, and can also be generated using standard techniques. Thus, 30 ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) Nature 334:585-591)) can be used to catalytically cleave mRNA transcripts to thereby inhibit translation of the protein encoded by the mRNA. A ribozyme having specificity for a nucleic acid molecule encoding a polypeptide of interest can be designed based upon the nucleotide sequence of a cDNA disclosed herein. For example, a derivative of a 35 Tetrahymena L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in a Cech et al. U.S.

Patent No. 4,987,071; and Cech et al. U.S. Patent No. 5,116,742. Alternatively, an mRNA encoding a polypeptide of interest can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, e.g., Bartel and Szostak (1993) Science 261:1411-1418.

Triple helical structures can also be generated using well known techniques. For example, expression of a polypeptide of interest can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the gene encoding the polypeptide (e.g., the promoter and/or enhancer) to form triple helical structures that prevent transcription of the gene in target cells. See generally Helene (1991) Anticancer Drug Des. 10 6(6):569-84; Helene (1992) Ann. N.Y. Acad. Sci. 660:27-36; and Maher (1992) Bioassays 14(12):807-15.

In various embodiments, nucleic acid compositions can be modified at the base moiety, sugar moiety or phosphate backbone to improve, e.g., the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acids can be modified to generate peptide nucleic acids (see Hyrup et al. (1996) Bioorganic & Medicinal Chemistry 4(1): 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, e.g., DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for 20 specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup et al. (1996), supra; Perry-O'Keefe et al. (1996) Proc. Natl. Acad. Sci. USA 93: 14670-675.

PNAs can, for example, be modified, e.g., to enhance their stability or cellular 25 uptake, by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras can be generated which may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, e.g., RNAse H and DNA polymerases, to interact with the DNA portion while the 30 PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup (1996), supra). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup (1996), supra, and Finn et al. (1996) Nucleic Acids Res. 24(17):3357-63. For example, a DNA chain can be synthesized 35 on a solid support using standard phosphoramidite coupling chemistry and modified nucleoside analogs. Compounds such as 5'-(4-methoxytrityl)amino-5'-deoxy-thymidine

phosphoramidite can be used as a link between the PNA and the 5' end of DNA (Mag et al. (1989) Nucleic Acids Res. 17:5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn et al. (1996) Nucleic Acids Res. 24(17):3357-63). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment (Peterser et al. (1975) Bioorganic Med. Chem. Lett. 5:1119-11124).

In other embodiments, the oligonucleotide may include other appended groups such as peptides (e.g., for targeting host cell receptors in vivo), or agents facilitating transport across the cell membrane (see, e.g., Letsinger et al. (1989) Proc. Natl. Acad. Sci. USA 86:6553-6556; Lemaitre et al. (1987) Proc. Natl. Acad. Sci. USA 84:648-652; PCT Publication No. W0 88/09810) or the blood-brain barrier (see, e.g., PCT Publication No. W0 89/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (see, e.g., Krol et al. (1988) Bio/Techniques 6:958-976) or intercalating agents (see, e.g., Zon (1988) Pharm. Res. 5:539-549). To this end, the oligonucleotide may be conjugated to another molecule, e.g., a peptide, hybridization triggered cross-linking agent, transport agent, hybridization-triggered cleavage agent, etc.

ANTIBODY COMPOSITIONS

In one embodiment, anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3
20 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies or anti-SOCS3 antibodies are
administered to a mammal, preferably a human, to reduce immune rejection. In another
embodiment, any combination of anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3
antibodies, anti-Stat4, anti-SOCS1 antibodies and anti-SOCS3 antibodies are administered
to a mammal, preferably a human, to reduce immune rejection. In a preferred embodiment,
25 anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4 antibodies, antiSOCS1 antibodies or anti-SOCS3 antibodies are administered to a mammal, preferably a
human, in combination with other types of treatments (e.g., immunosuppressive agents) to
reduce immune rejection. In yet another preferred embodiment, any combination of antiStat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4, anti-SOCS1
30 antibodies and anti-SOCS3 antibodies are administered to a mammal, preferably a human,
in combination with other types of treatments (e.g., immunosuppressive agents) to reduce
immune rejection.

Anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can be administered to a mammal, preferably a human, using various delivery systems are known to those of skill in the art. For example, anti-Stat1 antibodies, anti-Stat2 antibodies,

anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can be administered by encapsulation in liposomes, microparticles or microcapsules. See, e.g., U.S. Patent No. 5,762,904, U.S. Patent No. 6,004,534, and PCT Publication WO 99/52563. In addition, anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can be administered using recombinant cells capable of expressing the antibodies, or retroviral, other viral vectors or non-viral vectors capable of expressing the antibodies.

Anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4, antiSOCS1 antibodies and anti-SOCS3 antibodies can be obtained from any known source. For example, anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4, antiSOCS1 antibodies and anti-SOCS3 antibodies can be obtained from Santa Cruz
Biotechnology, Inc. (Santa Cruz, CA), Research Diagnostics, Inc. (Flanders, NJ) or Zymed
Laboratories (South San Francisco, CA). Alternatively, anti-Stat1 antibodies, anti-Stat2
antibodies, anti-Stat3 antibodies, anti-Stat4, anti-SOCS1 antibodies and anti-SOCS3
antibodies can be produced by any method known in the art for the synthesis of antibodies, in particular, by chemical synthesis or preferably, by recombinant expression techniques.

Antibodies of the invention include, but are not limited to, polyclonal, monoclonal, multispecific, human, humanized or chimeric antibodies, single chain antibodies, Fab

20 fragments, F(ab') fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies (including, e.g., anti-Id antibodies to antibodies of the invention), and epitope-binding fragments. The term "antibody," as used herein, refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, i.e., molecules that contain an antigen binding site that immunospecifically binds an antigen.

25 The immunoglobulin molecules of the invention can be of any type (e.g., IgG, IgE, 1gM, IgD, IgA and IgY), class (e.g., IgG₁, IgG₂, IgG₃, IgG₄, IgA₁ and IgA₂) or subclass of immunoglobulin molecule. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments which can be generated by treating the antibody with an enzyme such as pepsin or papain.

An isolated Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3, or a fragment thereof, can be used as an immunogen to generate antibodies using standard techniques for polyclonal and monoclonal antibody preparation. The full-length Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 polypeptide or protein can be used or, alternatively, the invention provides antigenic peptide fragments of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 for use as immunogens. An antigenic peptide comprises at least 8 (preferably 10, 15, 20, or 30) amino acid residues of the amino acid sequence of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3,

and encompasses an epitope of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 such that an antibody raised against the peptide forms a specific immune complex with Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3.

The antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized polypeptide. If desired, the antibody molecules can be isolated from the mammal (e.g., from the blood) and further purified by well-known techniques, such as protein A chromatography to obtain the IgG fraction. Alternatively, antibodies specific for a protein or polypeptide of the invention can be selected for (e.g., partially purified) or 10 purified by, e.g., affinity chromatography. For example, a recombinantly expressed and purified (or partially purified) protein of the invention is produced as described herein, and covalently or non-covalently coupled to a solid support such as, for example, a chromatography column. The column can then be used to affinity purify antibodies specific for the proteins of the invention from a sample containing antibodies directed against a large 15 number of different epitopes, thereby generating a substantially purified antibody composition, i.e., one that is substantially free of contaminating antibodies. By a substantially purified antibody composition is meant, in this context, that the antibody sample contains at most only 30% (by dry weight) of contaminating antibodies directed against epitopes other than those on the desired Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 20 protein or polypeptide, and preferably at most 20%, yet more preferably at most 10%, and most preferably at most 5% (by dry weight) of the sample is contaminating antibodies. A purified antibody composition means that at least 99% of the antibodies in the composition are directed against the desired Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 protein or polypeptide.

At an appropriate time after immunization, e.g., when the specific antibody titers are highest, antibody-producing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the hybridoma technique originally described by Kohler and Milstein (1975) Nature 256:495-497, the human B cell hybridoma technique (Kozbor et al. (1983) Immunol. Today 4:72), the EBV-hybridoma technique (Cole et al. (1985), Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing hybridomas is well known (see generally Current Protocols in Immunology (1994) Coligan et al. (eds.) John Wiley & Sons, Inc., New York, NY). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind the polypeptide of interest, e.g., using a standard ELISA assay.

Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal antibody directed against Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 polypeptide can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (e.g., an antibody phage display library) with Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3.

Kits for generating and screening phage display libraries are commercially available (e.g., the Pharmacia Recombinant Phage Antibody System, Catalog No. 27-9400-01; and the Stratagene SurfZAPTM Phage Display Kit, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, U.S. Patent No. 5,223,409; PCT Publication No. WO 92/18619; PCT Publication No. WO 91/17271; PCT Publication No. WO 92/20791; PCT Publication No. WO 92/15679; PCT Publication No. WO 93/01288; PCT Publication No. WO 92/01047; PCT Publication No. WO 92/09690; PCT Publication No. WO 90/02809; Fuchs et al. (1991) Bio/Technology 9:1370-1372; Hay et al. (1992) Hum. Antibod. Hybridomas 3:81-85; Huse et al. (1989) Science 246:1275-1281; Griffiths et al. (1993) EMBO J. 12:725-734.

Additionally, recombinant antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, can be made using standard recombinant DNA techniques. A chimeric antibody is a molecule in which different portions are derived from different animal species, such as those having a variable region 20 derived from a murine mAb and a human immunoglobulin constant region. (See, e.g., Cabilly et al., U.S. Patent No. 4,816,567; and Boss et al., U.S. Patent No. 4,816397, which are incorporated herein by reference in their entirety.) Humanized antibodies are antibody molecules from non-human species having one or more complementarily determining regions (CDRs) from the non-human species and a framework region from a human 25 immunoglobulin molecule. (See, e.g., Queen, U.S. Patent No. 5,585,089, which is incorporated herein by reference in its entirety.) Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in PCT Publication No. WO 87/02671; European Patent Application 184,187; European Patent Application 171,496; European Patent Application 30 173,494; PCT Publication No. WO 86/01533; U.S. Patent No. 4,816,567; European Patent Application 125,023; Better et al. (1988) Science 240:1041-1043; Liu et al. (1987) Proc. Natl. Acad. Sci. USA 84:3439-3443; Liu et al. (1987) J. Immunol. 139:3521-3526; Sun et al. (1987) Proc. Natl. Acad. Sci. USA 84:214-218; Nishimura et al. (1987) Canc. Res. 47:999-1005; Wood et al. (1985) Nature 314:446-449; and Shaw et al. (1988) J. Natl. 35 Cancer Inst. 80:1553-1559); Morrison (1985) Science 229:1202-1207; Oi et al. (1986) Bio/Techniques 4:214; U.S. Patent 5,225,539; Jones et al. (1986) Nature 321:552-525;

Verhoeyan et al. (1988) Science 239:1534; and Beidler et al. (1988) J. Immunol. 141:4053-4060.

Completely human antibodies are particularly desirable for therapeutic treatment of human patients. Such antibodies can be produced, for example, using transgenic mice which are incapable of expressing endogenous immunoglobulin heavy and light chains genes, but which can express human heavy and light chain genes. The transgenic mice are immunized in the normal fashion with a selected antigen, e.g., all or a portion of a polypeptide of the invention. Monoclonal antibodies directed against the antigen can be obtained using conventional hybridoma technology. The human immunoglobulin 10 transgenes harbored by the transgenic mice rearrange during B cell differentiation, and subsequently undergo class switching and somatic mutation. Thus, using such a technique, it is possible to produce therapeutically useful IgG, IgA and IgE antibodies. For an overview of this technology for producing human antibodies, see Lonberg and Huszar (1995, Int. Rev. Immunol. 13:65-93). For a detailed discussion of this technology for 15 producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, e.g., U.S. Patent 5,625,126; U.S. Patent 5,633,425; U.S. Patent 5,569,825; U.S. Patent 5,661,016; and U.S. Patent 5,545,806. In addition, companies such as Abgenix, Inc. (Fremont, CA), can be engaged to provide human antibodies directed against a selected antigen using technology similar to that described above.

Completely human antibodies which recognize a selected epitope can be generated using a technique referred to as "guided selection." In this approach a selected non-human monoclonal antibody, e.g., a mouse antibody, is used to guide the selection of a completely human antibody recognizing the same epitope. (Jespers et al. (1994) Bio/technology 12:899-903).

As described herein, anti-Stat1, anti-Stat2, anti-Stat3, anti-Stat4, anti-SOCS1 or anti-SOCS3 antibodies can be used diagnostically to monitor protein levels within affected tissue (e.g., a transplant cell, tissue, organ or organ system, or a cell, tissue, organ or organ system that is, or is suspected of being affected by an autoimmune disorder) as part of a clinical testing procedure, e.g., to, for example, determine transplant rejection or the 30 efficacy of a given treatment regimen. Detection can be facilitated by coupling the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, beta-galactosidase, or acetylcholinesterase; examples of suitable 35 prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate,

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rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ³⁵S or ³H.

Further, as described herein, anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can be conjugated to a therapeutic moiety and administered to a mammal, preferably a human, to reduce or prevent immune rejection. Examples of therapeutic moieties that can be conjugated to antibodies include, but are not limited to, a 10 cytotoxin, a therapeutic agent or a radioactive metal ion. A cytotoxin or cytotoxic agent includes any agent that is detrimental to cells such as taxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, 15 lidocaine, propranolol, and puromycin and analogs or homologs thereof. Therapeutic agents include, but are not limited to, antimetabolites (e.g., methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine), alkylating agents (e.g., mechlorethamine, thioepa chlorambucil, melphalan, carmustine (BSNU) and lomustine (CCNU), cyclothosphamide, busulfan, dibromomannitol, streptozotocin, 20 mitomycin C, and cis-dichlorodiamine platinum (II) (DDP) cisplatin), anthracyclines (e.g., daunorubicin (formerly daunomycin) and doxorubicin), antibiotics (e.g., dactinomycin (formerly actinomycin), bleomycin, mithramycin, and anthramycin (AMC)), and anti-mitotic agents (e.g., vincristine and vinblastine).

The antibodies can also be conjugated a drug moiety that modifies a given 25 biological response. For example, a drug moiety may be a protein or polypeptide possessing a desired biological activity. Such proteins may include, for example, a toxin such as abrin, ricin A, pseudomonas exotoxin, or diphtheria toxin; and a lymphokine such as IL-4 or IL-13.

The invention also provides kits comprising an anti-Stat1 antibody, an anti-Stat2 30 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody, an anti-SOCS1 antibody, an anti-SOCS3 antibody, or any combination thereof conjugated to a detectable substance, and instructions for use. Still another aspect of the invention is a pharmaceutical composition comprising an anti-Stat1 antibody, an anti-Stat2 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody, an anti-SOCS1 antibody, an anti-SOCS3 antibody and a pharmaceutically 35 acceptable carrier. In preferred embodiments, the pharmaceutical composition contains an anti-Stat1 antibody, an anti-Stat2 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody,

an anti-SOCS1 antibody, or an anti-SOCS3 antibody, a therapeutic moiety, and a pharmaceutically acceptable carrier.

In instances wherein an anti-Stat1 antibody, an anti-Stat2 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody, an anti-SOCS1 antibody, an anti-SOCS3 antibody is to be utilized as a therapeutic, characterization of the antibody can routinely be assayed and ascertained via the methods presented herein. For example, the fact that lymphocytes and animal models for transplants and autoimmune disorders are readily available, coupled with the availability of multiple assays for Stat and SOCS expression and activity provide for routine testing and analysis (e.g., for in vitro and in vivo testing and analysis) of such 10 antibodies. The antibodies described herein can be tested, for example, for their ability to modulate the expression and/or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, SOCS3, or any combination thereof, and for their specificity and toxicity.

GENE THERAPY TECHNIQUES

Gene therapy refers to therapy performed by the administration to a subject of an 15 expressed or expressible nucleic acid. Any of the methods for gene therapy available in the art can be used according to the present invention. Exemplary methods are described below.

In one embodiment, Stat4 antisense oligonucleotides are administered to reduce immune rejection by way of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more anti-Stat4 antibodies are administered to reduce immune rejection, by way of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Jak2 for binding to the IL-12 receptor β_2 ("IL-12R β_2 ") are administered to immune rejection, by way 25 of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Tyk2 for binding to the IL-12R β_1 are administered to reduce immune rejection, by way of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat4 for binding to the IL-12R β_2 are administered to reduce immune rejection, by way 30 of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 from binding to the IL-12R β_2 are administered to reduce immune rejection, by way of gene therapy. In yet another embodiment, nucleic acid molecules comprising sequences encoding one or more dominant-negative Stat4 polypeptides (e.g., a Stat4 polypeptide 35 lacking its SH2 domain or a Stat4 polypeptide lacking its DNA binding domain) are administered to reduce immune rejection, by way of gene therapy.

In specific embodiments, Stat1 antisense oligonucleotides, Stat2 antisense oligonucleotides, Stat3 antisense oligonucleotides, or the combination thereof are administered to reduce immune rejection by way of gene therapy. In other embodiments, nucleic acid molecules comprising sequences encoding one or more anti-Stat1 antibodies, anti-Stat2 antibodies, or anti-Stat3 antibodies are administered to reduce immune rejection. by way of gene therapy. In other embodiments, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Jak1 or Tyk2 for binding to the IFN-α receptor or IFN-β receptor are administered to reduce immune rejection, by way of gene therapy. In yet another embodiment, nucleic acid molecules comprising sequences 10 encoding one or more peptides that compete with Jak1 or Jak2 for binding to the IFN-y receptor are administered to reduce immune rejection, by way of gene therapy. In other embodiments, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat1 or Stat2 for binding to the IFN-α receptor or IFN-β receptor are administered to reduce immune rejection, by way of gene therapy. In other embodiments, 15 nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat1 for binding to the IFN-y receptor are administered to reduce immune rejection, by way of gene therapy. In yet other embodiments, nucleic acid molecules comprising sequences encoding one or more dominant-negative Stat1 polypeptides (e.g., a Stat1 polypeptide lacking its SH2 domain or a Stat1 polypeptide lacking its DNA binding 20 domain), dominant-negative Stat2 polypeptides (e.g., a Stat2 polypeptide lacking its SH2 domain or a Stat2 polypeptide lacking its DNA binding domain), or dominant-negative Stat3 polypeptides (e.g., a Stat3 polypeptide lacking its SH2 domain or a Stat3 polypeptide lacking its DNA binding domain) are administered to reduce immune rejection, by way of gene therapy.

For general reviews of the methods of gene therapy, see Goldspiel et al., 1993, 25 Clinical Pharmacy 12:488-505; Wu and Wu, 1991, Biotherapy 3:87-95; Tolstoshev, 1993, Ann. Rev. Pharmacol. Toxicol. 32:573-596; Mulligan, 1993, Science 260:926-932; and Morgan and Anderson, 1993, Ann. Rev. Biochem. 62:191-217; May, 1993, TIBTECH 11(5):155-215). Methods commonly known in the art of recombinant DNA technology 30 which can be used are described in Ausubel et al. (eds.), 1993, Current Protocols in Molecular Biology, John Wiley & Sons, NY; and Kriegler, 1990, Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY.

In one aspect, a composition of the invention comprises nucleic acid sequences encoding one or more anti-Stat 1, anti-Stat 2, anti-Stat 3, anti-Stat 4, anti-SOCS 1, or anti-35 SOCS 3 antibodies or fragments thereof, said nucleic acid sequences being part of expression vectors that express one or more anti-Stat 1, anti-Stat 2, anti-Stat 3, anti-Stat 4,

anti-SOCS 1, or anti-SOCS 3 antibodies or fragments thereof in a suitable host. In particular, such nucleic acid sequences have promoters operably linked to the anti-Stat 1, anti-Stat 2, anti-Stat 3, anti-Stat 4, anti-SOCS 1, or anti-SOCS 3 antibodies or fragments thereof, said promoter being inducible or constitutive, and, optionally, tissue-specific.

In another aspect, a composition of the invention comprises nucleic acid sequences encoding dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 polypeptides, said nucleic acid sequences being part of expression vectors that express dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 polypeptides in a suitable host. In particular, such nucleic acid sequences have promoters operably linked to the dominant-negative Stat1, 10 Stat2, Stat3, Stat4, SOCS1, or SOCS3, said promoter being inducible or constitutive, and, optionally, tissue-specific. In another particular embodiment, nucleic acid molecules are used in which the dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 coding sequences and any other desired sequences are flanked by regions that promote homologous recombination at a desired site in the genome, thus providing for intrachromosomal 15 expression of the dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 nucleic acids (Koller and Smithies, 1989, Proc. Natl. Acad. Sci. USA 86:8932-8935; Zijlstra et al., 1989, Nature 342:435-438).

In another aspect of the invention, a composition of the invention comprises nucleic acid molecules comprising sequences encoding one or more peptides that compete with 20 Tyk2 for binding to the IL-12R β_1 , said nucleic acid sequences being part of expression vectors that express one or more peptides in a suitable host. In another aspect of the invention, a composition of the invention comprises nucleic acid sequences nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat4 for binding to the IL-12Rβ₂, said nucleic acid sequences being part of expression vectors that 25 express one or more peptides in a suitable host. In yet another aspect of the invention, a composition of the invention comprises nucleic acid molecules comprising sequences encoding one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 from binding to the IL-12Rβ₂, said nucleic acid sequences being part of expression vectors that express one or more peptides in a suitable host. In particular a embodiment of the 30 invention, the nucleic acid sequences encoding peptides of the invention have promoters operably linked to said nucleic acid sequences, said promoter being inducible or constitutive, and, optionally, tissue-specific.

Delivery of the nucleic acids into a patient may be either direct, in which case the patient is directly exposed to the nucleic acid or nucleic acid-carrying vectors, or indirect, in 35 which case, cells are first transformed with the nucleic acids in vitro, then transplanted into

the patient. These two approaches are known, respectively, as in vivo or ex vivo gene therapy.

In a specific embodiment, the nucleic acid sequence is directly administered in vivo, where it is expressed to produce the encoded product. This can be accomplished by any of numerous methods known in the art, e.g., by constructing it as part of an appropriate nucleic acid expression vector and administering it so that they become intracellular, e.g., by infection using defective or attenuated retrovirals or other viral vectors (see U.S. Patent No. 4,980,286), or by direct injection of naked DNA, or by use of microparticle bombardment (e.g., a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or 10 transfecting agents, encapsulation in liposomes, microparticles, or microcapsules, or by administering them in linkage to a peptide which is known to enter the nucleus, by administering it in linkage to a ligand subject to receptor-mediated endocytosis (see, e.g., Wu and Wu, 1987, J. Biol. Chem. 262:4429-4432) (which can be used to target cell types specifically expressing the receptors), etc. In another embodiment, nucleic acid-ligand 15 complexes can be formed in which the ligand comprises a fusogenic viral peptide to disrupt endosomes, allowing the nucleic acid to avoid lysosomal degradation. In yet another embodiment, the nucleic acid can be targeted in vivo for cell specific uptake and expression. by targeting a specific receptor (see, e.g., PCT Publications WO 92/06180 dated April 16, 1992 (Wu et al.); WO 92/22635 dated December 23, 1992 (Wilson et al.); WO92/20316 20 dated November 26, 1992 (Findeis et al.); WO93/14188 dated July 22, 1993 (Clarke et al.), WO 93/20221 dated October 14, 1993 (Young)). Alternatively, the nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination (Koller and Smithies, 1989, Proc. Natl. Acad. Sci. USA 86:8932-8935; Zijlstra et al., 1989, Nature 342:435-438).

In one embodiment, viral vectors that contain Stat4 antisense oligonucleotides are used (see Miller et al., 1993, Meth. Enzymol. 217:581-599). In another embodiment, viral vectors that contain nucleic acids encoding one or more anti-Stat4 antibodies are used. In another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that compete with Jak2 for binding to the IL-12 receptor β_2 ("IL-12R β_2 ") are used. In 30 another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that compete with Tyk2 for binding to the IL-12RB, are used. In another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that compete with Stat4 for binding to the IL-12Rβ₂ are used. In another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 35 from binding to the IL-12R β_2 are used. In yet another embodiment, viral vectors that contain nucleic acids encoding dominant-negative Stat4 polypeptides (e.g., Stat4 lacking its

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SH2 domain or Stat4 lacking its DNA binding domain) are used. For example, a retroviral vector can be used. These retroviral vectors have been modified to delete retroviral sequences that are not necessary for packaging of the viral genome and integration into host cell DNA. The nucleic acid sequences encoding the Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 antibodies, or polypeptides or peptides of the invention to be used in gene therapy are cloned into one or more vectors, which facilitates delivery of the gene into a patient. More detail about retroviral vectors can be found in Boesen et al., 1994, *Biotherapy* 6:291-302, which describes the use of a retroviral vector to deliver the mdr1 gene to hematopoietic stem cells in order to make the stem cells more resistant to chemotherapy.

10 Other references illustrating the use of retroviral vectors in gene therapy are: Clowes et al., 1994, *J. Clin. Invest.* 93:644-651; Kiem et al., 1994, *Blood* 83:1467-1473; Salmons and Gunzberg, 1993, *Human Gene Therapy* 4:129-141; and Grossman and Wilson, 1993, *Curr. Opin. in Genetics and Devel.* 3:110-114.

Adenoviruses are other viral vectors that can be used in gene therapy. Adenoviruses are especially attractive vehicles for delivering genes to respiratory epithelia. Adenoviruses naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are liver, the central nervous system, endothelial cells, and muscle. Adenoviruses have the advantage of being capable of infecting non-dividing cells. Kozarsky and Wilson, 1993, Current Opinion in Genetics and Development

20 3:499-503 present a review of adenovirus-based gene therapy. Bout et al., 1994, Human Gene Therapy 5:3-10 demonstrated the use of adenovirus vectors to transfer genes to the respiratory epithelia of rhesus monkeys. Other instances of the use of adenoviruses in gene therapy can be found in Rosenfeld et al., 1991, Science 252:431-434; Rosenfeld et al., 1992, Cell 68:143-155; Mastrangeli et al., 1993, J. Clin. Invest. 91:225-234; PCT Publication

25 WO94/12649; and Wang, et al., 1995, Gene Therapy 2:775-783. In a preferred embodiment, adenovirus vectors are used.

Adeno-associated virus (AAV) has also been proposed for use in gene therapy (Walsh et al., 1993, *Proc. Soc. Exp. Biol. Med.* 204:289-300; U.S. Patent No. 5,436,146).

Another approach to gene therapy involves transferring a gene to cells in tissue culture by such methods as electroporation, lipofection, calcium phosphate mediated transfection, or viral infection. Usually, the method of transfer includes the transfer of a selectable marker to the cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene. Those cells are then delivered to a patient.

In this embodiment, the nucleic acid is introduced into a cell prior to administration in vivo of the resulting recombinant cell. Such introduction can be carried out by any

method known in the art, including but not limited to transfection, electroporation, microinjection, infection with a viral or bacteriophage vector containing the nucleic acid sequences, cell fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion, etc. Numerous techniques are known in the art for the introduction of foreign genes into cells (see, e.g., Loeffler and Behr, 1993, Meth. Enzymol. 217:599-618; Cohen et al., 1993, Meth. Enzymol. 217:618-644; Cline, 1985, Pharmac. Ther. 29:69-92) and may be used in accordance with the present invention, provided that the necessary developmental and physiological functions of the recipient cells are not disrupted. The technique should provide for the stable transfer of the nucleic acid to the cell, so that 10 the nucleic acid is expressible by the cell and preferably heritable and expressible by its cell progeny.

The resulting recombinant cells can be delivered to a patient by various methods known in the art. Recombinant blood cells (e.g., hematopoietic stem or progenitor cells) are preferably administered intravenously. The amount of cells envisioned for use depends on 15 the desired effect, patient state, etc., and can be determined by one skilled in the art.

Cells into which a nucleic acid can be introduced for purposes of gene therapy encompass any desired, available cell type, and include but are not limited to epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes; blood cells such as T lymphocytes, B lymphocytes, monocytes, macrophages, neutrophils, eosinophils, 20 megakaryocytes, granulocytes; various stem or progenitor cells, in particular hematopoietic stem or progenitor cells, e.g., as obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, etc.

In a preferred embodiment, the cell used for gene therapy is autologous to the patient.

In one embodiment in which recombinant cells are used in gene therapy, nucleic acid 25 sequences encoding Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 antibodies, or polypeptides or peptides of the invention are introduced into the cells such that they are expressible by the cells or their progeny, and the recombinant cells are then administered in vivo for therapeutic effect. In a specific embodiment, stem or progenitor cells are used.

30 Any stem and/or progenitor cells which can be isolated and maintained in vitro can potentially be used in accordance with this embodiment of the present invention (see e.g. PCT Publication WO 94/08598, dated April 28, 1994; Stemple and Anderson, 1992, Cell 71:973-985; Rheinwald, 1980, Meth. Cell Bio. 21A:229; and Pittelkow and Scott, 1986, Mayo Clinic Proc. 61:771).

Promoters that may be used to control the expression of nucleic acid sequences encoding Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 antibodies, or polypeptides or

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peptides of the invention include, but are not limited to, the SV40 early promoter region (Bernoist and Chambon, 1981, Nature 290:304-310), the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto, et al., 1980, Cell 22:787-797), the herpes thymidine kinase promoter (Wagner et al., 1981, Proc. Natl. Acad. Sci. USA 78:1441-1445), the regulatory sequences of the metallothionein gene (Brinster et al., 1982, Nature 296:39-42); prokaryotic expression vectors such as the β -lactamase promoter (Villa-Kamaroff et al., 1978, Proc. Natl. Acad. Sci. USA 75:3727-3731), or the tac promoter (DeBoer et al., 1983, Proc. Natl. Acad. Sci. USA 80:21-25); see also "Useful proteins from recombinant bacteria" in Scientific American, 1980, 242:74-94; plant 10 expression vectors comprising the nopaline synthetase promoter region (Herrera-Estrella et al., Nature 303:209-213) or the cauliflower mosaic virus 35S RNA promoter (Gardner et al., 1981, Nucl. Acids Res. 9:2871), and the promoter of the photosynthetic enzyme ribulose biphosphate carboxylase (Herrera-Estrella et al., 1984, Nature 310:115-120); promoter elements from yeast or other fungi such as the Gal 4 promoter, the ADC (alcohol 15 dehydrogenase) promoter, PGK (phosphoglycerol kinase) promoter, alkaline phosphatase promoter, and the following animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals: elastase I gene control region which is active in pancreatic acinar cells (Swift et al., 1984, Cell 38:639-646; Ornitz et al., 1986, Cold Spring Harbor Symp. Quant. Biol. 50:399-409; MacDonald, 1987, Hepatology 20 7:425-515); insulin gene control region which is active in pancreatic beta cells (Hanahan, 1985, Nature 315:115-122), immunoglobulin gene control region which is active in lymphoid cells (Grosschedl et al., 1984, Cell 38:647-658; Adames et al., 1985, Nature 318:533-538; Alexander et al., 1987, Mol. Cell. Biol. 7:1436-1444), mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells 25 (Leder et al., 1986, Cell 45:485-495), albumin gene control region which is active in liver (Pinkert et al., 1987, Genes and Devel. 1:268-276), alpha-fetoprotein gene control region which is active in liver (Krumlauf et al., 1985, Mol. Cell. Biol. 5:1639-1648; Hammer et al., 1987, Science 235:53-58; alpha 1-antitrypsin gene control region which is active in the liver (Kelsey et al., 1987, Genes and Devel. 1:161-171), beta-globin gene control region which is 30 active in myeloid cells (Mogram et al., 1985, Nature 315:338-340; Kollias et al., 1986, Cell 46:89-94; myelin basic protein gene control region which is active in oligodendrocyte cells in the brain (Readhead et al., 1987, Cell 48:703-712); myosin light chain-2 gene control region which is active in skeletal muscle (Sani, 1985, Nature 314:283-286), and gonadotropic releasing hormone gene control region which is active in the hypothalamus 35 (Mason et al., 1986, Science 234:1372-1378).

In a specific embodiment, the nucleic acid to be introduced for purposes of gene therapy comprises an inducible promoter operably linked to the coding region, such that expression of the nucleic acid is controllable by controlling the presence or absence of the appropriate inducer of transcription.

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PHARMACEUTICAL COMPOSITIONS

The nucleic acid molecules, polypeptides, antibodies and small molecules (also referred to herein as "active compounds") described herein can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the active compound and a pharmaceutically acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

The invention includes methods for preparing pharmaceutical compositions for modulating the expression or activity of a polypeptide or nucleic acid of interest (e.g., Stat1, Stat2, Stat3, Stat4, Stat6, SOS 1, or SOCS3). Such methods comprise formulating a pharmaceutically acceptable carrier with an agent which modulates expression or activity of a polypeptide or nucleic acid of interest (e.g., Stat1, Stat2, Stat3, Stat4, Stat6, SOS 1, or SOCS3). Such compositions can further include additional active agents. Thus, the invention further includes methods for preparing a pharmaceutical composition by formulating a pharmaceutically acceptable carrier with an agent which modulates expression or activity of a polypeptide or nucleic acid of interest and one or more additional active compounds.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, e.g., intravenous, intradermal, subcutaneous, oral (e.g., inhalation), transdermal (topical), transmucosal, and rectal administration. Intravenous administration is preferred. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic

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acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor ELTM (BASF; Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be 10 sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyetheylene glycol, and the like), and suitable 15 mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be 20 preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound 25 (e.g., a polypeptide or antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation 30 of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral 35 therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared

using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed.

Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl 10 salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from a pressurized container or dispenser which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For 15 transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are 20 formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect 25 the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be 30 obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used

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herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

For antibodies, the preferred dosage is 0.1 mg/kg to 100 mg/kg of body weight (more preferably, 0.1 to 20 mg/kg, 0.1-10 mg/kg, or 0.1 to to 1.0 mg/kg). If the antibody is to act in the brain, a dosage of 50 mg/kg to 100 mg/kg is usually appropriate. Generally, partially human antibodies and fully human antibodies have a longer half-life within the human body than other antibodies. Accordingly, lower dosages and less frequent administration is often possible. Modifications such as lipidation can be used to stabilize antibodies and to enhance uptake and tissue penetration (e.g., into the brain). A method for lipidation of antibodies is described by Cruikshank et al. ((1997) J. Acquired Immune Deficiency Syndromes and Human Retrovirology 14:193).

As defined herein, a therapeutically effective amount of protein or polypeptide (*i.e.*, an effective dosage) ranges from about 0.001 to 30 mg/kg body weight, preferably about 0.01 to 25 mg/kg body weight, more preferably about 0.1 to 20 mg/kg body weight, and even more preferably about 0.1 to 1.0 mg/kg, 1 to 10 mg/kg, 2 to 9 mg/kg, 3 to 8 mg/kg, 4 to 7 mg/kg, or 5 to 6 mg/kg body weight.

The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of a protein, polypeptide, or antibody can include a single treatment or, preferably, can include a series of treatments.

In a representative, non-limiting example, a subject is treated at the time of transplantation, or when diagnosed as exhibiting a reoccurrence of rejection, or an occurrence of a rejection (e.g., an autoimmune rejection) with one to several (for example, between 3 and 7) doses of an appropriate modulator of Stat1, Stat2, Stat3, Stat4, Stat6, SOS 1, and/or SOCS3 for a maximum of one week. In a preferred embodiment of such an example, treatment would further comprise additional administration approximately once per month for about 3 to 6 months. The preferred route of administration is intravenous bolus injection. It will also be appreciated that the effective dosage of the modulator used for treatment may increase or decrease over the course of a particular treatment. Changes in

dosage may result and become apparent from the results of diagnostic assays as described herein.

Preferably, administration of modulator is by intravenous injection, and can also be are or near the site of the cells or tissue to be treated, e.g., administration is at or near the site of the transplant or autoimmune disorder lesion.

In addition to those compounds described above, the present invention encompasses agents and use of agents which modulate expression or activity of a nucleic acid or polypeptide of interest. An agent may, for example, be a small molecule. For example, such small molecules include, but are not limited to, peptides, peptidomimetics, amino 10 acids, amino acid analogs, polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic compounds (i.e,. including heteroorganic and organometallic compounds) having a molecular weight less than about 10,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 5,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 1,000 grams per 15 mole, organic or inorganic compounds having a molecular weight less than about 500 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of such compounds. It is understood that appropriate doses of small molecule agents depends upon a number of factors within the ken of the ordinarily skilled physician, veterinarian, or researcher. The dose(s) of the small molecule will vary, for example, depending upon the 20 identity, size, and condition of the subject or sample being treated, further depending upon the route by which the composition is to be administered, if applicable, and the effect which the practitioner desires the small molecule to have upon the nucleic acid or polypeptide of the invention. Exemplary doses include milligram or microgram amounts of the small molecule per kilogram of subject or sample weight (e.g., about 1 microgram per kilogram to 25 about 500 milligrams per kilogram, about 100 micrograms per kilogram to about 5 milligrams per kilogram, or about 1 microgram per kilogram to about 50 micrograms per kilogram. It is furthermore understood that appropriate doses of a small molecule depend upon the potency of the small molecule with respect to the expression or activity to be modulated. Such appropriate doses may be determined using the assays described herein. 30 When one or more of these small molecules is to be administered to an animal (e.g., a human) in order to modulate expression or activity of a polypeptide or nucleic acid of the invention, a physician, veterinarian, or researcher may, for example, prescribe a relatively low dose at first, subsequently increasing the dose until an appropriate response is obtained. In addition, it is understood that the specific dose level for any particular animal subject will

35 depend upon a variety of factors including the activity of the specific compound employed,

the age, body weight, general health, gender, and diet of the subject, the time of

administration, the route of administration, the rate of excretion, any drug combination, and the degree of expression or activity to be modulated.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

In one embodiment, one or more compositions for modulation of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3 can be used combinatorially. For example, compositions for decreasing expression or activity of Stat4 can be utilized in combination (either simultaneously or serially) with compositions or techniques for increasing expression or activity of Stat6 can be utilized.

In another embodiment, one or more compositions of the present invention that modulate expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be administered to a mammal, preferably a human, in combination with one or more standard immunosuppressive or immunomodulatory compounds to reduce or prevent immune rejection resulting from an autoimmune disorder or an allograft. Examples of immunosuppressive agents include, but are not limited to, azathioprine, corticosteriods (e.g., prednisone), cyclosporine, OKT3 (anti-CD3 monoclonal human antibody), mycophenolate mofetil, rapamycin (sirolimus), mizoribine, deoxyspergualin, macrolide antibiotics such as, for example, FK506 (tacrolimus), brequinar, malononitriloamindes.(e.g., leflunamide), and anti-IL-2R antibodies (e.g., anti-Tac monoclonal antibody and BT 536).

See, e.g., Grummet et al., 1999, J. Am. Soc. Nephrol. 10:1366-1388; and Norman and Wadi, eds., 1998, "Primer on Transplantation," Am. Soc. Tx. Phys, 1st ed.).

Immunosuppressive agents may be administered at high doses initially and then tapered off over time to reduce or prevent immune rejection. For example, one or more compositions of the invention in combination with an initial dose of cyclosporine ranging from between 5 and 10 mg/kg per day, an initial dose of 10mg/kg per day prednisone, or an initial dose of 10mg/kg per day mycophenolate mofetil may be administered to animal to reduce or prevent immune rejection. Alternatively, one or more compositions of the invention in combination with an initial dose of cyclosporine ranging from between 5 and 10 mg/kg per day, an initial dose of 10mg/kg per day prednisone, and an initial dose of 10mg/kg per day mycophenolate mofetil may be administered to animal to reduce or prevent immune rejection. Preferably, corticosteroids are not administered children.

In yet another embodiment, one or more compositions of the present invention that modulate expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be administered to a mammal, preferably a human, in combination with one or more standard autoimmune therapeutic agents. used for treating a particular autoimmune disorder. For example, one or more compositions of the present invention may be administered in

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combination with one or more conventional anti-lupus therapeutic agents or drugs such as, for example, salicylates, corticosteroids, and immunosuppresants.

In another embodiment, one or more compositions of the present invention for modulating the expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3 are administered to a mammal, preferably a human, in combination with one or more T celltargeted or B cell-targeted agents. Examples of such agents include, but are limited to, CTLA-4Ig, IL-2 antagonists (e.g., anti-IL-2 receptor antibodies and IL-2 toxin conjugates), B7 monoclonal antibodies, anti-CD40L monoclonal antibodies, CD4 antagonists (e.g., anti-CD4 monoclonal antibodies), CD3 antagonists (e.g., anti-CD3 monoclonal antibodies), and 10 IL-12 antagonists (e.g., anti-IL-12 monoclonal antibodies and IL-12 toxin conjugates) to reduce or prevent immune rejection an autoimmune disorder or an allograft.

In order that the invention described herein may be more fully understood, the following examples are set forth. It should be understood that these examples are for illustrative purposes only and are not to be construed as limiting this invention in any manner.

EXAMPLE: QUANTITATIVE ANALYSIS OF NF-KB AND IKB 6. PROTEINS IN MOUSE CARDIAC ALLOGRAFTS

NF-κB proteins are transcription factors complexed with IκB proteins in the cytoplasm but which upon cell activation are released, translocate to the nucleus and bind κB motifs in the promoters of many genes, in particular of the promoters of genes whose expression is involved the immune response. Since NF-κB plays an important role in the transcription of genes involved in immune responses, the expression levels of NF-kB and IkB proteins and their localization were determined in mouse cardiac allografts. The data generated and analyzed represents the first comprehensive analysis of NF-kB and IkB protein expression, phosphorylation, and localization as detected by Western blotting and immunohistology in serially harvested allografts (BALB/c - B6), isoftafts and native hearts from recipients treated with IgG (rejection by day 8) or CD40 ligand monoclonal antibody ("CD40L mAb"; permanent survival).

Heterotopic cardiac allografting was performed with anastomoses to the abdominal aorta and vena cava (Hancock et al., 1998, Nature Medicine 4:1392-1396), using BALB/c donors and B6/129 wild-type or other Balb/c wild-type mice as recipients. Recipients were treated with hamster IgG or hamster anti-mouse (CD40L mAb; 250 µg, administered intravenously) plus DST (5 x 10⁶ splenic mononuclear cells) at the time of transplantation (Hancock et al., 1998, Nature Medicine 4:1392-1396). The protein expression, phosphorylation and localization of NF-κB and IκB were detected by Western blot analysis

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and immunohistology using serially harvested allografts, isografts and native hearts from recipients treated with IgG (rejection by day 8) or CD40L mAb (permanent survival).

The following tissue-specific expression patterns in the basal cardiac state relative to other organs were detected: low p50; low p52; low p65; low C-rel; low relB; low IkBa; high IkBb; low IkBe; low BCL-3 and high p105. The level of NF-κB proteins was significantly increased (2-4 fold) upon allografting and these levels were only modestly affected by CD40L mAb. Significant increases in the levels of IκBa (greater than 2 fold) and IκBe (greater than 6 fold) proteins were also detected in cardiac allografts. In contrast, a significant decrease in the level of IκBb protein, low to undetectable levels of p105 protein, and trace levels of BCL-3 were detected in cardiac allografts, but were only modestly affected by CD40L mAb. Thus, these results demonstrate that in cardiac allografts the NF-κB regulatory apparatus is highly activated at the protein level and is only modestly affected by CD40L mAb.

Cardiac samples principally express p105 and IκBb, but these are down-regulated during rejection, presumably through the action of the proteasome. By contrast, cardiac allograft rejection is associated primarily with expression by infiltrating leukocytes of p65, p50 and c-rel NF-κB proteins, plus IκBa and IκBc proteins.

The results, therefore, suggest that monitoring of the levels of NF-κB and IκB proteins in biopsies from transplant recipients may be of diagnostic and/or prognostic significance.

7. EXAMPLE: DIFFERENTIAL EFFECTS OF IMMUNOSUPPRESSIVE AGENTS ON ANTI-CD40L ANTIBODY-MEDIATED TOLERANCE INDUCTION

The data presented herein demonstrate that concomitant use of the immunosuppressive agents cyclosporin A or methylprednisolone, but not rapamycin, blocks CD154 mAb efficacy in experimental allograft recipients. Indeed, the differential effects of these agents on CD154 mAb-induced tolerance correlates with their capacity to inhibit activation-induced CD154 expression on CD4+ T cells. Full expression of CD154 expression was found to require NF-κB activation, and CD154 mAb was ineffective in NF-κB/p50 deficient allograft recipients or control mice in which NF-κB activation was blocked by a proteasome inhibitor. Hence, these data indicate that strategies to use CD154 mAb clinically must take into account the effects of immunosuppressive agents on CD154 induction, which appears to be at least partially NF-κB dependent, and suggest that ligation of surface-expressed CD154 provides an important signal that modulates T cell activation.

MATERIALS & METHODS

Media and Reagents:

Cell culture media, serum and supplements were purchased from Gibco BRL (Rockville. MD) and all mAbs were from PharMingen (San Diego, CA). Cyclosporin A (catalog C-3662, Sigma. St. Louis. MO) was prepared as a 5 mg/ml stock solution in 0.9% saline; rapamycin (catalog 380-004-M001, Alexis, San Diego. CA) as a 1 mg/ml stock solution in ethanol: 6α-methylprednisolone (catalog M-0369, Sigma) as a 5 mg/ml stock solution in 80% ethanol; mycophenolate mofetil (catalog M-5255, Sigma) as a 20 mM stock solution in DMSO; and the 3 proteasome inhibitors (Grisham, M.B., et al., 1999, Methods 10 Enzymol 300:345-63), clasto-lactacystin β-lactone (catalog 426102, Calbiochem, San Diego, CA) and its derivative PS-519 (Proscript, Cambridge, MA), and dipeptide-boronate (MG-273. ProScript), each as a 10 mM stock solution in DMSO.

Mice:

5

BALB/c (H-2^d) and B6/129 (H-2^b) mice were obtained from Jackson Labs (Bar 15 Harbor, ME), and NF-κB/p50 knockout (p50 KO) B6/129 mice (H-2^b) (Sha, W.C. et al., 1995, Cell 80:321-30.) were provided by Dr. David Baltimore (MIT, Cambridge, MA), and were housed under specific pathogen-free conditions.

20 Cardiac Transplantation:

Heterotopic cardiac allografting was performed with anastomoses to the abdominal aorta and vena cava (Hancock, W.W. et al., 1998, Nature Medicine 4:1392-1396), using BALB/c donors and B6/129 wild-type or NF-kB/p50 KO mice as recipients (n=6/group). Recipients were treated with hamster IgG or hamster anti-mouse CD 154 mAb (250 µg, i.v.) 25 plus DST ("donor specific transfusion"; 5 x 106 splenic mononuclear cells) at the time of transplantation (Hancock, W.W. et al., 1998, Nature Medicine 4:1392-1396). Additional groups of allografted wild-type mice were treated with CD 154 mAb/DST plus (i) rapamycin (0.2 mg/kg/d i.p.) or (ii) cyclosporin A (10 mg/kg/d i.p.) on day 0 and every other day until day 14; (iii) methylprednisolone (1 mg/kg i.p.) on day 0, 1 and 2; and (iv) 30 PS-519 (1 mg/kg/d i.p.) daily from the time of transplantation. Graft survival was monitored by daily palpation, and rejection was confirmed by laparotomy and histologic evaluation.

Measurement of activation-induced CD154 expression:

Six-well plates (Costar) were pre-coated overnight with 1 µg/ml of rat anti-mouse 35 CD3 mAb (2C11). After washing with media (RPMI supplemented with 10% fetal bovine

serum, 50 U/ml penicillin, 50 μg/ml streptomycin, 2 mM glutamine and 50 μM 2mercaptoethanol). one ml of media containing 2X final drug concentration was added to each well. After testing of a range of concentrations of each agent, the optimal final physiologic drug concentrations for the data to be reported were 125 ng/ml cyclosporin A, 20 ng/ml rapamycin. 100 µg/ml methylprednisolone, 1 μM mycophenolate mofetil, 10 μm lactacystin and 10 μM MG-273. Cells were obtained from mechanically disrupted spleens and, after collection by centrifugation and washing once with media, one ml aliquots containing two million viable splenocytes were added to each well. Cultures were incubated at 37°C in 5% CO₂ for 7 hr. 10 and were then diluted two-fold with chilled media containing 0.1% sodium azide (FACS media). Cells were collected by centrifugation at 4°C, resuspended in FACS media containing 1 µg Fc Block (PharMingen) and incubated on ice for 15 min. Samples were then split and either a mixture of 1 µg each of FITC-conjugated CD4-specific mAb and PEconjugated control hamster IgG mAb or FITC-conjugated CD4-specific mAb and 15 PE-conjugated CD 154-specific mAb were added. After a 30 min incubation on ice, cells were washed twice and analyzed on a Becton Dickinson FACScan using Cell Quest software. Viable CD4+ lymphocytes were gated using a combination of forward/side scatter and CD4 staining.

20 Statistics

Flow cytometry data was analyzed using student t-test and cardiac allograft survival was evaluated by the two-tailed Mann-Whitney U test using the program InStat (GraphPad Software, San Diego. CA); p.<0.05 indicated a significant result.

25 RESULTS

Given the apparent efficacy of CD154 mAb in inducing long-term cardiac allograft survival in various mouse strains (Larsen, C.P. et.al., 1996, Nature 381:434-438; Hancock. W. W. et al., 1996 Proc. Natl. Acad. Sci. USA. 93:13967-13972), without development of transplant arteriosclerosis or other sequelae of chronic rejection (Hancock, W.W. et al., 1998, Nature Medicine 4:1392-1396), investigators have begun clinical testing. As human allograft recipients presently derive significant benefit from conventional immunosuppressant therapy, initial CD 154 mAb trial designs have included pharmacologic immunosuppression. However, conventional immunosuppression with glucocorticoids or cyclosporin A has prevented the success of otherwise potent experimental approaches for tolerance induction, such as that seen with intrathymic injection of donor MHC antigen in rodents (Perico, N. et al.,1995, J. Amer. Soc. Nephrol. 5:1618-1623). Likewise, CD154

mAb-induced prolongation of renal allograft survival in the initial primate studies was diminished by concomitant use of either glucocorticoids or FK506 (Kirk, A.D.et. Al, 1999, Nature Medicine 5:686-693): FK-506, like cyclosporin A, blocks activation of the calciumdependent serine phosphatase calcineurin.

To understand the potential for inhibitory effects of immunosuppressive agents on the therapeutic efficacy of CD154 mAb, the effectiveness of CD154 mAb was investigated in conjunction with adjunct therapies in a completely MHC-mismatched (H2^d->N2^b) mouse cardiac allograft model. The results of concomitant administration of cyclosporin A. methylprednisolone or rapamycin on allograft survival in mice treated with CD154 mAb 10 plus DST are summarized in Figure 10. Whereas CD154 mAb therapy induced permanent cardiac allograft survival (>100 days), the effects of CD154 mAb were blocked by addition of cyclosporin A or methylprednisolone (p<0.001 vs. CD154 mAb/DST alone), but not by rapamycin (Figure 10).

Given these markedly contrasting differences in effects of standard 15 immunosuppressive agents on the efficacy of CD154 mAb therapy, and recent evidence that CD154 may signal to T cells (Blair, P.J. et al., 2000, J Exp Med 191:651-660), it was hypothesized that inhibition of CD 154 expression was involved. Accordingly, the in vitro effects of standard immunosuppressive agents on the expression of CD154 by activated T cells were tested in vitro. It was found that whereas resting CD4+ splenic cells lacked CD 20 154 expression, 25-30% of cells expressed the molecule within 7 hours of activation with plate-bound CD3 mAb (Fig. 11A). This upregulation was markedly suppressed by therapeutic doses of cyclosporin A or methylprednisolone, but not by rapamycin or mycophenolate mofetil (Figure 11a). Analysis of three separate experiments showed >90% suppression by methylprednisolone (p<0.01) and >70% suppression by cyclosporin A 25 (p<0.01) whereas the effects of rapamycin (<20% inhibition) and mycophenolate mofetil (<10% inhibition) were not statistically distinguishable from control activated cells (Figure 11B). Thus, pharmacologic immunosuppressives that reduce the efficacy of CD154 mAb in vivo also inhibit activation-induced CD154 expression in vitro.

Since the allograft response is highly T cell-dependent, immunosuppressants must 30 modulate one or more aspects of the T cell response. As reviewed in this context (Gummert, J.F. et al., 1999, J. Am. Soc. Nephrol. 10:1366-1380), full T cell activation requires three signals: signal 1 is the triggering of the T cell antigen receptor, signal 2 is costimulation through CD28 and related molecules, and signal 3 is provided by cytokines. Cyclosporin A, like FK-506, blocks activation of calcineurin, an early event in T cell 35 activation, just downstream of signal 1. Calcineurin dephosphorylates the transcription factor, nuclear factor of activated T cells (NFAT), and though it is well known that NFAT is

essential to the transcription of IL-2 and (FN-γ (Ullman, K.S. et al., 1990, Anna Rev Immunol 8:421-52), it is less appreciated that NFATp-binding sites are also present in the CD154 promoter (Schubert, L.A.,et al., 1995, J. Biol Chem 270:29624-7). It was found here that CD154 induction on murine CD4+ T cells was markedly inhibited by cyclosporin A, as was reported for human T cells (Fuleihan. R., et al., 1994., J. Clin Invest 93:1315-20).

In contrast to cyclosporin A, the macrolide rapamycin blocks a relatively late stage of T cell activation, as the target of rapamycin ("TOR"), is downstream of signal 3, cytokine receptor activation. Notably, though rapamycin is known to block various CD28-mediated events, and costimulation though CD28 can augment CD154 expression (Klaus. S.J., et al., 1994, J. Immunol 152:5643-5652), it was found here that rapamycin had no significant inhibitory effect on CD154 induction, suggesting that the effects of CD28 costimulation on CD154 expression are not critical. Hence, our data suggest that the contrasting efficacies of rapamycin and other immunophilin-binding agents such as cyclosporin A or FK506 in trials of CD154 mAb correlate with their capacities to block CD154 induction and may reflect inhibition of late and early T cell activation events, respectively.

The immunosuppressive agent mycophenolate mofetil blocks the final stage of T cell activation. T cell proliferation, which requires de novo synthesis of purine and pyrimidine nucleotides. Mycophenolate mofetil is converted within lymphocytes to its active 20 metabolite, mycophenolic acid, which is a reversible inhibitor of a key enzyme in the denovo purine synthesis pathway, inosine monophosphate dehydrogenase (Gummert, J.F. et al., 1999, J. Am. Soc. Nephrol. 10:1366-1380). Mycophenolate mofetil was notable in the current studies for its complete lack of effect on CD154 expression by T cells, consistent with an antagonistic function at late stages of T cell activation. Accordingly, use of CD 154 mAb with mycophenolate mofetil and/or rapamycin may be particularly efficacious in the management of transplant recipients.

Methylprednisolone resulted in almost complete inhibition of CD154 induction in mouse CD4+ T cells, consistent with a single previous report of the effects of dexamethasone on human CD4+ T cells in which activation was induced by a non-TCR-dependent mechanism (PMA/ionomycin) (Bischof, F. et al., 1998, Cell Immunol 187:3844). Glucocorticoids enter target cells, bind cytoplasmic receptors and form complexes which translocate to the nucleus and bind specific response elements in the promoters of target genes (Gummert, J.F. et al., 1999, J. Am. Soc. Nephrol. 10:1366-1380). In addition to inhibiting late events in T cell activation including proliferation and cytokine production, glucocorticoids block early events such as the activation of transcription factors AP-1 and NF-κB. Indeed, ligand-bound glucocorticoid receptors bind ReIA and NF-κB p50 subunits

in vitro (Epinat, J.C. et al., 1999, Oncogene 18:6896-6909), affecting the transactivation potential of RelA/p65, and also enhance transcription of the NF-κB antagonist IκB-α. Thus, we considered whether NF-kB activation was required for CD154 expression.

Compared with the responses in normal CD3 mAb-activated CD4+ T cells, CD154 induction was consistently inhibited by about 65% in NF-kB/p50 KG cells (Figure 12) (p<0.02). Since activation of NF-KB requires the signal-coupled phosphorylation and proteolysis of IκB-α through the 26S proteasome (Lin, Y.C., et al., 1995, Proc Natl Acad Sci USA 92:552-6; Traenckner, E.B. et al., 1995, EMBO J 14:2876-83), we also tested the effects of 2 different types of proteasome inhibitors on CD154 induction by activated T 10 cells (Fig. 12). Lactacystin irreversibly blocks proteasome activity by acylating a threonine residue in the active site of the mammalian proteasome subunit X, whereas dipeptide boronates, such as MG-273, act by irreversible inhibition of proteasomal chymotryptic activity (Grisham, M.B. et al., 1999, Methods Enzymol 300:345-63). Use of either agent in vitro significantly decreased CD154 expression by CD3-stimulated CD4+ cells (p<0.05) 15 (Figure 12). Together, these data indicate that NF-kB activation is required for optimal induction of CD154 by activated T cells.

To assess the validity of these in vitro data as guides to the in vivo efficacy of CD154 mAb therapy, CD154 mAb-induced cardiac allograft survival in normal vs. NFκB/p50 KO mice, as well as in wild-type mice treated with a proteasome inhibitor; all 20 experiments involved the same H2d->H2b MHC disparity. As anticipated from the in vitro data using spleen cells from NF-κB/p50 KO mice was evaluated, the efficacy of CD154 mAb therapy in vivo was abrogated in NF-κB/p50 KG mice (p<0.001) (Figure 13). Moreover, administration of a proteasome inhibitor to wild-type mice also blocked the effects of CD154 mAb therapy in vivo (p<0.001) (Figure 13). In this light, even agents 25 such as the calcineurin-inhibitors, cyclosporin A and FK-506, which are known to inhibit NF-KB activation (Epinat, J.C. et al., 1999, Oncogene 18:6896-6909), may exert their effects on CD154 induction and CD154 mAb therapy as consequences of NF-kB inhibition.

The first conclusion of these in vitro and in vivo studies is that CD154 mAb fails to prolong allograft survival under conditions that diminish upregulation of CD154 upon 30 CD4+ T cell activation. Indeed, the flow cytometric assay of CD154 expression provides a ready approach to evaluating potential clinical utility of CD154 mAb in combination with other immunosuppressive agents, and predicting those combinations which are likely to be successful (e.g. CD154 mAb plus rapamycin or mycophenolate mofetil) or not (e.g. CD154 mAb plus cyclosporin A or glucocorticoids). These studies further suggest that agents 35 blocking early stages of T cell activation, thereby suppressing CDI 54 induction, will not be useful adjuncts to CD154 mAb therapy.

A second conclusion from the data is that NP-κB is required for CD154 induction, a finding that has not been previously described. Interestingly, the phenotype of NF-κB/p50 KO is one of moderate immunodeficiency associated with defective antibody responses (Sha, W.C. et al.,1995, Cell, <u>80</u>:321-30). These data indicate, therefore, that much of this phenotype may relate to defects in CD154 induction, which is key to development of B cell responses.

In addition, these findings provide an alternate interpretation for recent experimental data emphasizing an important role for CD40/CD154 interactions in the development of apoptosis in vivo, and which have suggested that concomitant use of cyclosporin A prevents tolerance induction by impairing activation-induced cell death (Li, X.C. et al., 1999, J. Immunol 163:2500-2 507; Wells. A.D. et al., 1999 Nat Med 5:1303-7). These data are more consistent with models suggesting that CD154mAb functions by stimulating CD154-dependent events (Blair, P.J. et al., 2000, J Exp Med 191:651-660), at least transiently. Further, these data demonstrate, for the first time, that NF-κB is required for CD154 induction.

8. <u>EXAMPLE</u>: Post-Transplantation Stat and SOCS Levels and Their Involvement in Immune Reduction and Tolerance Induction

This Example presents the first data that has been generated on the post-20 transplantation levels of Stat and SOCS members post-transplantation. Using a vascularized cardiac transplant model, the levels of Stats (Stats 1-6) and SOCS (inhibitors of Stats) were measured during rejection and tolerance (via CD40L antibody treatment). During rejection, a rapid upregulation of Stat1, Stat2 and Stat3 mRNA was observed, followed by upregulation of these Stats in the animals' own hearts, indicating a systemic expression pattern. In contrast, RNA and protein analysis demonstrate that changes in Stat4 and Stat6 expression are confined to the transplant tissue. Specifically, Stat4 mRNA, however, was upregulated only later, and only in the transplant heart. Likewise, Stat6 mRNA was upregulated locally in the transplant tissue. Treatment with CD40L monoclonal antibody (MR-1) resulted in the downregulation of all the Stats, with the exception of Stat6, 30 which was upregulated upon MR-1 treatment. Thus, reduction of immune rejection, via induction of tolerance, was accompanied by a downregulation of Stat4 and an increase in Stat6 levels. Cardiac transplants in Stat6 knockout mice were rejected despite treatment with MR-1. These results indicate that an immune deviation from TH1 (mediated by IL-12 and Stat4) to TH2 (mediated by IL-4 and Stat6) is a prerequisite in the induction of tolerance, and that Stat4 and Stat6, at a minimum, are citrical signals in graft survival.

Further, these results indicate that Stat6, at a minimum, is a prerequisite in the induction of tolerance.

MATERIALS & METHODS

5 Cloning by RT-PCR:

5 μg of IL-6 (Pharmingen, San Diego, CA) was injected intravenously to a B6/129 strain female mouse. The heart, liver and spleen from the mouse was collected 1 hr after the intravenous injection of IL-6. Total RNA from these three organs were prepared using the acid-guanidine thiocyanate-phenol-chloroform method (Promega, Madison, WI). The RNA was then dissolved in water, quantitated, and a portion of the RNA was combined in equal amounts, and 1 μg of the combined RNA was used in a 50 μl reaction volume for the synthesis of first-strand cDNA. The ProStar Ultra HF RT-PCR System (Stratagene, La Jolla, CA) reagents were used both for the first-strand cDNA generation and for the subsequent steps during the amplification of the cDNA template. The following upstream and downstream primers were used:

```
5'-GAACTTTCAGCTGTTACTTTCC-3' (SEQ ID NO:35)
   Stat1:
           5'-CTGTGCTCATCATACTGTC-3' (SEQ ID NO:36)
           5'-GTGTTACAGTCACTCCCACTG-3 (SEQ ID NO:37)
   Stat2:
           5'-CCTCAGGCAAATCTGACTCTG-3' (SEQ ID NO:38)
20
           5'-GAAAGTACTGTAGGCCCGAG-3' (SEQ ID NO:39)
   Stat3:
           5'-CTGGAACCACAAGTTAGGAG-3' (SEQ ID NO:40)
   Stat4:
           5'-GAAGTGAGATTCCACTCTGTAG-3' (SEQ ID NO:41)
          5'-CACTCTCCAGTTTCATCTGC-3' (SEQ ID NO:42)
   Stat5A: 5'-CGAAAGCAGTTGACGGATACG-3' (SEQ ID NO:43)
25
          5'-CTCCAACTTAGTTGCCTAAACC-3'
                                        (SEQ ID NO:44)
   Stat5B: 5'-CAAGCCGTTAGAAGCAGGAG-3' (SEQ ID NO:45)
          5'CCATGGTTCACAACCTACAG-3' (SEQ ID NO:46)
           5'-GATGAGGCTTTCCGGAGTCAC-3'
   Stat6:
                                        (SEQ ID NO:47)
          5'-CAGTTGTATCACATTCGAGC-3'
                                      (SEQ ID NO:48)
  SOCS1:
           5'-CTGTGCCGCAGCATTAAGTG-3' (SEQ ID NO:49)
          5'-GTTTATTACCTAAACTGGCTG-3'
                                        (SEQ ID NO:50)
   SOCS2: 5'-CCAGGTATAAGTATTTCTCTC-3'
                                        (SEQ ID NO:51)
          5'-GGCCATTTGATCTTGAGCAGC-3'
                                        (SEQ ID NO:52)
  SOCS3:
           5'-GCAGATTGGCTTCTTCCTCAG-3'
                                        (SEQ ID NO:53)
          5'-GGCATTTAAGGCGAGTCTCC-3' (SEO ID NO:54)
  SOCS5:
           5'-GGAGCTTACTCGCAGTAGGCTC-3'
                                         (SEQ ID NO:55)
          5'-GTAGGAGTCTCTCCGTGCAAGC-3'
                                         (SEQ ID NO:56)
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CIS: 5'-CCAACTCTGACTGAGCCAGG-3' (SEQ ID NO:57) 5'-CATCCATACGCAGGTGGATG-3' (SEQ ID NO:58)

Amplification reactions included 5μl 10x Ultra HF PCR buffer, 1 μl dNTP (40 mM) mix, 0.5 μl upstream primer (approximately 500 ng/ul), 0.5 μl downstream primer (~500 ng/μl), 1 μl first-strand cDNA reaction, 41 μl H₂O, 1 μl Pfu Turbo DNA polymerase (2.5 U/μl). The same PCR program was used for all the amplifications: samples were heated for 1 min at 95 °C, followed by 40 cycles of 1 min at 95 °C, 1 min at 58 °C, 2 min at 68 °C, and final extension at 68 °C for 5 min. The PCR samples were then loaded onto agarose or acrylamide gels, the cDNA fragments were isolated and cloned directly into SfrI cut PCR-Script (Stratagene). Sequence analysis of the plasmids were performed by Tufts Core Facility (Boston, MA). The length of the cloned fragments were as follows: Stat1 (334 bp); Stat2 (694 bp); Stat3 (373 bp); Stat4 (442 bp); stat 5A (634 bp); Stat 5B (458 bp); Stat6 (894 bp); SOCS1 (381 bp); SOCS 2 (266 bp); SOCS3 (381 bp); SOCS 5 (558 bp); and CIS (688 bp).

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Cardiac Transplantation:

Heterotopic cardiac allografting was performed with anastomoses to the abdominal aorta and vena cava, using BALB/c donors and B6/129 wild-type or Stat-6 KO mice as recipients (n = 6/group). See, Hancock et al., 1998, Nature Med. 4:1392-1396. Recipients were intravenously administered donor specific transfusion ("DST"; 5 x 10⁶ splenic mononuclear cells) ("DST + IgG") plus 250 μg hamster anti-mouse CD40L mAb (CD154 mAb; BioExpress, West Lebanon, NH) and DST ("DST + MR-1") or control hamster IgG ("DST + IgG") at the time of transplantation. Graft survival was monitored by daily palpation and rejection was confirmed by laparotomy and histologic evaluation.

25

RNA isolations and Northern blot analysis:

Total RNA from native or transplanted hearts of Balb/c or B6/129 mice was prepared using the acid-guanidine thiocyanate-phenol-chloroform method (Chomezynski, P. and Sacchi, N., 1987, Anal. Biochem. 162:156-159). 25 μg of RNA was loaded onto each lane of 1.2 % agarose-formaldehyde gels. The 0.24 kB RNA ladder (GIBCO-BRL, Rockville, MD) was used as a size control. After electrophoresis the RNA was blotted overnight onto Nytran Supercharge membranes (Schleicher & Schuell, Keene, NH) with 20 x SSC and cross-linked onto the membranes by irradiation with UV light using a Stratalinker (Stratagene). 32P-labeled probes were prepared by using the Multiprime DNA labelling system and 32 P-dCTP (both from Amersham Pharmacia Biotech, Piscataway, NJ). Hybridizations with the 32P-labeled probes were done at 68°C in roller bottles using

ExpressHyb Solution (Clontech Laboratories, Palo Alto, CA). For re-use, the membranes were deprobed in 0.5 % SDS at 95-100°C and exposed to film to assure complete removal of previous hybridization signals.

The nucleotide sequences of the probes utilized are presented below:

5 <u>Stat1</u> (SEQ ID NO:59):

gaactttcagctgttactttcccagatattattcgcaactacaaagtcatggctgccgagaacataccagagaatcccctgaa gtatctgtaccccaatattgacaaagaccacgcetttgggaagtattattccagaccaaaggaagcaccagaaccgatgga gcttgacgaccctaagcgaactggatacatcaagactgagttgatttctgtgtctgaagtccaccettctagacttcagacca cagacaacctgcttccagaggaggtttgatgagatgtcccggatagtgggccccgaatttgacagtatgatga gcacag

10 <u>Stat2</u> (SEQ ID NO:60):

Stat3 (SEQ ID NO:61):

gaaagtactgtaggcccgagagccaggagcacccgaagccgacccaggtagtgctgccccgtacctgaagaccaag ttcatctgtgtgacaccaacgacctgcagcaataccattgacctgccgatgtcccccgcactttagattcattgatgcagttt ggaaataacggtgaaggtgctgagccctcagcaggagggcagtttgagtcgctcacgtttgacatggatctgacctcgga gtgtgctacctcccccatgtgaggagctgaaaccagaagctgcagagacgtgacttgagacacctgcccgtgctccacccctaagcagccgaaccccatatcgtctgaaactcctaactttgtggttccag

<u>Stat4</u> (SEQ ID NO:62):

gaagtgagattccactctgtagaaccctacaacaaagggagactgtcggctctggccttcgctgacatcctgcgagactac aaggttatcatggctgaaaacatccctgaaaaccctctgaagtacctctaccctgacattcccaaagacaaagcctttggca aacactacagctcccagccgtgcgaagtctcaagaccaaccgaacgggagagacaagggttacgtcccctctgtttttatc cccatttcaacaatccgaagcgattccacggagccacaatctccttcagaccttctccccatgtctccaagtgcatatgctgt gctgagagaaaacctgagcccaacgacaattgaaactgcaatgaattccccatattctgctgaatgacggtgcaaacgga cactttaaagaaggaagcagatgaaactggagagtg

<u>Stat 5A</u> (SEQ ID NO:63):

35 <u>Stat 5B</u> (SEQ ID NO:64):

caagccaagccgttagaagcaggagccctggccagtgcctggtcacggagctgagctgtgtttagatgtgttggctgct gcgtggtgaaggaagacccgtctccagaaaagcaatttaggcaaaagggattccgtttgatggcagagtcccagtgcta gaaaggtagcgaaggtggacagcttacagtctcaactcatttcgtcgtaaatgtcctcgtaacgacattgattcttctacctg

15

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Stat6 (SEQ ID NO:65):

SOCS1 (SEQ ID NO:66):

SOCS2 (SEQ ID NO:67):

SOCS3 (SEQ ID NO:68):

<u>SOCS 5</u> (SEQ ID NO:69):

ggagettaetegeagtaggetetegetettetaateaatggataaagtggggaaaatgtggaacaacttaaaatacagatge cagaatetetteagceacgagggaggaagcegtaatgagaacgtggaggatgaacceaacagatgteegtetgteaaag agaaaagcateagtetgggagaggeageteeccagcaagaggaggteeettaaggagaaaatgttgeettacagetgggactgagecetteeaagagcateegagageteeteaagtggtgaaateagcateg agaaagacagtgactegggtgccaccccaggaacgaggettgcacggagagacteetac

20

25

30

<u>CIS</u> (SEQ ID NO:70):

Western blotting:

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Hearts obtained from mice were frozen in liquid nitrogen and homogenized with Tissue Tearor (model 985370, Biospec Products, Inc.) in 800 µl lysis buffer containing 4% SDS, 125 mM Tris-HCl, pH 6.8, and protease inhibitors Antipain, Benzamidin, BeStatin, Chymostatin, Leupeptin, Pefabloc C, Pepstatin A, PMSF, TLCK, TPCK. The protease inhibitors were prepared and used as suggested by the manufacturer, Roche Molecular Biochemicals, Indianapolis, IN. Following homogenization, the DNA in the samples was sheared by sonication for 30 sec at 5 Watts (RMS) output power, with a Virtis sonicator (model Virsonic 60, Virtis Company, Gardiner, NY). Samples were then heated at 95°C for 10 minutes and centrifuged at 14,000 x g for 30 minutes, to get rid of particulate material. Protein concentrations in the extracts were determined using DC Protein Assay (Bio-Rad Laboratories, Hercules, CA). The protein concentration in all the samples was equalized to 10 mg/ml by adding lysis buffer, and 1 vol. 2 x Sample buffer (20 % Glycerol, 0.005 % Bromophenol blue) was added to yield samples with a protein concentration of 5 mg/ml. The samples were kept at '80°C until use. Prior to loading onto 10% SDS-polvacrylamide gels, 2.5 % $\beta\text{-mercaptoethanol}$ was added to every sample (100 $\mu\text{g})$ followed by heating to 95°C for 10 min. After electrophoresis, proteins were transferred onto Immobilon-P membranes (Millipore Corporation, Bedford, MA 01730) for 2.5 hrs at 100 V, using a Hoefer Transphor Electrophoresis unit. The Transfer buffer consisted of 12 mM Tris base, 96 mM Glycine, and 200 ml Methanol in 1 liter. After the transfer, the membranes were placed in roller bottles and rotated for 1 hr (at 4°C) in a Blocking solution (1 x PBS + 0.1 % Tween-20 + 5 % non-fat dry milk + 0.02 % sodium azide), the membranes were rinsed once with the Rinse solution (1 x PBS + 0.1 % Tween-20) and rolled overnight with the primary antibody in the Blocking solution. After washing times in the Rinse solution (15, 10, 5 minutes), the blots were incubated for 1 hour with the secondary antibody conjugated to horseradish peroxidase, followed by another set of washes with the Rinse solution and one 35 final wash in 1 x TBS (10 mM Tris-HCl, pH 8.0, 150 mM NaCl). Chemiluminescence was performed by using the Luminol Reagent (Santa Cruz Biotechnology, Inc., Santa Cruz, CA)

and detection was with Biomax MR film (Eastman Kodak Company, Rochester, NY). All the antibodies were used at 1:1000 dilution. Stat1, phospo-Stat1, Stat3, phospho-Stat3, Stat4, and Stat5A were from UpState Biotechnology (Lake Placid, NY). Stat2, Stat6, HSC70, and the secondary antibodies were from Santa Cruz Biotechnology.

5

Quantification of RNA and Protein

Quantification of the RNA and protein bands on the Kodak Biomax MR film was performed on a Macintosh Performa 6300CD computer using the public domain NIH Image program (developed at the U.S. National Institutes of Health and available on the Internet at http://rsb.info.nih-image/).

RESULTS

STAT and SOCS RNA levels in heart allografts during the 5 days following transplantation

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Hearts from Balb/c mice were transplanted into B6/129 female mice using a vascularized mouse cardiac allograft model. The heart transplant resulted with animals having two hearts: their own hearts and a second functional heart grafted adjacent to the kidneys. To determine the kinetics of Stat and SOCS RNA expression following cardiac allograft surgery, total RNA was prepared from the animals' own hearts (native), the transplanted hearts and from the hearts of healthy controls (designated as "N", "T", and "C", respectively) one, two, three, and five days post-transplantation. RNA (25 μ g/lane) was then electrophoresed on three identical gels, transferred to Nytran Supercharge membranes and the membranes were hybridized with Stat probes, SOCS probes, and a GAPDH probe as a control (Figures 14-16).

25

As shown in Figures 14A and 14B, increases of approximately 5-fold in Stat1, Stat2 and Stat3 RNA levels were detected in the transplanted hearts relative to the control hearts as early as one day post-transplantation, and were more evident by the second day. During the first two days, the increase appeared to be localized to only the transplanted tissue. By the third day, and much more strongly by the fifth day, higher levels of Stat1, 2, and 3 RNA were also detected in the native hearts.

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Stat4 RNA levels, almost undetectable the first three days post-transplantation, increased significantly (approximately 60-fold) in the transplanted hearts at around day five post-transplantation, but not earlier. Stat5A RNA levels did not change in the native and transplanted hearts during the five days following the transplant surgery. An increase in Stat6 RNA expression was detected in transplanted hearts as early as day one post-

35 Stat6 RNA expression was detected in transplanted hearts as early as day one posttransplantation (approximately 2-fold), and remained at approximately the same level

during the five days following transplantation. The fact that increases in Stat4 and Stat6 RNA expression were only detected in the transplanted hearts by day five posttransplantation suggests that the up-regulation of the expression of these two Stats was localized to the transplanted hearts (Figures 14A, 14B).

An increase in SOCS1 RNA expression levels in the transplanted hearts began to increase by day two post-transplantation and by day five post-transplantation SOCS1 RNA expression in the transplanted hearts had increased approximately 300-fold (Figure 15). Between days 1 and 3, there was an approximately 6-fold difference in such SOCS1 RNA levels, and between days 3 and 5, there was an approximately 25-fold difference in such 10 SOCS1 RNA levels. At approximately 38-fold, the SOCS1 RNA increase in native hearts was much less pronounced (Figure 15).

A considerably high level of SOCS3 RNA expression was detected in the transplanted hearts as early as one day post-transplantation (approximately 100-fold increase), and reached an approximately 230-fold increase at five days post-transplantation. 15 (Figure 15). However, SOCS3 RNA expression remained low in native hearts, and was hardly detectable in the native hearts five days following the transplant surgery. As shown in Figure 16, Stat4 and SOCS3 RNA expression patterns were found to be quite similar to each other, both of them being expressed only in the transplanted hearts, and both of them being significantly up-regulated at approximately day 5 post-transplantation..

Two species of SOCS5 RNA were detected (4.4 kb and 3.8 kb) in the naive and transplanted hearts. The shorter species 3.8 kb SOCS5 RNA species was much more abundant in the transplanted hearts than in naive hearts or in control hearts. The level of CIS RNA expression detected in naive hearts remained almost unchanged relative to the control hearts, while a lower level of CIS RNA expression was detected in the transplanted 25 hearts relative to the naive hearts or control hearts (Figure 15).

STAT Protein Levels in Heart Allografts and Native Hearts During the 5 Days Post-transplantation

To determine the effect of cardiac allografts on Stat and SOCS protein expression, protein extracts were prepared from transplanted and native hearts at day one, day two, day three, and day five, post-transplantation. In the transplanted hearts an increase in Stat1 protein levels relative to the control hearts was detected as early as one day posttransplantation, increased steadily, and by day five post-transplantation Stat1 protein levels were approximately 17-fold higher (Figure 17). In the native hearts an increase in Stat1 protein levels began increasing at day two post-transplantation and by day five posttransplantation Stat1 levels were 15-fold higher.

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An increase in Stat2 protein expression levels in the transplanted hearts was initially detected three days post-transplantation, and by day five post-transplantation had increased sharply by approximately 27-fold. In native hearts an increase in Stat2 protein levels was detected by day three post-transplantation and by day five post-transplantation a 10-fold increase in the levels of Stat2 protein was detected.

An increase in Stat3 protein levels in the transplanted hearts was detected as early as day two post-transplantation and by day five post-transplantation the levels of Stat3 protein in the transplanted hearts exhibited an approximately 3.5-fold increase. In the native hearts a 2-fold increase in the levels of Stat3 protein was detectable only by day five post-transplantation.

Stat4 levels began increasing around one day post-transplantation in the transplanted heart, with a dramatic increase (approximately 16-fold) between day 2 and day 5 post-transplantation. Stat4 was detected only in the transplant tissue, not in the native heart tissue (Figure 17).

Stat5A protein levels did not change in the transplanted or native hearts relative to the control hearts.

Stat6 is normally made at a low level in hearts (see, e.g., Figure 17, control lanes). As was observed with Stat4, Stat6 expression, however, only increased (aproximately 1.5-3.5-fold) in transplanted hearts.

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Phosphorylated forms of Stat1 and Stat3 during graft rejection

To determine whether the levels of phosphorylated Stat1 and phosphorylated Stat3 protein parallel that of the newly synthesized Stat1 and Stat3 in the native and transplanted hearts following cardiac allograft surgery, the levels of these Stats and their phosphorylated forms were analyzed (Figure 18). As shown in Figure 18, an increase in phosphorylated Stat1 protein levels paralleled the increase in the total Stat1 protein levels detected both in native and transplanted hearts on days three and five post-transplantation. In contrast, the level of phosphorylated Stat3 protein detected in native and transplanted hearts over the five day post-transplantation period did not parallel the increase in the total Stat3 protein levels detected during this period. Ijn particular, although there was a steady increase of total Stat3 over the five day period, there was no major change in the levels of phosphorylated Stat3 protein detected over this same period, with the exception of the appearance of a slightly higher molecular weight form of phosphorylated Stat3 protein.

35 The Effect of Anti-CD4OL Monoclonal Antibody MR-1 on the Stat and SOCS RNA Levels in 5-day Post-Transplant Hearts

To determine how Stat and SOCS RNA expression levels are affected in cardiac allografts when tolerance has been induced, six mice having undergone cardiac transplant surgery were divided into three groups. One group of mice were intravenously administered DST ("donor specific transfusion") + MR-1 anti-CD40L antibody which has been shown to induce tolerance), on the day of the transplant surgery. The second group of mice were intravenously administered DST + IgG as control on the day of the transplant surgery and the third group received no treatment. The animals were sacrificed five days post-transplantation and total RNA was prepared from the animals' own hearts (native), the transplanted hearts and from the hearts of healthy controls (designated as "N", "T", and 10 "C", respectively). The RNA (25 µg /lane) was electrophoresed on four identical gels and the Northern blots were hybridized to Stat probes, SOCS probes and a GAPDH probe as a control. (Figures 19, 20)

Stat1, Stat2, and Stat3 mRNA were found to be expressed at some level in the normal heart (see control lanes of the no treatment group). Five days post-transplantation, 15 the transplant allograft tissue exhibited a dramatic upregulation of Stat1, Stat2 (approximately 45-fold), and Stat3 (approximately 6-fold) mRNAs. This upregulation was found to be systemic in nature in that the increases were also observed in the native heart tissue (approximately 43-fold, 48-fold, and 5.5-fold, respectively). Treatment with DST + IgG caused minimal changes n the transplant Stat RNA levels, but the single does of DST + 20 MR-1 resulted in lower levels of Stat1, Stat2, and Stat3 mRNAs (62%, 76%, and 50% lower than the animals that received no treatment).

Stat4 mRNA was found only in the transplanted heart tissue, as a local signal, and was not detected in the control or native hearts (Figure 19; see the control and native lanes of the no treatment group). Stat4 mRNA was significantly reduced (approximately 6-fold) in transplants obtained from MR-1-treated animals.

Only minimal differences in Stat 5A mRNA expression in transplant tissue of the group that received no treatment and the controls (approximately 1.4-fold increase) were observed. MR-1 treatment, however, lowered the Stat 5 A expression level by 70%.. MR-1 treatment resulted in Stat 5A levels that were 60% lower than control heart levels. Stat 5B 30 mRNA levels were too low to be detected (data not shown).

Stat6 mRNA is normally expressed at some level in the heart, and five days posttransplantation, Stat6 mRNA levels doubled only in the transplanted, not native, hearts. Upon MR-1 treatment, however, Stat6 mRNA levels were downregulated to pretransplantation (control) levels.

The same membranes were used to determine the mRNA expression levels of SOCS1, SOCS 2, SOC, and CIS (Figure 20). SOCS1 and SOCS3 mRNA levels were

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almost undetectable in control hearts. SOCS1 and SOCS3 mRNA levels were increased five days post-transplantation in transplant tissue in both DST + IgG treated mice and untreated mice (some low level of SOCS1 expression was also detected in the native hearts of the untreated groups). Treatment with DST + MR-1 resulted in a 93% and 73% reduction in the level of SOCS1 and SOCS3 mRNA expression, respectively, relative to that in transplanted hearts of untreated mice.

SOCS 2 RNA levels remained unchanged in the control, native, and transplanted hearts in the three experimental groups. Interestingly, CIS levels were 50% higher in the transplanted hearts of mice, regardless of treatment.

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The Effect of Anti-CD40L Monoclonal Antibody MR-1 on Stat Protein Levels in 5-day and 7-day Post-Transplant Hearts

To determine how Stat protein levels are affected in cardiac allografts when tolerance has been induced, the day of the surgery mice were intravenously administered a single dose of either DST + IgG or DST + MR-1, and the Stat protein levels were analyzed five and seven days post-transplantation. As shown in Figure 21, at day 5 post-transplantation thee were mnimal changes in the Stat levels between IgG and MR-1 treated animals, but a day 7 post-transplantation, there was significant changes in such levels.

Stat1 and Stat2 levels were rduced 64% and 52%, respectively, in native hearts of MR-1 treated animals. Stat3 levels were reduced in the transplant tissue by 29%. The most dramatic change at day 7 post-transplantation was the amount of Stat4 reduction (approximately 63%) in the transplanted hearts. Stat6, on the other hand, tended to increase about 2-fold upon MR-1 treatment.

25 Stat and Bax Protein Levels in Isografts and Allografts

The level of Stat1, Stat2, Stat3, Stat4, Stat5A, and Stat6 were compared in isografts and allografts to exclude the possibility that the observed changes in Stats were due to the surgery and the healing process that follows the surgery. As shown in Figure 22, between day 1 and day 2 post-transplantation, the isografts showed a small increase in different Stats. The low levels, however, of Stat1, Stat2, and Stat5A remained constant between days 2 and 7 post-transplantation, and Stat3 and Stat6 levels declined between these days. In allografts, on the other hand, all of the Stats reached high levels by day 5. Bax, an unrelated protein, showed a very different profile, with its levels increasing in both isografts and allografts.

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Transplantation in Stat6 Knockout Mice

Stat6 is activated by IL-4 and IL-13, and Stat6 knock-out animals have been shown to be deective in TH2 differentiation (Kaplan, M.H. et al., 1996, Imm. 4:313-319; Takeda, K. et al., 1996, Nature 380:627-630; and Shimoda, K. et al., 1996, Nature 380:630-633). The results presented herein indicate that induction of tolerance, by causing lower Stat4 and higher Stat6 levels, shifts the differentiation of T cells toward the TH2 lineage.

To assess the importance of Stat6 and the TH2 response in MR-1 mediated tolerance induction, hearts from B6/129 background mice were transplanted into three Stat6 (-/-) mice with a Balb/c background. The day of the surgery the animals were injected with MR-1 and splenocytes from the donor animals. In normal animals, such a transplant results in indefinite graft survival. Stat6 (-/-) animals, however, readily rejected the transplanted hearts at day 11. This result points out the importance of Stat6 in the induction of tolerance, e.g., the induction of tolerance via MR-1 adiministration.

The contents of all references, patents and published patent applications cited throughout this application are hereby incorporated by reference.

Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

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WHAT IS CLAIMED IS:

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1. A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of at least one of (i) to (iv), as follows:

- (i) Stat4 mRNA or Stat4 protein,
- (ii) Stat6 mRNA or Stat6 protein,
- (iii) SOCS1 mRNA or SOCS1 protein, or
- (iv) SOCS3 mRNA or SOCS3 protein,
- 10 present in a transplant sample from the subject.
 - 2. The method of Claim 1, comprising determining the amount of at least two of (i) to (iv) present in the transplant sample.
- 15 3. The method of Claim 1, comprising determining the amount of at least three of (i) to (iv) present in the transplant sample.
 - 4. The method of Claim 1, comprising determining the amount of (i) to (iv) present in the transplant sample.
 - 5. A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a transplant sample from the subject.
- 25 6. The method of Claim 5, further comprising determining the ratio of Stat4 to Stat6 amounts.
 - 7. The method of Claim 1, 2, 3, 4, 5, or 6, comprising comparing the amount or ratio determined to that present in a control sample.
 - 8. The method of Claim 7, wherein the control sample is a corresponding pretransplant subject sample.
- 9. The method of Claim 7, wherein the control sample is a subject blood 35 sample.

10. The method of Claim 7, wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is greater than, or the amount of Stat6 mRNA or protein in the transplant sample is less than that of the control sample, thereby indicating that acceptance of the transplant has not been induced or is not being maintained.

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- 11. The method of Claim 7, wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is less than, or the amount of Stat6 mRNA or protein in the transplant sample is equal to or greater than that of the control sample, thereby indicating that acceptance of the transplant has occurred, is being induced or is being maintained.
 - 12. The method of Claim 1 or 5, wherein the amount of mRNA is determined.
 - 13. The method of Claim 1 or 5, wherein the amount of protein is determined.

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- 14. The method of Claim 1 or 5, wherein the transplant is an allograft.
- 15. The method of Claim 1 or 5, wherein the transplant is a heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, intestine or cornea transplant.

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- 16. The method of Claim 1, further comprising assaying the transplant sample for evidence of cell injury.
- 17. A method for monitoring an autoimmune disorder in a subject mammal, comprising: determining the amount of at least one of (i) to (iv), as follows:
 - (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS protein, or
 - (iv) SOCS3 mRNA or SOCS protein,
- 30 present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder.
 - 18. The method of Claim 17, comprising determining the amount of at least two of (i) to (iv) present in the sample.

19. The method of Claim 17, comprising determining the amount of at least three of (i) to (iv) present in the sample.

- 20. The method of Claim 17, further comprising determining the amount of (i) to (iv) present in the sample.
- 21. A method for monitoring an autoimmune disorder in a subject mammal comprising: determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a sample from a subject mammal being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder.
 - 22. The method of Claim 21, further comprising determining the ratio of Stat4 to Stat6 amounts.
- 15 23. The method of Claim 17, 18, 19, 20, 21 or 22, further comprising comparing the amount or ratio determined to that present in a control sample.
 - 24. The method of Claim 23, wherein the control sample is a corresponding tissue not affected by the disorder.
 - 25. The method of Claim 23, wherein the control sample is a subject blood sample.
- The method of Claim 17, wherein the amount of Stat4, SOCS1, or SOCS3
 mRNA or protein in the sample is greater than, or the amount of Stat6 mRNA or protein in the sample is less than, that of the control sample, thereby indicating that the subject mammal exhibits or continues to exhibit the disorder.
- 27. The method of Claim 17, wherein the amount of Stat4, SOCS1, or SOCS3
 30 mRNA or protein in the sample is less than, or the amount of Stat6 mRNA or protein in the sample is equal to or greater than, that of the control sample, thereby indicating that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective.
 - 28. The method of Claim 17 or 21, wherein the amount of mRNA is determined.
 - 29. The method of Claim 17 or 21, wherein the amount of protein is determined.

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30. The method of Claim 17 or 21, wherein the autoimmune disorder is systemic lupus erythematosus, glomerulonephritis, rheumatoid arthritis, Wegener's granulomatosis, chronic active hepatitis, or vasculitis.

- 5 31. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:
 - (a) contacting an activated T cell sample with a test compound;
 - (b) determining the amount of at least one of (i) to (iv), as follows:
 - (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS1 protein, or
 - (iv) SOCS3 mRNA or SOCS3 protein, present in (a); and
 - (c) comparing the amount(s) in (a) to that/those present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the amount of (i), (iii) or (iv) is decreased, or the amount of (ii) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

- 20 32. The method of Claim 31, comprising determining the amount of at least two of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.
- 33. The method of Claim 31, further comprising determining the amount of at least three of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.
 - 34. The method of Claim 31, further comprising determining the amounts of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.
 - 35. The method of Claim 31, wherein the amount of mRNA is determined.
 - 36. The method of Claim 31, wherein the amount of protein is determined.
 - 37. The method of Claim 32, comprising:

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(a) contacting an activated T cell sample with a test compound;

- (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and
- (c) comparing the amounts in (a) to those present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

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- 38. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:
 - (a) contacting an activated T cell sample with a test compound;
 - (b) determining the ratio Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and
 - (c) comparing the ratio in (a) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

- 39. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:
 - (a) contacting a resting T cell sample, a T cell activator and a test compound;
 - (b) determining the amount of at least one of (i) to (iv), as follows:
 - (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS1 protein, or
 - (iv) SOCS3 mRNA or SOCS3 protein, present in (a); and
 - (c) comparing the amount(s) in (a) to that/those present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the amount of (i), (iii) or (iv) is decreased, or the amount of (ii) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

40. The method of Claim 39, comprising determining the amount of at least two of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

- 5 41. The method of Claim 39, further comprising determining the amount of at least three of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.
- 42. The method of Claim 39, comprising determining the amounts of (i) to (iv) 10 present in (a), and comparing the amounts to those present in the control sample.
 - 43. The method of Claim 39, wherein the resting T cell is a primary T cell.
- 15 44. The method of Claim 37, wherein the resting T cell is a T cell line.
 - 45. The method of Claim 39, wherein the T cell activator is a mitogen or a T cell receptor stimulatory factor.
- 20 46. The method of Claim 45, wherein the T cell activator is an anti-CD3 antibody.
 - 47. The method of Claim 40, comprising:
 - (a) contacting a resting T cell sample, a T cell activator and a test compound;
 - (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and
 - (c) comparing the amounts in (b) to those present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

48. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:

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- (a) contacting a resting T cell sample, a T cell activator and a test compound;
- (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and
- (c) comparing the ratio to in (a) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

- 49. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:
 - (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine;
 - (b) determining the amount of at least one of (i) to (iv), as follows:
 - (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS1 protein, or
 - (iv) SOCS3 mRNA or SOCS3 protein, present in (a); and
 - (c) comparing the amount(s) in (a) to that/those present in a corresponding control T cell sample that has not been contacted with the test compound,

so that if the amount of (i), (iii) or (iv) is decreased, or the amount of (ii) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

- 50. The method of Claim 49, comprising determining the amount of at least two of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.
- 51. The method of Claim 49, further comprising determining the amount of at least three of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

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52. The method of Claim 49, comprising determining the amounts of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

- 5 53. The method of Claim 49, wherein the T cell is a T cell line.
 - 54. The method of Claim 49, wherein the cytokine is IL-2, IL-4, IL-12, or IL-13.
 - 55. The method of Claim 50, comprising:

10 (a) contacting a resting T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine;

- (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and
- (c) comparing the amounts in (b) to those present in a corresponding control T cell sample that has not been contacted with the test compound,

so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

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- 56. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:
 - (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine;
 - (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and
 - (c) comparing the ratio to in (b) to that present in a corresponding control T cell sample that has not been contacted with the test compound,
- 30 so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.
- 57. A method for reducing immune rejection in a subject mammal, comprising: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to reduce the level of Stat4 mRNA or protein in the subject relative to

that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-κB activation in CD40L⁺ cells.

- 58. The method of Claim 57, wherein upon administration of the compound, the level of Stat6 mRNA or protein in the subject is maintained or increased relative to that observed in the subject in the absence of the compound.
- 59. A method for reducing immune rejection in a subject mammal, comprising: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to increase the level of Stat6 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF-κB activation in CD40L⁺ cells.
- 60. The method of Claim 57, 58 or 59, wherein the subject mammal has undergone a transplant.
 - 61. The method of Claim 60, wherein administering the compound induces tolerance in the subject mammal.
- 20 62. The method of Claim 57, 58 or 59, wherein the subject mammal being treated exhibits an autoimmune disorder.
 - 63. The method of Claim 57, 58 or 59, wherein the compound is a tyrosine kinase inhibitor.
 - 64. The method of Claim 63, wherein the tyrosine kinase inhibitor is a tyrphostin compound.
- 65. The method of Claim 63, wherein the tyrosine kinase inhibitor inhibits a Jak 30 enzyme.
 - 66. The method of Claim 65, wherein the Jak inhibited is Jak2.
- 67. The method of Claim 57, 58, or 59, wherein the compound is a Jak2 35 inhibitor.

gcc ttg	atco	tcg cta	agag acgt	ctgt gctg gg a M	ct a tg c tg t	ggtt gtag ct c	aacg ctgc ag t	t to t co gg t	gcac tttg ac g	tetg gttg aa c	tgt: aat: tt.c:	atata cccca ag ca	aac o agg o ag ci ln Lo	ctoga cccti tt ga	ctgtat acagtc tgttgg ac tca sp Ser	60 120 180 232
aaa Lys	ttc Phe	ctg Leu 15	gag Glu	cag Gln	gtt Val	cac His	cag Gln 20	ctt Leu	tat Tyr	gat Asp	gac Asp	agt Ser 25	ttt Phe	ccc Pro	atg Met	280
gaa Glu	acc Ile 30	aga Arg	cag Gln	tac Tyr	ctg Leu	gca Ala 35	cag Gln	tgg Trp	tta Leu	gaa Glu	aag Lys 40	caa Gln	gac Asp	tgg Trp	gag Glu	<i>3</i> 29
			aat Asn													376
ctg Leu	tca Ser	cag Gln	ctg Leu	gat Asp 65	gat Asp	caa Gln	tat Tyr	agt Ser	cgc Arg 70	ttt Phe	tct Ser	ttg Leu	gag Glu	aat Asn 75	aac Asn	424
ttc Phe	ttg Leu	cta Leu	cag Gln 80	cat His	aac Asn	ata Ile	agg Arg	aaa Lys 85	agc Ser	aag Lys	egt Arg	aat Asn	ctt Leu 90	cag Gln	gat Asp	472
aat Asn	ttt Phe	cag Gln 95	gaa Glu	gac Asp	cca Pro	atc Ile	cag Gln 100	atg Met	tct Ser	atg Met	atc Ile	att Ile 105	tac Tyr	agc Ser	tgt Cys	520
ctg Leu	aag Lys 110	gaa Glu	gaa Glu	agg Arg	aaa Lys	att Ile 115	ctg Leu	gaa Glu	aac Asn	gcc Ala	cag Gln 120	aga Arg	ttt Phe	aat Asn	cag Gln	568
gct Ala 125	cag Gln	tcg Ser	ggg Gly	aat Asn	att Ile 130	cag Gln	agc Ser	aca Thr	gtg Val	atg Met 135	tta Leu	gac Asp	aaa Lys	cag Gln	aaa Lys 140	616
gag Glu	ctt Leu	gac Asp	agt Ser	aaa Lys 145	gtc Val	aga Arg	aat Asn	gtg Val	aag Lys 150	gac Asp	aag Lys	gtt Val	atg Met	tgt Cys 155	ata Ile	664
gag Glu	cat His	gaa Glu	atc Ile 160	aag Lys	agc Ser	ctg Leu	gaa Glu	gat Asp 165	tta Leu	caa Gln	gat Asp	gaa Glu	tat Tyr 170	gac Asp	ttc Phe	712
aaa Lys	tgc Cys	aaa Lys 175	acc Thr	ttg Leu	cag Gln	aac Asn	aga Arg 180	gaa Glu	cac His	gag Glu	acc Thr	aat Asn 185	ggt Gly	gtg Val	gca Ala	760
aag Lys	agt Ser 190	gat Asp	cag Gln	aaa Lys	caa Gln	gaa Glu 195	cag Gln	ctg Leu	tta Leu	ctc Leu	aag Lys 200	aag Lys	atg Met	tat Tyr	tta Leu	808
atg Met 205	ctt Leu	gac Asp	aat Asn	aag Lys	aga Arg 210	aag Lys	gaa Glu	gta Val	gtt Val	cac His 215	aaa Lys	ata Ile	ata Ile	gag Glu	ttg Leu 220	856

Figure 1

2/48 ctg aat gtc act gaa ctt acc cag aat gcc ctg att aat gat gaa cta 904 Leu Asn Val Thr Glu Leu Thr Gln Asn Ala Leu Ile Asn Asp Glu Leu 225 230 gtg gag tgg aag cgg aga cag cag agc gcc tgt att ggg ggg ccg ccc 9,52 Val Glu Trp Lys Arg Arg Gln Gln Ser Ala Cys Ile Gly Gly Pro Pro aat get tge ttg gat cag etg cag aac tgg tte act ata gtt geg gag 1000 Asn Ala Cys Leu Asp Gln Leu Gln Asn Trp Phe Thr Ile Val Ala Glu agt ctg cag caa gtt cgg cag cag ctt aaa aag ttg gag gaa ttg gaa 1048 Ser Leu Gln Gln Val Arg Gln Gln Leu Lys Lys Leu Glu Glu Leu Glu 275 cag aaa tac acc tac gaa cat gac cct atc aca aaa aac aaa caa gtg 1096 Gln Lys Tyr Thr Tyr Glu His Asp Pro Ile Thr Lys Asn Lys Gln Val 300 tta tgg gac cgc acc ttc agt ctt ttc cag cag ctc att cag agc tcq 1144 Leu Trp Asp Arg Thr Phe Ser Leu Phe Gln Gln Leu Ile Gln Ser Ser 310 ttt gtg gtg gaa aga cag ccc tge atg cca acg cac cct cag agg ccg Phe Val Val Glu Arg Gln Pro Cys Met Pro Thr His Pro Gln Arg Pro 1192 325 ctg gtc ttg aag aca ggg gtc cag ttc act gtg aag ttg aga ctg ttg 1240 Leu Val Leu Lys Thr Gly Val Gln Phe Thr Val Lys Leu Arg Leu Leu 340 gtg aaa ttg caa gag ctg aat tat aat ttg aaa gtc aaa gtc tta ttt 1288 Val Lys Leu Gln Glu Leu Asn Tyr Asn Leu Lys Val Lys Val Leu Phe 350 355 gat aaa gat gtg aat gag aga aat aca gta aaa gga ttt agg aag ttc 1336 Asp Lys Asp Val Asn Glu Arg Asn Thr Val Lys Gly Phe Arg Lys Phe 370 aac att ttg ggc acg cac aca aaa gtg atg aac atg gag gag tcc acc 1384 Asn Ile Leu Gly Thr His Thr Lys Val Met Asn Met Glu Glu Ser Thr 385 aat ggc agt ctg gcg gct gaa ttt cgg cac ctg caa ttg aaa gaa cag 1432 Asn Gly Ser Leu Ala Ala Glu Phe Arg His Leu Gln Leu Lys Glu Gln 400 405 410 aaa aat get gge ace aga acg aat gag ggt cet etc ate gtt act gaa 1480 Lys Asn Ala Gly Thr Arg Thr Asn Glu Gly Pro Leu Ile Val Thr Glu 415 gag ett cac tee ett agt ttt gaa ace caa ttg tge cag eet ggt ttg 1528 Glu Leu His Ser Leu Ser Phe Glu Thr Gln Leu Cys Gln Pro Gly Leu 430 gta att gac etc gag acg acc tet etg ecc gtt gtg gtg ate tec aac 1576 Val Ile Asp Leu Glu Thr Thr Ser Leu Pro Val Val Val Ile Ser Asn

Figure 1 (cont'd)

455

450

gt Va	c ago l Sei	c cag	g cto n Leu	ccg Pro 465	Ser	ggt Gly	tgg Trp	gcc Ala	tcc Ser 470	atc Ile	ctt Leu	tgg Trp	tac Tyr	aac Asn 475	atg Met	1624
cte Lei	g gto ı Val	g gcg Ala	gaa Glu 480	ccc Pro	agg Arg	aat Asn	ctg Leu	tcc Ser 485	ttc Phe	ttc Phe	ctg Leu	act Thr	cca Pro 490	cca Pro	tgt Cys	1672
gca Ala	a cga a Arg	tgg Trp 495	Ala	cag Gln	ctt Leu	tca Ser	gaa Glu 500	gtg Val	ctg Leu	agt Ser	tgg Trp	cag Gln 505	ttt Phe	tct Ser	tct Ser	1720
gto Val	acc Thr 510	Lys	aga Arg	ggt Gly	ctc Leu	aat Asn 515	gtg Val	gac Asp	cag Gln	ctg Leu	aac Asn 520	atg Met	ttg Leu	gga Gly	gag Glu	1768
aag Lys 525	Leu	ctt Leu	ggt Gly	cct Pro	aac Asn 530	gcc Ala	agc Ser	ccc Pro	gat Asp	ggt Gly 535	ctc Leu	att Ile	ccg Pro	tgg Trp	acg Thr 540	1816
agg Arg	ttt Phe	tgt Cys	aag Lys	gaa Glu 545	aat Asn	ata Ile	aat Asn	gat Asp	aaa Lys 550	aat Asn	ttt Phe	ccc Pro	ttc Phe	tgg Trp 555	ctt Leu	1864
tgg Trp	att Ile	gaa Glu	agc Ser 560	atc Ile	cta Leu	gaa Glu	ctc Leu	att Ile 565	aaa Lys	aaa Lys	cac His	ctg Leu	ctc Leu 570	cct Pro	ctc Leu	1912
tgg Trp	aat Asn	gat Asp 575	ggg Gly	tgc Cys	atc Ile	atg Met	ggс Gly 580	ttc Phe	atc Ile	agc Ser	aag Lys	gag Glu 585	cga Arg	gag Glu	cgt Arg	1960
Ala	Leu 590	Leu	Lys	gac Asp	Gln	Gln 595	Pro	Gly	Thr	Phe	Leu 600	Leu	Arg	Phe	Ser	2008
Glu 605	Ser	Ser	Arg	gaa Glu	Gly 610	Ala	Ile	Thr	Phe	Thr 615	Trp	Val	Glu	Arg	Ser 620	2056
Gln	Asn	Gly	Gly	gaa Glu 625	Pro	Asp	Phe	His	Ala 630	Val	Glu	Pro	Tyr	Thr 635	Lys	2104
aaa Lys	gaa Glu	ctt Leu	tct Ser 640	gct Ala	gtt Val	act Thr	ttc Phe	cct Pro 645	gac Asp	atc Ile	att Ile	cgc Arg	aat Asn 650	tac Tyr	aaa Lys	2152
Val	Met	Ala 655	Ala	gag Glu	Asn	Ile	Pro 660	Glu	Asn	Pro	Leu	Lys 665	Tyr	Leu	Tyr	2200
cca Pro	aat Asn 670	att Ile	gac Asp	aaa Lys	Asp :	cat His 7 675	gcc Ala	ttt Phe	gga Gly	aag Lys	tat Tyr 680	tac Tyr	tcc Ser	agg Arg	cca Pro	2248

Figure 1 (cont'd)

\SDOCID: <WO____0179555A2_I_>

			cca Pro													2296
			act Thr													2344
			aca Thr 720													2392
			cgg Arg		_			_	-		_	_	_	_		2440
aca Thr	_	tag *	agca	tga'a	tt t	tttt	cato	t to	tctg	idcds	caç	tttt	cct			2489
															gaaat	2549
															taact	2609
															aaggc	2669
															cagga	2729 2789
	-		_	_			-	_						_	actgc	2849
															taagt	2909
															ataca	2969
															tgtat	3029
															agaag	3089
															aagga	3149
															ctttc	3209
															caacc	3269
															catgt	3329 3389
															agata	3449
															gcata	3509
															gtact	3569
															ccttt	3629
															cttcc	3689
															caact	3749
															tttgt	3809 3869
															gaacc	3929
															aaaaa	3989
caata		-	-			- 5									-	4003

aagtc	gcgac	cagage	catt g	ıgaggg	icaca (gggac	tgcaa	a cc	ctaat	cag	agco		atg Met 1	60
gcg ca Ala Gl	g tgg n Trp	gaa a Glu Mo 5	tg ctg et Leu	cag Gln	Asn Le	it gae eu Asi .0	c ago o Ser	ccc Pro	ttt Phe	cag Gln 15	Asp	cag Gin		108
ctg ca Leu Hi	c cag s Gln 20	ctt ta Leu Ty	ac tcg /r Ser	cac His	agc ct Ser Le 25	c cto u Lei	g cct Pro	gtg Val	gac Asp	att Ile	cga Arg	ı cag ı Gln		156
tac tte Tyr Le 3	u Ala	gtc to Val Tr	g att p Ile	gaa (Glu / 40	gac ca Asp Gl	g aac n Asr	tgg Trp	cag Gln 45	Glu	gct Ala	gsa Ala	sti Leu		204
ggg agt Gly Sei 50	t gat c Asp	gat to Asp Se	c aag r Lys 55	gct a Ala 7	acc at Thr Me	g cta t Leu	ttc Phe 60	ttc Phe	cac His	ttc Phe	ttg Leu	gat Asp 65		252
cag cto Gln Lei	1 Asn	Tyr GI 7	u Cys 0	Gly F	Arg Cy.	s Ser 75	Gln	Asp	Pro	Glu	Ser 80	Leu		300
ttg ctg Leu Leu	GIn	His As 85	n Leu	Arg L	ys Phe 90	e Cys)	Arg	`Asp	Ile	Gln 95	Pro	Phe		348
tcc cag Ser Gln	100	Pro Thi	c Gln	Leu A 1	la Glu 05	ı Met	Ile	Phe	Asn 110	Leu	Leu	Leu		3,96
gaa gaa Glu Glu 115	Lys A	Arg Ile	e Leu	Ile G 120	ln Ala	Gln	Arg	Ala 125	Gln	Leu	Glu	Gln		444
gga gag Gly Glu 130	Pro V	/ai Leu	135	Thr P	ro Val	Glu	Ser 140	Gln	Gln	His	Glu	Ile 145		492
gaa tcc Glu Ser	Arg 1	le Leu 150	Asp 1	Leu Ai	rg Ala	Met 155	Met	Glu	Lys	Leu	Val 160	Lys		540
tcc atc Ser Ile	Ser G	65 Leu	Lys A	Asp Gl	In Gln 170	Asp	Val	Phe	Cys	Phe 175	Arg	Tyr		588
aag atc Lys Ile	180	la Lys	Gly I	ys Th	ır Pro 35	Ser	Leu i	Asp	Pro 1 190	His :	Gln	Thr		636
aaa gag Lys Glu 195	Gln L	ys Ile	Leu G	ln G1 00	u Thr	Leu	Asn (Glu : 205	Leu i	Asp :	Lys	Arg		684
aga aag Arg Lys 210	gağ g: Glu V	tg ctg al Leu	gat g Asp A 215	cc tc la Se	c aaa r Lys	Ala	ctg d Leu J 220	cta Leu (ggc (Gly)	ega : Arg :	Leu	act Thr 225		732

Figure 2

acc Thr	cta Leu	atc Ile	gag Glu	cta Leu 230	ctg Leu	ctg Leu	cca Pro	aag Lys	ttg Leu 235	gag Glu	gag Glu	tgg Trp	aag Lys	gcc Ala 240	cag Gln	780
cag Gln	caa Gln	aaa Lys	gcc Ala 245	tgc Cys	atc Ile	aga Arg	gct Ala	ccc Pro 250	att Ile	gac Asp	cac His	ggg Gly	ttg Leu 255	gaa Glu	cag Gln	828
ctg Leu	gag Glu	aca Thr 260	tgg Trp	ttc Phe	aca Thr	got Ala	gga Gly 265	gca Ala	aag Lys	ctg Leu	ttg Leu	ttt Phe 270	cac His	ctg Leu	agg Arg	8-6
cag Gln	ctg Leu 275	ctg Leu	aag Lys	gag Glu	ctg Leu	aag Lys 280	gga Gly	ctg Leu	agt Ser	tgc Cys	ctg Leu 285	gtt Val	agc Ser	tat Tyr	cag Gln	924
gat Asp 290	gac Asp	cct Pro	ctg Leu	acc Thr	aaa Lys 295	gly ggg	gtg Val	gac Asp	cta Leu	cgc Arg 300	aac Asn	gcc Ala	cag Gln	gtc Val	aca Thr 305	972
gag Glu	ttg Leu	cta Leu	cag Gln	cgt Arg 310	ctg Leu	ctc Leu	cac His	aga Arg	gcc Ala 315	ttt Phe	gtg Val	gta Val	gaa Glu	acc Thr 320	cag Gln	1020
ccc Pro	tgc Cys	atg Met	ccc Pro 325	caa Gln	act Thr	ccc Pro	cat His	cga Arg 330	ccc Pro	ctc Leu	atc Ile	ctc Leu	aag Lys 335	act Thr	ggc Gly	1068
agc Ser	aag Lys	ttc Phe 340	acc Thr	gtc Val	cga Arg	aca Thr	agg Arg 345	ctg Leu	ctg Leu	gtg Val	aga Arg	ctc Leu 350	câg Gln	gaa Glu	ggc Gly	1116
aat Asn	gag Glu 355	tca Ser	ctg Leu	act Thr	gtg Val	gaa Glu 360	gtc Val	tcc Ser	att Ile	gac Asp	agg Arg 365	aat Asn	cct Pro	cct Pro	caa Gln	1164
tta Leu 370	caa Gln	ggc Gly	ttc Phe	cgg Arg	aag Lys 375	ttc Phe	aac Asn	att Ile	ctg Leu	act Thr 380	tca Ser	aac Asn	cag Gln	aaa Lys	act Thr 385	1212
ttg Leu	acc Thr	ccc Pro	gag Glu	aag Lys 390	Gly ggg	cag Gln	agt Ser	cag Gln	ggt Gly 395	ttg Leu	att Ile	tgg Trp	gac Asp	ttt Phe 400	ggt Gly	1260
tac Tyr	ctg Leu	act Thr	ctg Leu 405	gtg Val	gag Glu	caa Gln	cgt Arg	tca Ser 410	ggt Gly	ggt Gly	tca Ser	gga Gly	aag Lys 415	ggc Gly	agc Ser	1308
aat Asn	aag Lys	ggg Gly 420	cca Pro	cta Leu	ggt Gly	gtg Val	aca Thr 425	gag Glu	gaa Glu	ctg Leu	cac His	atc Ile 430	atc Ile	agc Ser	ttc Phe	1356
acg Thr	gtc Val 435	aaa Lys	tat Tyr	acc Thr	tac Tyr	cag Gln 440	ggt Gly	ctg Leu	aag Lys	cag Gln	gag Glu 445	ctg Leu	aaa Lys	acg Thr	gac Asp	1404

ac Th: 45	r Le	c cc u Pr	t gto o Val	g gto l Val	g att 1 116 455	e Ile	tec Ser	aac Asn	atg Met	aac Asn 460	Gln	ctc Leu	tca Ser	att Ile	gcc Ala 465	1452
tg: Tr	g gct p Ala	t to a Se	a gtt r Val	- ctc Leu 470	ı Trp	ttc Phe	: aat : Asn	ttg Leu	ctc Leu 475	Ser	cca Pro	aac Asn	ctt Leu	Cag Gln 480	Asn	1500
caç Glr	g caq n Glr	g tte n Phe	c tto ≥ Phe 485	e Ser	aac Asn	ccc Pro	Pro	aag Lys 490	gcc Ala	ccc Pro	tgg Trp	agc Ser	ttg Leu 495	ctg Leu	ggc Gly	1548
cct Pro	gct Ala	cto Lev 500	agt Ser	tgg Trp	cag Gln	ttc Phe	tec Ser 505	Ser	tat Tyr	gtt Val	ggc Gly	cga Arg 510	ggc Gly	ctc Leu	aac Asn	1596
tca Ser	gac Asp 515	Glr	ctg Leu	agc Ser	atg Met	ctg Leu 520	aga Arg	aac Asn	aag Lys	ctg Leu	ttc Phe 525	G] A BBB	cag Gln	aac Asn	tgt Cys	1644
agg Arg 530	Thr	gaç Glu	gat Asp	cca Pro	tta Leu 535	ttg Leu	tcc Ser	tgg Trp	gct Ala	gac Asp 540	ttc Phe	act Thr	aag Lys	cga Arg	gag Glu 545	1692
agc Ser	Pro	ect Pro	ggc	aag Lys 550	tta Leu	cca Pro	ttc Phe	tgg Trp	aca Thr 555	tgg Trp	ctg Leu	gac Asp	aaa Lys	att Ile 560	ctg Leu	1740
gag Glu	ttg Leu	gta Val	cat His 565	gac Asp	cac His	ctg Leu	aag Lys	gat Asp 570	ctc Leu	tgg Trp	aat Asn	gat Asp	gga Gly 575	ege Arg	atc Ile	1788
atg Met	ggc Gly	ttt Phe 580	gtg Val	agt Ser	cgg Arg	agc Ser	cag Gln 585	gag Glu	ege Arg	cgg Arg	ctg Leu	ctg Leu 590	aag Lys	aag Lys	acc Thr	1836
atg Met	tct Ser 595	ggc Gly	acc Thr	ttt Phe	cta Leu	ctg Leu 600	cgc Arg	ttc Phe	agt Ser	gaa Glu	tcg Ser 605	tca Ser	gaa Glu	GJ À aaa	Gly ggc	1884
Ile 610	Thr	Cys	tcc Ser	Trp	Val 615	Glu	His	Gln	Asp	Asp 620	Asp	Lys	Val	Leu	Ile 625	1932
tac Tyr	tct Ser	gtg Val	caa Gln	ccg Pro 630	tac Tyr	acg Thr	aag Lys	gag Glu	gtg Val 635	ctg Leu	cag Gln	tca Ser	ctc Leu	ccg Pro 640	ctg Leu	1980
act Thr	gaa Glu	atc Ile	atc Ile 645	egc Arg	cat His	tac Tyr	cag Gln	ttg Leu 650	ctc Leu	act Thr	gag Glu	gag Glu	aat Asn 655	ata Ile	cct Pro	2028
Glu	Asn	Pro 660	ctg Leu	Arg	Phe	Leu	Tyr 665	Pro	Arg	Ile	Pro	Arg 670	Asp	Glu	Ala	2076
Phe	ggg Gly 675	tgc Cys	tac Tyr	tac Tyr	Gln	gag Glu 680	aaa Lys	gtt Val	aat Asn	ctc Leu	cag Gln 685	gaa Glu	cgg Arg	agg Arg	aaa Lys	2124

tac ctg aaa cac agg ctc att gtg gtc tct aat aga cag gtg gat gaa 2172 Tyr Leu Lys His Arg Leu Ile Val Val Ser Asn Arg Gln Val Asp Glu 690 ctg caa caa ccg ctg gag ctt aag cca gag cca gag ctg gag tca tta 2220 Leu Gln Gln Pro Leu Glu Leu Lys Pro Glu Pro Glu Leu Glu Ser Leu 710 715 gag ctg gaa cta ggg ctg gtg cca gag cca gag ctc agc ctg gac tta 2268 Glu Leu Glu Leu Gly Leu Val Pro Glu Pro Glu Leu Ser Leu Asp Leu gag cca ctg ctg aag gca ggg ctg gat ctg ggg cca gag cta gag tct 2316 Glu Pro Leu Leu Lys Ala Gly Leu Asp Leu Gly Pro Glu Leu Glu Ser 745 gtg ctg gag tcc act ctg gag cct gtg ata gag ccc aca cta tgc atg 2364 Val Leu Glu Ser Thr Leu Glu Pro Val Ile Glu Pro Thr Leu Cys Met 755 760 gta toa caa aca gtg coa gag coa gac caa gga cot gta toa cag coa 2412 Val Ser Gln Thr Val Pro Glu Pro Asp Gln Gly Pro Val Ser Gln Pro gtg cca gag cca gat ttg ccc tgt gat ctg aga cat ttg aac act gag 2460 Val Pro Glu Pro Asp Leu Pro Cys Asp Leu Arg His Leu Asn Thr Glu cca atg gaa atc ttc aga aac tgt gta aag att gaa gaa atc atg ecg 2508 Pro Met Glu Ile Phe Arg Asn Cys Val Lys Ile Glu Ile Met Pro 805 810 815 aat ggt gac cca ctg ttg gct ggc cag aac acc gtg gat gag gtt tac 2556 Asn Gly Asp Pro Leu Leu Ala Gly Gln Asn Thr Val Asp Glu Val Tyr 825 830 gto too ego coo ago cao tto tao act gat gga coo ttg atg cot tot 2604 Val Ser Arg Pro Ser His Phe Tyr Thr Asp Gly Pro Leu Met Pro Ser 840 845 gac ttc tag gaaccacatt tcctctgttc ttttcatatc tctttgccct 2653 Asp Phe * 850 tectactect catageatga tattgttete caaggatggg aateaggeat gtgteeette 2713 caagetgtgt taactgttca aactcaggee tgtgtgacte cattggggtg agaggtgaaa 2773 qcataacatg ggtacagagg ggacaacaat gaatcagaac agatgctgag ccataggtct 2833 aaataggatc ctggaggctg cctgctgtgc tgggaggtat aggggtcctg ggggcaggcc 2893 agggcagttg acaggtactt ggagggctca gggcagtggc ttctttccag tatggaagga 2953 tttcaacatt ttaatagttg gttaggctaa actggtgcat actggcattg gccttggtgg 3013 ggagcacaga cacaggatag gactccattt ctttctcca ttccttcatg tctaggataa 3073 cttgctttct tctttccttt actcctggct caagccctga atttcttctt ttcctgcagg 3133 ggttgagagc tttctgcctt agcctaccat gtgaaactct accctgaaga aagggatgga 3193 taggaagtag acetetttt ettaccagte tectecceta etetgeceee taagetgget 3253 gtacetgttc eteccecata aaatgateet gecaatet 3291

Figure 2 (cont'd)

ago	cgaç	gga	acaa	gccc agag	ca a gc c	ccgg cttc	atec ggcc	t gg. t ga	acag ggga	gcac gcct	ccc cgc	gget egee gee	tgg cgt caa	cgct cccc tgg	cgtcgc gtctct ggcaca aat Asn 5	60 120 180 235
		cag Gln			Asp											283
		gac Asp		Phe												331
		agt Ser 40	Gln													379
		gtg Val														427
		ctg Leu														-475
		cag Gln														523
		att Ile														571
		gcc Ala 120														619
gca Ala	gcc Ala 135	gtg Val	gtg Val	acg Thr	gag Glu	aag Lys 140	cag Gln	cag Gln	atg Met	ctg Leu	gag Glu 145	cag Gln	cac His	ctt Leu	cag Gln	667
gat Asp 150	gtc Val	cgg Arg	aag Lys	aga Arg	gtg Val 155	cag Gln	gat Asp	cta Leu	gaa Glu	cag Gln 160	aaa Lys	atg Met	aaa Lys	gtg Val	gta Val 165	715
		ctc Leu														763
caa Gln	gga Gly	gac Asp	atg Met 185	caa Gln	gat Asp	ctg Leu	aat Asn	gga Gly 190	aac Asn	aac Asn	cag Gln	tca Ser	gtg Val 195	acc Thr	agg Arg	811
		atg Met 200														859

Figure 3

cgg Arg	aga Arg 215	agc Ser	atc Ile	gtg Val	agt Ser	gag Glu 220	ctg Leu	gcg Ala	GJ A GGG	ctt Leu	ttg Leu 225	tca Ser	gcg Ala	atg Met	gag Glu	907
tac Tyr 230	gtg Val	cag Gln	aaa Lys	act Thr	ctc Leu 235	acg Thr	gac Asp	gag Glu	gag Glu	ctg Leu 240	gct Ala	gac Asp	tgg Trp	aag Lys	agg Arg 245	955
cgg Arg	caa Gln	cag Gln	att Ile	gcc Ala 250	tgc Cys	att Ile	gga Gly	ggc Gly	ccg Pro 255	ccc Pro	aac neA	atc Ile	tgc Cys	cta Leu 260	gat Asp	1003
cgg Arg	cta Leu	gaa Glu	aac Asn 265	tgg Trp	ata Ile	acg Thr	tca Ser	tta Leu 270	gca Ala	gaa Glu	tct Ser	caa Gln	ctt Leu 275	cag Gln	acc Thr	1051
cgt Arg	caa Gln	caa Gln 280	att Ile	aag Lys	aaa Lys	ctg Leu	gag Glu 285	gag Glu	ttg Leu	cac His	caa Gln	aaa Lys 290	gtt Val	t <i>c</i> c Ser	tac Tyr	1099
aaa Lys	ggg Gly 295	gac Asp	ccc Pro	att Ile	gta Val	cag Gln 300	cac His	cgg Arg	ccg Pro	atg Met	ctg Leu 305	gag Glu	gag Glu	agg Arg	atc Ile	1147
gtg Val 310	gag Glu	ctg Leu	ttc Phe	aga Arg	aac Asn 315	tta Leu	atg Met	aaa Lys	agt Ser	gcc Ala 320	ttt Phe	gtg Val	gtg Val	gag Glu	cgg Arg 325	1195
cag Gln	ccc Pro	tgc Cys	atg Met	ccc Pro 330	atg Met	cat His	cct Pro	gac Asp	cgg Arg 335	ccc Pro	ctc Leu	gtc Val	atc Ile	aag Lys 340	acc Thr	1243
ggc Gly	gtc Val	cag Gln	ttc Phe 345	act Thr	act Thr	aaa Lys	gtc Val	agg Arg 350	ttg Leu	ctg Leu	gtc Val	aag Lys	ttc Phe 355	cct Pro	gag Glu	1291
ttg Leu	aat Asn	tat Tyr 360	cag Gln	ctt Leu	aaa Lys	att Ile	aaa Lys 365	gtg Val	tgc Cys	att Ile	gac Asp	aaa Lys 370	gac Asp	tct Ser	G1 y ggg	1339
gac Asp	gtt Val 375	gca Ala	gct Ala	ctc Leu	aga Arg	gga Gly 380	tcc Ser	cgg Arg	aaa Lys	Phe	aac Asn 385	att Ile	ctg Leu	G] A āāc	aca Thr	1387
aac Asn 390	aca Thr	aaa Lys	gtg Val	atg Met	aac Asn 395	atg Met	gaa Glu	gaa Glu	tcc Ser	aac Asn 400	aac Asn	ggc Gly	agc Ser	ctc Leu	tct Ser 405	1435
gca Ala	gaa Glu	ttc Phe	aaa Lys	cac His 410	ttg Leu	acc Thr	ctg Leu	agg Arg	gag Glu 415	cag Gln	aga Arg	tgt Cys	ggg Gly	aat Asn 420	GŢĀ āāā	1483
ggc Gly	cga Arg	gcc Ala	aat Asn 425	tgt Cys	gat Asp	gct Ala	tcc Ser	ctg Leu 430	att Ile	gtg Val	act Thr	gag Glu	gag Glu 435	ctg Leu	cac His	1531
ctg Leu	atc Ile	acc Thr 440	ttt Phe	gag Glu	acc Thr	gag Glu	gtg Val 445	tat Tyr	cac His	caa Gln	ggt Gly	ctc Leu 450	aag Lys	att Ile	gac Asp	1579

Figure 3 (cont'd)

cta Leu	gag Glu 455	acc Thr	cac His	tcc Ser	ttg Leu	tca Ser 460	gtt Val	gtg Val	gtg Val	atc Ile	tcc Ser 465	aac Asn	atc Ile	tgt Cys	cag Gln	1627
atg Met 470	cca Pro	aat Asn	gcc Ala	tgg Trp	gcg Ala 475	tcc Ser	atc Ile	ctg Leu	tgg Trp	tac Tyr 480	aac Asn	atg Met	ctg Leu	acc Thr	aac Asn 485	1675
aat Asn	ccc Pro	aag Lys	aat Asn	gtg Val 490	aac Asn	ttc Phe	ttc Phe	act Thr	aag Lys 495	ccg Pro	cca Pro	att Ile	gga Gly	acc Thr 500	tgg Trp	1723
gac Asp	caa Gln	gtg Val	gcc Ala 505	gag Glu	gtg Val	ctc Leu	agc Ser	tgg Trp 510	cag Gln	ttc Phe	tcg Ser	tcc Ser	acc Thr 515	acc Thr	aag Lys	1771
cgg Arg	ggg Gly	ctg Leu 520	agc Ser	atc Ile	gag Glu	cag Gln	ctg Leu 525	aca Thr	acg Thr	ctg Leu	gct Ala	gag Glu 530	aag Lys	ctc Leu	cta Leu	1819
G1 y ggg	cct Pro 535	ggt Gly	gtg Val	aac Asn	tac Tyr	tca Ser 540	Gly ggg	tgt Cys	cag Gln	atc Ile	aca Thr 545	tgg Trp	gct Ala	aac Asn	ttc Phe	1867
tgc Cys 550	aaa Lys	gaa Glu	aac Asn	atg Met	gct Ala 555	ggc Gly	aag Lys	ggc Gly	ttc Phe	tcc Ser 560	tac Tyr	tgg Trp	gtc Val	tgg Trp	cta Leu 565	1915
gaç Asp	aat Asn	atc Ile	atc Ile	gac Asp 570	ctt Leu	gtg Val	aaa Lys	aag Lys	tat Tyr 575	atc Ile	ttg Leu	gcc Ala	ctt Leu	tgg Trp 580	aat Asn	1963
gaa Glu	ggg Gly	tac Tyr	atc Ile 585	atg Met	ggt Gly	ttc Phe	atc Ile	agc Ser 590	aag Lys	gag Glu	cgg Arg	gag Glu	cgg Arg 595	gcc Ala	atc Ile	2011
ttg Leu	agc Ser	act Thr 600	aag Lys	ccc Pro	cca Pro	ggc Gly	acc Thr 605	ttc Phe	ctg Leu	ctg Leu	cgc	ttc Phe 610	agt Ser	gaa Glu	agc Ser	2059
agc Ser	aaa Lys 615	gaa Glu	gga Gly	ggc Gly	gtc Val	act Thr 620	ttc Phe	açt Thr	tgg Trp	gtg Val	gag Glu 625	aag Lys	gac Asp	atc Ile	agc Ser	2107
ggt Gly 630	aag Lys	acc Thr	cag Gln	atc Ile	cag Gln 635	tcc Ser	gtg Val	gaa Glu	cca Pro	tac Tyr 640	aca Thr	aag Lys	cag Gln	cag Gln	ctg Leu 645	2155
aac Asn	aac Asn	atg Met	tca Ser	ttt Phe 650	gct Ala	gaa Glu	atc Ile	atc Ile	atg Met 655	Gly ggc	tat Tyr	aag Lys	atc Ile	atg Met 660	gat Asp	2203
gct Ala	acc Thr	aat Asn	atc Ile 665	ctg Leu	ttg Leu	tct Ser	cca Pro	ctt Leu 670	gtc Val	tat Tyr	ctc Leu	tat Tyr	cct Pro 675	gac Asp	att Ile	2251

Figure 3 (cont'd)

ccc Pro	aag Lys	gag Glu 680	gag Glu	gca Ala	ttc Phe	G1A āāā	aag Lys 685	tat Tyr	tgt Cys	cgg Arg	cca Pro	gag Glu 690	agc Ser	cag Gln	gag Glu	2299
cat His	cct Pro 695	gaa Glu	gct Ala	gac Asp	cca Pro	ggt Gly 700	agc Ser	gct Ala	gcc Ala	cca Pro	tac Tyr 705	ctg Leu	aag Lys	acc Thr	aag Lys	2347
ttt Phe 710	atc Ile	tgt Cys	gtg Val	aca Thr	cca Pro 715	acg Thr	acc Thr	tgc Cys	agc Ser	aat Asn 720	acc Thr	att Ile	gac Asp	ctg Leu	cag Pro 725	2395
atg Met	tcc Ser	ccc Pro	cgc Arg	gct Ala 730	tta Leu	gat Asp	tca Ser	ttg Leu	atg Met 735	cag Gln	ttt Phe	gga Gly	aat Asn	aat Asn 740	ggt Gly	2443
gaa Glu	ggt Gly	Ala	gaa Glu 745	ccc Pro	tca Ser	gca Ala	Gly	999 Gly 750	cag Gln	ttt Phe	gag Glu	tcc Ser	ctc Leu 755	acc Thr	ttt Phe	2491
gac Asp	atg Met	gag Glu 760	ttg Leu	acc Thr	tcg Ser	Glu	tgc Cys 765	gct Ala	acc Thr	tcc Ser	ccc Pro	atg Met 770	tga *			2533
ccct ttta	caca atct aatg	ca g cc ta tg g	ccaa: actt gtga:	accc ctgc	c ag t at	atca cttt	tctg gagc	aaa aat	ctac	taa gca	cttt	gtgg taaa	tt c aa t	caga agag	gecac ttttt aaatg gaccc	2593 2653 2713 2773 2787

gct	ttet tget	ect gag	aggg agag	actg cgcta	tg ac	atg	tct	cag	tgg	aat	actt caa Gln	gtc	caa	cag	gggac tta Leu 10	60 111
gaa Glu	atc Ile	aag Lys	ttt Phe	ttg Leu 15	gag Glu	cag Gln	gtg Val	gat Asp	caa Gln 20	ttc Phe	tat Tyr	gat Asp	gac Asp	aac Asn 25	ttt Phe	159
ccc Pro	atg Met	gaa Glu	att Ile 30	cgg Arg	cat His	ctg Leu	ttg Leu	gcc Ala 35	caa Gln	tgg Trp	att Ile	gaa Glu	aat Asn 40	caa Gln	gac Asp	207
tgg Trp	gag Glu	gca Ala 45	gct Ala	tct Ser	aac Asn	aat Asn	gaa Glu 50	acc Thr	atg Met	gca Ala	acg Thr	att Ile 55	ctt Leu	ctt Leu	caa Gln	255
aac Asn	ttg Leu 60	tta Leu	ata Ile	caa Gln	ctg Leu	gat Asp 65	gaa Glu	cag Gln	tta Leu	ggt Gly	cgt Arg 70	gtt Val	tcc Ser	aaa Lys	gag Glu	303
aaa Lys 75	aac Asn	cta Leu	ctc Leu	ttg Leu	ata Ile 80	cac His	aat Asn	cta Leu	aaa Lys	aga Arg 85	att Ile	agg Arg	aag Lys	gtc Val	ctt Leu 90	351
cag Gln	gga Gly	aaa Lys	ttt Phe	cat His 95	gga Gly	aat Asn	cca Pro	atg Met	cat His 100	gta Val	gct Ala	gtg Val	gtt Val	att Ile 105	tca Ser	399
aac Asn	tgt Cys	tta Leu	agg Arg 110	gaa Glu	gag Glu	agg Arg	aga Arg	ata Ile 115	ttg Leu	gct Ala	gca Ala	gcc Ala	aac Asn 120	atg Met	cct Pro	447
gtc Val	cag Gln	ggg Gly 125	cct Pro	cta Leu	gag Glu	aaa Lys	tcc Ser 130	tta Leu	caa Gln	agt Ser	tct Ser	tca Ser 135	gtt Val	tca Ser	gaa Glu	495
aga Arg	cag Gln 140	agg Arg	aat Asn	gtg Val	gag Glu	cac His 145	aaa Lys	gtg Val	gct Ala	gcc Ala	att Ile 150	aaa Lys	aac Asn	agt Ser	gtg Val	543
cag G1n 155	atg Met	aca Thr	gaa Glu	caa Gln	gat Asp 160	acc Thr	aaa Lys	tac Tyr	tta Leu	gaa Glu 165	gat Asp	ctg Leu	caa Gln	gac Asp	gaa Glu 170	591
ttt Phe	gac Asp	tac Tyr	agg Arg	tat Tyr 175	aaa Lys	aca Thr	att Ile	cag Gln	aca Thr 180	atg Met	gat Asp	cag Gln	agt Ser	gac Asp 185	aag Lys	639
aat Asn	agt Ser	gcc Ala	atg Met 190	gtg Val	aat Asn	cag Gln	gaa Glu	gtt Val 195	ttg Leu	aca Thr	ctg Leu	cag Gln	gaa Glu 200	atg Met	ctt Leu	687
aac Asn	agc Ser	ctc Leu 205	gat Asp	ttc Phe	aag Lys	aga Arg	aag Lys 210	gag Glu	gct Ala	ctc Leu	agt Ser	aaa Lys 215	atg Met	acc Thr	caa Gln	735

Figure 4

ato Ile	220	e His	t gag s Glu	aca Thr	gac Asp	ctg Leu 225	Leu	atg Met	aac Asn	acc Thr	atg Met 230	ctc Leu	ata Ile	gaa Glu	gag Glu	783
cto Leu 235	ı Glr	gad Asp	tgg Trp	aag Lys	cgg Arg 240	cgg Arg	cag Gln	caa Gln	atc Ile	gcc Ala 245	tgc Cys	atc Ile	ggg Gly	ggt Gly	cca Pro 250	831
cto Leu	cac His	: aat : Asr	ggg Gly	ctc Leu 255	gac A s p	cag Gln	ctt Leu	cag Gln	aac Asn 260	tgc Cys	ttt Phe	aca Thr	cta Leu	ttg Leu 265	gca Ala	879
gaa Glu	agt Ser	ctt Leu	ttc Phe 270	caa Gln	ctg Leu	aga Arg	agg Arg	caa Gln 275	ttg Leu	gag Glu	aaa Lys	cta Leu	gag Glu 280	gag Glu	caa Gln	927
tct Ser	acc Thr	aaa Lys 285	atg Met	aca Thr	tat Tyr	gaa Glu	ggt Gly 290	gat Asp	ccc	att Ile	cca Pro	atg Met 295	caa Gln	aga Arg	act Thr	975
cac His	atg Met 300	cta Leu	gaa Glu	aga Arg	gtc Val	acc Thr 305	ttc Phe	ttg Leu	atc Ile	tac Tyr	aac Asn 310	ctt Leu	ttc Phe	aag Lys	aac Asn	1023
tca Ser 315	ttt Phe	gtg Val	gtt Val	gag Glu	cga Arg 320	cag Gln	cca Pro	tgt Cys	atg Met	cca Pro 325	acc Thr	cac His	cct Pro	cag Gln	agg Arg 330	1071
ccg Pro	ttg Leu	gta Val	ctt Leu	aaa Lys 335	acc Thr	cta Leu	att Ile	cag Gln	ttc Phe 340	act Thr	gta Val	aaa Lys	cta Leu	agg Arg 345	cta Leu	1119
cta Leu	ata Ile	aaa Lys	ttg Leu 350	cca Pro	gaa Glu	cta Leu	aac Asn	tat Tyr 355	cag Gln	gta Val	aag Lys	gtt Val	aag Lys 360	gca Ala	tca Ser	1167
att Ile	gac Asp	aag Lys 365	aat Asn	gtt Val	tca Ser	act Thr	cta Leu 370	agc Ser	aac Asn	cga Arg	aga Arg	ttt Phe 375	gta Val	ctt Leu	tgt Cys	1215
Gly	Thr 380	Asn	gtc Val	Lys	Ala	Met 385	Ser	Ile	Glu	Glu	Ser 390	Ser	Asn	Gly	Ser	1263
Leu 395	Ser	Val	gaa Glu	Phe	Arg 400	His	Leu	Gln	Pro	Lys 405	Glu	Met	Lys	Ser	Ser 410	1311
Ala	Gly	Gly	aaa Lys	Gly 415	Asn	Glu	Gly	Cys	His 420	Met	Val	Thr	Glu	Glu 425	Leu	1359
His	Ser	Ile	acg Thr 430	Phe	Glu '	Thr	Gln	Ile 435	Cys	Leu	Tyr	Gly	Leu 440	Thr	Ile	1407
gat Asp	Leu	gag Glu 445	acc Thr	agc Ser	tca : Ser :	Leu	cct Pro 450	gtg Val	gtg Val	atg Met	att Ile	tcc Ser 455	aat Asn	gtc Val	agt Ser	1455

		Pro	aat Asn									1503
			cag Gln									1551
			cta Leu									1599
			ctt Leu 510									1647
			tct Ser									1695
_	_	_	cat His									1743
_			ttg Leu	_		_						1791
_			gt <i>c</i> Val	_		_		 	4.00	 _	400	1839
			aaa Lys 590									1887
			gga Gly									1935
			ttc Phe									1983
			ttc Phe									2031
			cct Pro									2079
			gcc Ala 670									2127

	aga Arg															2175
	ccc Pro 700															2223
	gac Asp															2271
	ctg Leu	-					-			_	_					2319
gct Ala	gaa Glu	tga *	cagç	ataa	ac t	ctga	icgca	ic ca	agaa	agga	a ago	aaat	gaa			2368
ccag attg	gttc	ta g tg t	gaaa tgtg	tgtt actg	t ga a aa	cato tgct	tgaa tgaa	gct	ctet	tca	cact	cccc	jtg q	gcact	aaata cotoa igataa	2428 2488 2548 2588

Figure 4 (cont'd)

tg	aact	eget	ggad	cagaç	get a	acaga	iccta	it go	ggcc	tgga	agt	gccc tc a M	gct tg t	gaga ct c	cagcct aaggga tg tgg eu Trp	60 120 177
ggt Gly 5	cto Lei	g gto i Val	c tco L Ser	aaç Lys	g ato Met	p ccc	cca Pro	gaa Glu	aaa Lys	gtg Val 15	Gln	cgg Arg	ctc Leu	tat Tyr	gtc Val 20	225
gac Asp	ttt Phe	ccc Pro	caa Glr	cac His 25	Let	g egg Arg	cat His	ctt Leu	ctg Leu 30	ggt Gly	gac Asp	tgg Trp	ctg Leu	gag Glu 35	ago Seb	273
cag Gln	Pro	tgg Trp	gag Glu 40	Phe	ctg Leu	gtc Val	ggc Gly	tcc Ser 45	gac Asp	gcc Ala	ttc Phe	tgc Cys	tgc Cys 50	aac Asn	ttg Leu	321
gct Ala	agt Ser	gcc Ala 55	Leu	ctt Leu	tca Ser	gac Asp	act Thr 60	gtc Val	cag Gln	cac His	ctt Leu	cag Gln 65	gcc Ala	tcg Ser	gtg Val	369
gga Gly	gag Glu 70	cag Gln	ggg Gly	gag Glu	ggg Gly	agc Ser 75	acc Thr	atc Ile	ttg Leu	caa Gln	cac His 80	atc Ile	agc Ser	acc Thr	ctt Leu	417
gag Glu 85	Ser	ata Ile	tat Tyr	cag Gln	agg Arg 90	gac A sp	ccc	ctg Leu	aag Lys	ctg Leu 95	gtg Val	gcc Ala	act Thr	ttc Phe	aga Arg 100	465
caa Gln	ata Ile	ctt Leu	caa Gln	gga Gly 105	gag Glu	aaa Lys	aaa Lys	gct. Ala	gtt Val 110	atg Met	gaa Glu	cag Gln	ttc Phe	cgc Arg 115	cac His	513
ttg Leu	cca Pro	atg Met	cct Pro 120	ttc Phe	cac His	tgg Trp	aag Lys	cag Gln 125	gaa Glu	gaa Glu	ctc Leu	aag Lys	ttt Phe 130	aag Lys	aca Thr	561
ggc Gly	ttg Leu	cgg Arg 135	agg Arg	ctg Leu	cag Gln	cac His	cga Arg 140	gta Val	G] À aàa	gag Glu	atc Ile	cac His 145	ctt Leu	ctc Leu	cga Arg	609
gaa Glu	gcc Ala 150	ctg Leu	cag Gln	aag Lys	ggg Gly	gct Ala 155	gag Glu	gct Ala	ggc Gly	caa Gln	gtg Val 160	tct Ser	ctg Leu	cac His	agc Ser	657
ttg Leu 165	ata Ile	gaa Glu	act Thr	cct Pro	gct Ala 170	aat Asn	G 1 У	act Thr	GJA aaa	cca Pro 175	agt Ser	gag Glu	gec Ala	ctg Leu	gcc Ala 180	705
atg Met	cta Leu	ctg Leu	cag Gln	gag Glu 185	acc Thr	act Thr	gga Gly	gag Glu	cta Leu 190	gag Glu	gca Ala	gcc Ala	aaa Lys	gcc Ala 195	cta Leu	753
gtg Val	ctg Leu	aag Lys	agg Arg 200	atc Ile	cag Gln	att Ile	tgg Trp	aaa Lys 205	cgg Arg	cag Gln	cag Gln	cag Gln	ctg Leu 210	gca Ala	GJ A ādā	801

Figure 5

aa As	t gg n Gl	уΑ.	ca c la P l5	cg ro	ttt Phe	ga Gl	g ga u Gl	g ag i Se 22	r Lei	g gco 1 Ala	c cca a Pro	cto Leu	cag Glr 225	Glu	g ago	g tgt g Cys	849
ga Gl	a ag u Se 23	r Le	g g eu V	tg al	gac Asp	att Ile	tai Ty: 235	: Se	c caq r Glr	g cta n Leu	a caç ı Glr	caç Glr 240	Glu	gta Val	ggç Gly	g gcg / Ala	897
gc Al 24	a G1:	t gç y Gl	y G	ag lu	ctt Leu	gaç Glu 250	Pro	c aad b Lys	g acc	cgg Arg	g gca g Ala 255	Ser	ctg Leu	act Thr	ggc Gly	cgg Arg 260	945
ct: Le:	g gat ı Asp	t ga o Gl	agi uVa	all	ctg Leu 265	aga Arg	acc Thr	cto Lev	gto ıVal	: acc Thr 270	Ser	tgc Cys	ttc Phe	ctg Leu	gtg Val 275	gag Glu	993
aaq Lys	g cag s Glr	g cc n Pr	C CC O P1 28	0.0	cag Gln	gta Val	ctg Lev	aaç Lys	act Thr 285	Gln	acc Thr	aag L y s	ttc Phe	cag Gln 290	Ala	gga Gly	1041
gtt Val	cga Arg	tt Ph 29	е ње	g t u I	tg Leu	ggc	ttg Leu	agg Arg 300		ctg Leu	ggg Gly	gcc Ala	cca Pro 305	gcc Ala	aag Lys	ect Pro	1089
c c g Pro	ctg Leu 310	va.	c ag l Ar	g g	nja Icc	gac Asp	atg Met 315	gtg Val	aca Thr	gag Glu	aag Lys	cag Gln 320	gcg Ala	cgg Arg	gag Glu	ctg Leu	1137
agt Ser 325	val	Pro	ca Gl	g g n G	gt ly	cct Pro 330	Gly ggg	gct Ala	gga Gly	gca Ala	gaa Glu 335	agc Ser	act Thr	gga Gly	gaa Glu	atc Ile 340	1185
He	Asn	Asr	ı Th	r V 3	al 45	Pro	Leu	Glu	aac Asn	Ser 350	Ile	Pro	Gly	Asn	Cys 355	Cys	1233
Ser	ALa	Leu	36	9 L;	ys .	Asn	Leu	Leu	ctc Leu 365	Lys	Lys	Ile	Lys	Arg 370	Cys	Glu	1281
Arg	Lys	G1 y 375	Th	r G	lu a	Ser	Val	Thr 380	gag Glu	Glu	Lys	Cys	Ala 385	Val	Leu	Phe	1329
Ser	Ala 390	Ser	Ph∈	? Tł	nr 1	Leu	Gly 395	Pro	ggc Gly	Lys	Leu	Pro 400	Ile	Gln	Leu	Gln	1377
405	Leu	Ser	ьег	ı Pı	0 l	110	Val	Val	atc Ile	Val	His 415	Gly	Asn	Gln	Asp	Asn 420	1425
Asn	Ala	Lys	Ala	42	r 1 !5	[le	Leu	Trp	gac Asp	Asn 430	Ala	Phe	Ser	Glu	Met 435	Asp	1473
cgc Arg	gtg Val	ccc Pro	ttt Phe 440	Va	g g	gtg /al .	gct Ala	gag Glu	cgg Arg 445	gtg Val	ccc Pro	tgg Trp	Glu	aag Lys 450	atg Met	tgt Cy s	1521

ga Gl	a ac u Th	t ct r Le 45	u As	c ct n Le	g aaq u Ly:	g tto s Phe	e ato Met 460	: Ala	gaç a Glı	g gto ı Val	g ggg	g acc 7 Thr 465	Asr	egç Arç	l GJA l Gdd	1569
ct: Le:	g ct u Le 47	u Pr	a ga o Gl	g ca u Hi	c tto s Phe	cto Lev 475	ı Ph∈	ctç Lev	geo Ala	caç a Glr	g aaq n Lys 480	: Ile	tto Phe	aat Asn	gac Asp	1617
aa: As: 485	n Sei	c ct r Le	c ag u Se:	t ato r Mei	g gaq E Glu 490	ı Ala	ttc Phe	cag Gln	cac His	cgt Arc 495	Ser	gtg Val	tcc Ser	tgg Trp	tcg Ser 500	1665
caç Glr	g tto n Phe	aa a As:	c aaq n Lys	g gaq s Glu 505	ı Ile	ctg Leu	ctg Leu	ggc Gly	cgt Arg 510	Gly	tto Phe	acc Thr	ttt Phe	tgg Trp 515	Gln	1713
tgg Trp	ttt Phe	ga: Asj	t ggt 520	, Val	ctg Leu	gac Asp	ctc Leu	acc Thr 525	aaa Lys	egc Arg	tgt Cys	ctc Leu	cgg Arg 530	agc Ser	tac Tyr	1761
tgg Trp	tct Ser	gad Asp 539	c egg Arg	ctg Leu	atc Ile	att Ile	ggc Gly 540	ttc Phe	atc Ile	agc Ser	aaa Lys	cag Gln 545	tac Tyr	gtt Val	act Thr	1809
agc Ser	ctt Leu 550	Leu	cto Leu	aat Asn	gag Glu	ccc Pro 555	gac Asp	gga Gly	acc Thr	ttt Phe	ctc Leu 560	ctc Leu	cgc Arg	ttc Phe	agc Ser	1857
gac Asp 565	Ser	gag Glu	att Ile	Gly aaa	ggc Gly 570	atc Ile	acc Thr	att Ile	gcc Ala	cat His 575	gtc Val	atc Ile	cgg Arg	Gly ggc	cag Gln 580	1905
gat Asp	ggc Gly	tct Ser	cca Pro	cag Gln 585	ata Ile	gag Glu	aac Asn	atc Ile	cag Gln 590	cca Pro	ttc Phe	tct Ser	gcc Ala	aaa Lys 595	gac Asp	1953
Leu	Ser	Ile	ege Arg 600	Ser	Leu	Gly	Asp	Arg 605	Ile	Arg	Asp	Leu	Ala 610	Gln	Leu	2001
aaa Lys	aat Asn	ctc Leu 615	tat Tyr	ccc Pro	aag Lys	aag Lys	ccc Pro 620	aag Lys	gat Asp	gag Glu	gct Ala	ttc Phe 625	cgg Arg	agc Ser	cac His	2049
tac Tyr	aag Lys 630	ect Pro	gaa Glu	cag Gln	atg Met	ggt Gly 635	aag Lys	gat Asp	ggc Gly	agg Arg	ggt Gly 640	tat Tyr	gtc Val	cca Pro	gct Ala	2097
645	Tie	rys	atg Met	Thr	Val 650	Glu	Arg	Asp	Gln	Pro 655	Leu	Pro	Thr	Pro	Glu 660	2145
Leu	Gln	Met	cct Pro	Thr 665	Met	Val	Pro	Ser	Tyr 670	Asp	Leu	Gly	Met	Ala 675	Pro	2193
gat Asp	tcc Ser	tcc Ser	atg Met 680	agc Ser	atg Met	cag Gln :	Leu	ggc Gly 685	cca Pro	gat Asp	atg Met	Val	ccc Pro 690	cag Gln	gtg Val	2241

			cac His													2289
gaa Glu	gaa Glu 710	tca Ser	gtc Val	aac Asn	gtg Val	ttg Leu 715	tca Ser	gcc Ala	ttc Phe	cag Gln	gag Glu 720	cct Pro	cac His	ctg Leu	cag Gln	2337
atg Met 725	ccc Pro	ccc Pro	agc Ser	ctg Leu	ggc Gly 730	cag Gln	atg Met	agc Ser	ctg Leu	ccc Pro 735	ttt Phe	gac Asp	cag Gln	cct Pro	cac His 740	2385
			ctg Leu													2433
cct Pro	gac Asp	ccc Pro	ctg Leu 760	ctc Leu	tgc Cys	tca Ser	gat Asp	gtg Val 765	acc Thr	atg Met	gtg Val	gaa Glu	gac Asp 770	agc Ser	tgc Cys	2481
ctg Leu	agc Ser	cag Gln 775	cca Pro	gtg Val	aca Thr	gcg Ala	ttt Phe 780	ect Pro	cag Gln	ggc Gly	act Thr	tgg Trp 785	att Ile	ggt Gly	gaa Glu	2529
			cct Pro													2577
ctt Leu 805	ctc Leu	ctg Leu	gag Glu	GJ A aaa	caa Gln 810	G] A aaa	gag Glu	tcg Ser	ggg Gly	gga Gly 815	ggg Gly	tcc Ser	ttg Leu	ggg Gly	gca Ala 820	2625
cag Gln	ccc Pro	ctc Leu	ctg Leu	cag Gln 825	ccc Pro	tcc Ser	cac His	tat Tyr	999 61y 830	caa Gln	tct Ser	GJ À GG À	atc Ile	tca Ser 835	atg Met	2673
		Met .	gac Asp 840								tga *	tecc	agct	.gg		2719
tcat cagg ctat	gece agga catt gece	tg c aa a cc c ca a	caag gact ctgc catg	cago aaca ccac cctg	a ga g ga c tc c ac	tggg gaat cttc ctgc	gagg gcac cagc agcg	gtg agt act	ccct gggt gact	cct gga gga	atcc gcca aggg	ccac atcc aagt	ct a ac t tc a	ctcc cctt ggct	ettgc tgggt cettt etgag agage	2779 2839 2899 2959 3019 3046

ccc	ett	ctgt	agg	atg Met 1	gta Val	gca Ala	cac His	aac Asn 5	cag Gln	gtg Val	gca Ala	gcc Ala	gac Asp 10	aat Asn	gca Ala	49
gto Val	: tcc Sei	c aca Thi	c Ala	gca Ala	gag Glu	Pro Coc	cga Arg 20	, Arg	cgg Arg	cca Pro	gaa Glu	cct Pro 25	Ser	tco Ser	tct Ser	97
tcc Ser	Ser 30	Ser	tcg Ser	ccc Pro	gcg Ala	gcc Ala 35	Pro	gcg Ala	cgc Arg	ccg Pro	cgg Arg 40	Pro	tgc Cys	c ccc Pro	gcg Ala	145
gtc Val 45	ccg Pro	gco Ala	ccg Pro	gcc Ala	ccc Pro 50	ggc	gac Asp	acg Thr	cac His	ttc Phe 55	Arg	aca Thr	ttc Phe	cgt Arg	tcg Ser 60	193
cac His	gcc Ala	gat Asp	tac Tyr	cgg Arg 65	cgc Arg	atc Ile	acg Thr	cgc Arg	gcc Ala 70	agc Ser	gcg Ala	ctc Leu	ctg Leu	gac Asp 75	Ala	241
tgc Cys	gga Gly	ttc Phe	tac Tyr 80	tgg Trp	GJ y Ggg	ccc Pro	ctg Leu	agc Ser 85	gtg Val	cac His	G] A GGG	gcg Ala	cac His 90	Glu	cgg Arg	289
ctg Leu	cgc Arg	gcc Ala 95	gag Glu	ccc Pro	gtg Val	ggc Gly	acc Thr 100	ttc Phe	ctg Leu	gtg Val	cgc Arg	gac Asp 105	agc Ser	cgc Arg	cag Gln	337
Arg Arg	aac Asn 110	tgc Cys	ttt Phe	ttc Phe	gcc Ala	ctt Leu 115	agc Ser	gtg Val	aag Lys	atg Met	gcc Ala 120	tcg Ser	gga Gly	ccc Pro	acg Thr	385
agc Ser 125	atc Ile	cgc Arg	gtg Val	cac His	ttt Phe 130	cag Gln	gcc Ala	ggc Gly	cgc Arg	ttt Phe 135	cac His	ctg Leu	gat Asp	Gly	agc Ser 140	433
cgc Arg	gag Glu	agc Ser	ttc Phe	gac Asp 145	tgc Cys	ctc Leu	ttc Phe	gag Glu	ctg Leu 150	ctg Leu	gag Glu	cac His	tac Tyr	gtg Val 155	gcg Ala	481
gcg Ala	ccg Pro	cgc Arg	cgc Arg 160	atg Met	ctg Leu	Gl y ggg	gcc Ala	ccg Pro 165	ctg Leu	cgc Arg	cag Gln	cgc Arg	cgc Arg 170	gtg Val	cgg Arg	529
ccg (Pro	ctg Leu	cag Gln 175	gag Glu	ctg Leu	tgc Cys	ege Arg	cag Gln 180	cgc Arg	atc Ile	gtg Val	gcc Ala	acc Thr 185	gtg Val	ggc Gly	cgc Arg	577
gag a Glu <i>l</i>	aac Asn 190	ctg Leu	gct Ala	ege Arg	Ile	ccc Pro 195	ctc Leu	aac Asn	ccc Pro	gtc Val	ctc Leu 200	cgc Arg	gac Asp	tac Tyr	ctg Leu	625
agc t Ser s 205	cc Ser	ttc Phe	ccc Pro	Phe	cag Gln 210	att Ile	tga *	ccgg	cagc	gc c	egec	gtgc	a cg	jcago	atta	679
cctgg	gtt	gg a	ggga accc	gegg	a tg: t ca:	ggtg	tagg ttga	gge	gagg gatc	cgc ctc	etec	cgcc	ct c	aact	cccgg ggaga cctct	700

Figure 6

			cac His							48
			ctg Leu 20							96
			aac Asn							144
			acc Thr							192
			ttt Phe							240
			gtc Val							288
			ggc Gly 100							336
			cgc Arg							384
			gga Gly							432
			ccc Pro							480
			gcc Ala							528
			cgg Arg 180							576
	Arg		acc Thr							624
Gln			G1A ddd	Ile .						672
ctt Leu 225	taa *	gggg								682

Figure 7

get egg aga eec tga gtg	tggge ggcce ggage egage gtage	eegg eete etgg ggee geeg	ggcd tcct cago ggta tttc	egge egge egge ectge eggae agaa	etc great transport to great transport transpo	gettg gegg gagg gtegt geece	ggtg gggtg ggatg tcag tgcg	pt to pe gt pt ga pa go pg go pg aa	egegt eggg gtgg egtg tega caag	cgcc gctg gagc cccg gggc atgt	act agg tga ccc gcg gaa	tegg getg geec eggg etet etgt	ctt ctg aca gct ggt ttc	ctcg cggc ctgg tcgc cgcc	tgacag gccggt gcaggg agggcc agacct cgatct ctgcag tgc atg	
gga Gly	atç Met	gcc Ala	tgc Cys 5	ctt Leu	acg Thr	atg Met	aca Thr	gaa Glu 10	atg Met	gag Glu	gga Gly	aca Thr	tcc Ser 15	acc Thr	tct Ser	528
tct Ser	ata Ile	tat Tyr 20	Gln	aat Asn	ggt Gly	gat Asp	att Ile 25	tct Ser	gga Gly	aat Asn	gcc Ala	aat Asn 30	tct Ser	atg Met	aag Lys	576
caa Gln	ata Ile 35	gat Asp	cca Pro	gtt Val	ctt Leu	cag Gln 40	gtg Val	tat Tyr	ctt Leu	tac Tyr	cat His 45	tcc Ser	ctt Leu	ejà aaa	aaa Lys	624
tct Ser 50	Glu	gca Ala	gat Asp	tat Tyr	ctg Leu 55	acc Thr	ttt Phe	cca Pro	tct Ser	60 Gly ggg	gag Glu	tat Tyr	gtt Val	gca Ala	gaa Glu 65	672
gaa Glu	atc Ile	tgt Cys	att Ile	gct Ala 70	gct Ala	tct Ser	aaa Lys	gct Ala	tgt Cys 75	ggt Gly	atc Ile	aca Thr	cct Pro	gtg Val 80	tat Tyr	720
cat His	aat Asn	atg Met	ttt Phe 85	gct Ala	tta Leu	atg Met	agt Ser	gaa Glu 90	aca Thr	gaa Glu	agg Arg	atc Ile	tgg Trp 95	tat Tyr	cca Pro	768
ccc Pro	aac Asn	cat His 100	gtc Val	ttc Phe	cat His	ata Ile	gat Asp 105	gag Glu	tca Ser	acc Thr	agg Arg	cat His 110	aat Asn	gta Val	ctc Leu	816
tac Tyr	aga Arg 115	ata Ile	aga Arg	ttt Phe	tac Tyr	ttt Phe 120	cct	cgt Arg	tgg Trp	tat Tyr	tgc Cys 125	agt Ser	ggc Gly	agc Ser	aac Asn	864
aga Arg 130	gcc Ala	tat Tyr	cgg Arg	cat His	gga Gly 135	ata Ile	tct Ser	cga Arg	ggt Gly	gct Ala 140	gaa Glu	gct Ala	cct Pro	ctt Leu	ctt Leu 145	912
		ttt Phe														960
gtg V al	cac His	gga Gly	tgg Trp 165	ata Ile	aaa Lys	gta Val	cct Pro	gtg Val 170	act Thr	cat His	gaa Glu	aca Thr	cag Gln 175	gaa Glu	gaa Glu	1008
		999 Gly 180														1056

caa Gln 195	Thr							1104
cca Pro								1152
aag Lys								1200
tgc Cys								1248
gaa Glu								1296
cct Pro 275								1344
act Thr								1392
gag Glu								1440
att Ile								1488
agc Ser								1536
gaa Glu 355								1584
gga Gly								1632
gta V al								1680
cca Pro								1728
cag Gln								1776

Figure 8 (cont'd)

	ttt Phe				Val								1824
His	tgt Cys			Lys									1872
	aag Lys	_		-	_		-	_		-			1920
 -	gaa Glu												1968
	ccc Pro 500												2016
	ggt Gly												2064
	atg Met												2112
	aat Asn											-	2160
 _	ċga Arg	_	-	 	-				_		_		2208
	tta Leu 580												2256
	gaa Glu											•	2304
	aat Asn												2352
	ttt Phe												2400
	tgt Cys												2448

gc Al	a tg a Tr	g gc p Al 66	a Me	g ca t Hi	t tt: s Ph	t cta e Lei	a gaa 1 Gl: 665	ı Glı	a aac 1 Asr	aco Thr	ctt Lei	att 1 Ile 670	His	e Gl7	, aat , Asn	2496
gt. Va.	a tg 1 Cy 67	s Al	c aa a Ly:	a aa s Ası	t att	t ctq E Leu 680	ı Leı	ato Ille	aga Arg	gaa Glu	gaa Glu 685	ı Asp	ago Aro	g aag g Lys	aca Thr	2544
gg: Gl: 690	Y As	t cc n Pr	t cc o Pro	t tto D Phe	c ato ∈ Il∈ 695	: Lys	t ctt	agt Ser	gat Asp	cct Pro 700	Gly	att Ile	agt Ser	att Ile	aca Thr 705	2592
gti Vai	tte	g cca u Pro	a aaq o Lys	g gad s Asp 710) Ile	ctt Leu	cag Gln	gag Glu	aga Arg 715	Ile	cca Pro	tgg Trp	gta Val	Pro 720	cct Pro	2640
gaa Glu	tgo Cys	c att	gaa e Glu 725	Asr.	cct Pro	aaa Lys	aat Asn	tta Leu 730	Asn	ttg Leu	gca Ala	aca Thr	gac Asp 735	Lys	tgg Trp	2688
agt Ser	ttt Phe	ggt Gly 740	/ Thr	act Thr	ttg Leu	tgg Trp	gaa Glu 745		tgc Cys	agt Ser	gga Gly	gga Gly 750	gat Asp	aaa Lys	cct Pro	2736
cta Leu	agt Ser 755	Ala	ctg Leu	gat Asp	tct Ser	caa G1n 760	aga Arg	aag Lys	cta Leu	caa Gln	ttt Phe 765	tat Tyr	gaa Glu	gat Asp	agg Arg	2784
cat His 770	GIn	ctt Leu	cct Pro	gca Ala	cca Pro 775	aag Lys	tgg Trp	gca Ala	gaa Glu	tta Leu 780	gca Ala	aac Asn	ctt Leu	ata Ile	aat Asn 785	2832
Asn	Cys	Met	Asp	Tyr 790	Glu	Pro	Asp	ttc Phe	Arg 795	Pro	Ser	Phe	Arg	Ala 800	Ile	2880
Ile	Arg	Asp	Leu 805	Asn	Ser	Leu	Phe	act Thr 810	Pro	Asp	Tyr	Glu	Leu 815	Leu	Thr	2928
Glu	Asn	820	Met	Leu	Pro	Asn	Met 825	agg Arg	Ile	Gly	Ala	Leu 830	Gly	Phe	Ser	2976
Gly	Ala 835	Phe	Glu	Asp	Arg	Asp 840	Pro	aca Thr	Gln	Phe	Glu 845	Glu	Arg	His	Leu	3024
Lys 850	Phe	Leu	GIn	Gln	Leu 855	Gly	Lys	ggt Gly	Asn	Phe 860	Gly	Ser	Val	Glu	Met 865	3072
Cys	Arg	Tyr	Asp	Pro 870	Leu	Gln	Asp	aac Asn	Thr 875	Gly	Glu	Val	Val	Ala 880	Val	3120
aaa Lys	aag Lys	Leu	cag Gln 885	cat His	agt Ser	act Thr	Glu	gag Glu 890	cac His	cta Leu	aga Arg	Asp	ttt Phe 895	gaa Glu	agg Arg	3168

	att lle		Ile													3216
	gga Gly 915	Val														3264
gaa Glu 930	tat Tyr	tta Leu	cca Pro	tat Tyr	gga Gly 935	agt Ser	tta Leu	cga Arg	gac Asp	tat Tyr 940	ctt Leu	caa Gln	aaa Lys	cat His	aaa Lys 945	3312
	cgg Arg															3360
aag Lys	ggt Gly	atg Met	gag Glu 965	tat Tyr	ctt Leu	ggt Gly	aca Thr	aaa Lys 970	agg Arg	tat Tyr	atc Ile	cac His	agg Arg 975	gat Asp	ctg Leu	3408
	acg Thr															3456
gat Asp	ttt Phe 99	Gly	tta Leu	acc Thr	aaa Lys	gtc Val 1000	Leu	cca Pro	caa Gln	gac Asp	aaa Lys 1005	Glu	tac Tyr	tat Tyr	aaa Lys	3504
gta Val 101	aaa Lys O	gaa Glu	cct Pro	ggt Gly	gaa Glu 1015	Ser	ccc Pro	ata Ile	ttc Phe	tgg Trp 1020	Tyr	gct Ala	cca Pro	gaa Glu	tca Ser 1025	3552
	aca Thr				Phe					Asp					Gly	3600
	gtt Val			Glu					Ile					Ser		3648
	gcg Ala		Phe					Gly					Gly			3696
	gtg Val 1075	Phe					Leu					Gly				3744
	cca Pro)					Asp					Ile					3792
	aac Asn				Asn					Phe					Leu	3840
	gtg V al	Asp		Ile					Ala			aaga	aatg	ac		3886

gc: gg: tg: gt:	gctt: gtct: gcaa: ggct!	gaca gtgc cacc tgct atg	ggct tgaa ctgaa tgaa	tgcae atgte aggae gttga ctg	ott q gta a cct q aca q cgc	ggato etato coctt	ggag gegga ttec gaget tgg	ge ad la et la ga le ea ggg	ctgo atat atggo aggao atg	tgcc tgaa gaaa tcac gcc	teg aca ctg cct agg	ggac ittac gaggo cago ggc	tgc aac cca tga agt	tecg cate gaat geca aag	etgget atgcce ettttga etgetaa ectgee ecc Pro	60 120 180 240 300 348
gtt Val	L Gly	g gat / Asp	t gga o Gly	a gco / Ala	c caç i Gln 20	g ccc Pro	atg Met	gct Ala	gcc Ala	atg Met 25	Gly	ggc Gly	ctg Leu	aag Lys	gtg Val 30	396
ctt Let	t etg Leu	g cac His	c tgg s Trp	get Ala 35	Gly	cca Pro	Gly ggc	Gly ggc	ggg Gly 40	gag Glu	ccc Pro	tgg Trp	gtc Val	act Thr 45	ttc Phe	444
agt Ser	gag Glu	tca Ser	tcg Ser 50	Leu	aca Thr	gct Ala	gag Glu	gaa Glu 55	Val	tgc Cys	atc	cac His	att Ile 60	gca Ala	cat His	492
aaa Lys	gtt Val	ggt Gly 65	Ile	act Thr	cct Pro	cct	tgc Cys 70	ttc Phe	aat Asn	ctc Leu	ttt Phe	gcc Ala 75	ctc Leu	ttc Phe	gat Asp	540
gct Ala	cag Gln 80	Ala	caa Gln	gtc Val	tgg Trp	ttg Leu 85	ccc Pro	cca Pro	aac Asn	cac His	atc Ile 90	cta Leu	gag Glu	atc Ile	ccc Pro	588
aga Arg 95	gat Asp	gca Ala	agc Ser	ctg Leu	atg Met 100	cta Leu	tat Tyr	ttc Phe	cgc Arg	ata Ile 105	agg Arg	ttt Phe	tat Tyr	ttc Phe	cgg Arg 110	636
aac Asņ	tgg Trp	cat His	ggc Gly	atg Met 115	aat Asn	cct Pro	cgg Arg	gaa Glu	ccg Pro 120	gct Ala	gtg Val	tac Tyr	cgt Arg	tgt Cys 125	ej aaa	684
Pro	Pro	Gly	Thr 130	Glu	Ala	tcc Ser	Ser	Asp 135	Gln	Thr	Ala	Gln	Gly 140	Met	Gln	732
ctc Leu	ctg Leu	gac Asp 145	cca Pro	gcc Ala	tca Ser	ttt Phe	gag Glu 150	tac Tyr	ctc Leu	ttt Phe	gag Glu	cag Gln 155	ggc Gly	aag Lys	cat His	780
gag Glu	ttt Phe 160	gtg Val	aat Asn	gac Asp	gtg Val	gca Ala 165	tca Ser	ctg Leu	tgg Trp	gag Glu	ctg Leu 170	tcg Ser	acc Thr	gag Glu	gag Glu	828
Glu 175	Ile	His	His	Phe	Lys 180	aat Asn	Glu	Ser	Leu	Gly 185	Met	Ala	Phe	Leu	His 190	876
Leu	Cys	His	Leu	Ala 195	Leu	cgc Arg	His	Gly	1le 200	Pro	Leu	Glu	Glu	Val 205	Ala	924
aag Lys	aag Lys	acc. Thr	agc Ser 210	ttc Phe	aag Ly s	gac Asp	tgc Cys	atc Ile 215	eeg Pro	cgc Arg	tcc Ser	ttc Phe	cgc Arg 220	Arg	cat His	972

Figure 9

ate Ile	e Arg	g ca g G1: 22:	n His	c ago s Sei	e geo Ala	ctg Leu	acc Thr 230	Arg	ctg Leu	cgc	ctt Leu	egg Arg 235	Asn	gtc Val	ttc Phe	1020
cgo Arg	agg Arg 240	y Phe	c cto	g egg i Arg	g gad g Asp	ttc Phe 245	Gln	ccg Pro	ggc Gly	.cga Arg	ctc Leu 250	Ser	cag Gln	cag Gln	atg Met	1068
gto Val 255	. Met	g gto : Val	c aaa Lys	tac Tyr	cta Leu 260	Ala	aca Thr	ctc Leu	gag Glu	cgg Arg 265	ctg Leu	gca Ala	ccc Pro	ege Arg	ttc Phe 270	1116
ggc Gly	aca Thr	gaç Glu	g cgt Arg	gtg Val 275	Pro	gtg Val	tgc Cys	cac His	ctg Leu 280	agg Arg	ctg Leu	ctg Leu	gcc Ala	cag Gln 285	gcc Ala	1164
gag Glu	ggg Gly	gaç Glu	Pro 290	Cys	tac Tyr	atc Ile	cgg Arg	gac Asp 295	agt Ser	GJ y ggg	gtg V al	gcc Ala	cct Pro 300	aca Thr	gac Asp	1212
cct Pro	ggc Gly	cct Pro 305	gag Glu	tct Ser	gct Ala	gct Ala	ggg Gly 310	ccc Pro	cca Pro	acc Thr	cac His	gag Glu 315	gtg Val	ctg Leu	gtg Val	1260
aca Thr	ggc Gly 320	act Thr	ggt Gly	ggc Gly	atc Ile	cag Gln 325	tgg Trp	tgg Trp	cca Pro	gta Val	gag Glu 330	gag Glu	gag Glu	gtg Val	aac Asn	1308
aag Lys 335	gag Glu	gag Glu	ggt Gly	tct Ser	agt Ser 340	ggc Gly	agc Ser	agt Ser	ggc Gly	agg Arg 345	aac Asn	ccc Pro	caa Gln	gcc Ala	agc Ser 350	1356
ctg Leu	ttt Phe	Gly ggg	aag Lys	aag Lys 355	gcc Ala	aag L <i>ys</i>	gct Ala	cac His	aag Lys 360	gca Ala	ttc Phe	ggc Gly	cag Gln	ccg Pro 365	gca Ala	1404
gac Asp	agg Arg	ccg Pro	cgg Arg 370	gag Glu	cca Pro	ctg Leu	tgg Trp	gcc Ala 375	tac Tyr	ttc Phe	tgt Cys	gac Asp	ttc Phe 380	cgg Arg	gac Asp	1452
atc Ile	acc Thr	cac His 385	gtg Val	gtg Val	ctg Leu	aaa Lys	gag Glu 390	cac His	tgt Cys	gtc Val	agc Ser	atc Ile 395	cac His	cgg Arg	cag Gln	1500
gac Asp	aac Asn 400	aag Lys	tgc Cys	ctg Leu	gag Glu	ctg Leu 405	agc Ser	ttg Leu	cct Pro	tcc Ser	cgg Arg 410	gct Ala	gcg Ala	gcg Ala	ctg Leu	1548
tcc Ser 415	ttc Phe	gtg Val	tcg Ser	ctg Leu	gtg Val 420	gac Asp	Gly ggc	tat Tyr	ttc Phe	cgc Arg 425	ctg Leu	acg Thr	gcc Ala	gac Asp	tcc Ser 430	1596
agc Ser	cac His	tac Tyr	ctg Leu	tgc Cys 435	cac His	gag Glu	gtg Val	gct Ala	ccc Pro 440	cca Pro	cgg Arg	ctg Leu	gtg Val	atg Met 445	agc Ser	1644
atc Ile	cgg Arg	Asp	999 Gly 450	atc Ile	cac His	gga Gly	Pro	ctg Leu 455	ctg Leu	gag Glu	cca Pro	ttt Phe	gtg Val 460	cag Gln	gcc Ala	1692

Figure 9 (cont'd)

aac Lys	g cto	g egg 1 Arg 465	g ccc g Pro	gaç Glu	gac Asp	ggc Gly	cto Leu 470	Tyr	ctc Leu	att	cac His	tgg Trp 475	Ser	acc Thr	agc Ser	174	10
cac His	e eee s Pro 480	туг	cgc Arg	ctg Leu	atc Ile	ctc Leu 485	Thr	gtg Val	gcc Ala	cag Gln	cgt Arg 490	Ser	cag Gln	gca Ala	cca Pro	178	38
gad Asp 499	> Gly	ato Met	cag Gln	agc Ser	ttg Leu 500	cgg Arg	ctc Leu	cga Arg	aag Lys	ttc Phe 505	ccc Pro	att Ile	gag Glu	cag Gln	cag Gln 510	183	36
gac Asp	: ggg : Gly	gcc Ala	ttc Phe	gtg Val 515	Leu	gag Glu	ggc Gly	tgg Trp	ggc Gly 520	cgg Arg	tcc Ser	ttc Phe	CCC Pro	agc Ser 525	gtt Val	188	34
cgg Arg	gaa Glu	ctt Leu	ggg Gly 530	gct Ala	gcc Ala	ttg Leu	cag Gln	ggc Gly 535	tgc Cys	ttg Leu	ctg Leu	agg Arg	gcc Ala 540	GJ A āāā	gat Asp	193	32
gac Asp	tgc Cys	ttc Phe 545	tct Ser	ctg Leu	cgt Arg	cgc Arg	tgt Cys 550	tgc Cys	ctg Leu	ccc Pro	caa Gln	cca Pro 555	gga Gly	gaa Glu	acc Thr	198	10
tcc Ser	aat Asn 560	ctc Leu	atc Ile	atc Ile	atg Met	cgg Arg 565	gly ggg	gct Ala	cgg Arg	gcc Ala	agc Ser 570	ccc Pro	agg Arg	aca Thr	ctc Leu	202	:8
aac Asn 575	ctc Leu	agc Ser	cag Gln	ctc Leu	agc Ser 580	ttc Phe	cac His	cgg Arg	gtt Val	gac Asp 585	cag Gln	aag Lys	gag Glu	atc Ile	acc Thr 590	207	6
cag Gln	ctg Leu	tcc Ser	cac His	ttg Leu 595	Gly ggc	cag Gln	ggc Gly	aca Thr	agg Arg 600	acc Thr	aac Asn	gtg Val	tat Tyr	gag Glu 605	ggc Gly	212	:4
cgc Arg	ctg Leu	cga Arg	gtg Val 610	gag Glu	ggc Gly	agc Ser	ggg Gly	gac Asp 615	cct Pro	gag Glu	gag Glu	ggc Gly	aag Lys 620	atg Met	gat Asp	217	2
Asp	Glu	Asp 625	ccc Pro	Leu	Val	Pro	Gly 630	Arg	Asp	Arg	Gly	Gln 635	Glu	Leu	Arg	222	:0
gtg Val	gtg Val 640	ctc Leu	aaa Lys	gtg Val	Leu	gac Asp 645	ect Pro	agt Ser	cac His	cat His	gac Asp 650	atc Ile	gcc Ala	ctg Leu	gcc Ala	226	-8
Phe 655	Tyr	Glu	aca Thr	Ala	Ser 660	Leu	Met	Ser	Gln	Val 665	Ser	His	Thr	His	Leu 670	231	6
gcc Ala	ttc Phe	gtg Val	cat (ggc Gly 675	gtc Val	tgt Cys	gtg Val	cgc Arg	ggc Gly 680	ect Pro	gaa Glu	aat Asn	agc Ser	atg Met 685	gtg Val	236	4
aca Thr	gag Glu	Тут	gtg (Val (690	gag Glu	cac (His (gga Gly	Pro	ctg Leu 695	gat Asp	gtg Val	tgg Trp	ctg Leu	cgg Arg 700	agg Arg	gag Glu	241	2

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	32/48	

cgg Arg	ggc	cat His 705	gtg Val	ccc Pro	atg Met	gct Ala	tgg Trp 710	aag Lys	atg Met	gtg Val	gtg Val	gcc Ala 715	cag Gln	cag Gln	ctg Leu	2460
gcc Ala	agc Ser 720	gcc Ala	ctc Leu	agc Ser	tac Tyr	ctg Leu 725	gag Glu	aac Asn	aag Lys	aac Asn	ctg Leu 730	gtt Val	cat His	ggt Gly	aat Asn	2508
	tgt Cys															2556
	agc Ser															2604
ctc Leu	tcc Ser	agg Arg	gag Glu 770	gag Glu	cgg Arg	gtg Val	gag Glu	agg Arg 775	atc Ile	ccc Pro	tgg Trp	ctg Leu	gcc Ala 780	ccc Pro	gaa Glu	2652
tgc Cys	cta Leu	cca Pro 785	ggt Gly	G1y 999	gcc Ala	aac Asn	agc Ser 790	cta Leu	agc Ser	acc Thr	gcc Ala	atg Met 795	gac Asp	aag Lys	tgg Trp	2700
GJ A GGG	ttt Phe 800	ggc Gly	gcc Ala	acc Thr	ctc Leu	ctg Leu 805	gag Glu	atc Ile	tgc Cys	ttt Phe	gac Asp 810	gga Gly	gag Glu	gcc Ala	cct Pro	2748
	cag Gln										Phe					2796
cac His	cgg Arg	ctg Leu	ccc Pro	gag Glu 835	ccc Pro	tcc Ser	tgc Cys	cca Pro	cag Gln 840	ctg Leu	gcc Ala	aca Thr	ctc Leu	acc Thr 845	agc Ser	2844
cag . Gln	tgt Cys	ctg Leu	acc Thr 850	tat Tyr	gag Glu	cca Pro	acc Thr	cag Gln 855	agg Arg	Pro	tca Ser	ttc Phe	cgc Arg 860	acc Thr	atc Ile	2892
	cgt Arg															2940
act Thr	gtg Val 880	aac Asn	cgg Arg	gac Asp	tca Ser	ccg Pro 885	gcc Ala	gtc Val	gga Gly	cct Pro	act Thr 890	act Thr	ttc Phe	cac His	aag Lys	2988
	tat Tyr															3036
gtc Val	agc Ser	ttg Leu	tac Tyr	tgc Cys 915	tac Tyr	gat Asp	ccg Pro	acc Thr	aac Asn 920	gac Asp	ggc Gly	act Thr	Gly Ggc	gag Glu 925	atg Met	3084
gtg Va l	gcg Ala	gtg Val	aaa Lys 930	gcc Ala	ctc Leu	aag Lys	gca Ala	gac Asp 935	tgc Cys	ggc Gly	ccc Pro	cag Gln	cac His 940	cgc Arg	tcg Ser	3132

GJ7 ggc	c tgg / Trp	aag Lys 945	cag Gln	gag Glu	att	gac Asp	att Ile 950	Leu	cgc Arg	acg Thr	ctc Leu	tac Tyr 955	cac His	gag Glu	cac His	3180
ato Ile	ato 11e 960	Lys	tac Tyr	aag Lys	Gly	tgc Cys 965	tgc Cys	gag Glu	gac Asp	caa Gln	ggc Gly 970	gag Glu	aag Lys	tcg Ser	ctg Leu	3228
cag Gln 975	Leu	gtc Val	atg Met	gag Glu	tac Tyr 980	gtg Val	ecc	ctg Leu	ggc Gly	agc Ser 985	ctc Leu	cga Arg	gac Asp	tac Tyr	ctg Leu 990	3276
			agc Ser		Gly					Leu					Gln	3324
atc Ile	tgc Cys	gag Glu	ggc Gly 1010	Met	gcc	tat Tyr	ctg Leu	cac His 101	Ala	cac His	gac Asp	tac Tyr	atc Ile 1020	His	cga Arg	3372
gac Asp	cta Leu	gcc Ala 102	gcg Ala 5	cgc Arg	aac Asn	gtg Val	ctg Leu 103	Leu	gac Asp	aac Asn	gac Asp	agg Arg 1039	Leu	gtc Val	aag Lys	3420
atc Ile	ggg Gly 104	Asp	ttt Phe	ggc Gly	cta Leu	gcc Ala 1049	Lys	gcc Ala	gtg Val	ccc	gaa Glu 1050	Gly	cac His	gag Glu	tac Tyr	3468
	Arg		cgc Arg			Gly					Phe					3516
			aag Lys		Tyr					Ala					Ser	3564
ttc Phe	GJA āāā	gtg Val	acc Thr 1090	Leu	tat Tyr	gag Glu	ctg Leu	ctg Leu 1095	Thr	cac His	tgt Cys	gac Asp	tcc Ser 1100	Ser	cag Gln	3612
			acg Thr					Leu			Ile		Gln			3660
		Val	ctg Leu				Glu					Gly				3708
	Arg		gac Asp			Pro					His					3756
			aca Thr		Ala					Thr					Ile	3804
		Leu	aag Lys 1170						Tyr					Pro		3852

gtg ttc agc gtg tgc tga ggcacaatgg cagccctgcc tgggaggact Val Phe Ser Val Cys * 1185	3900
ggaccaggca gtggctgcag agggagcctc ctgctccctg ctccaggatg aaaccaagag ggggatgtca gcctcaccca caccgtgtgc cttactcctg tctagagacc ccacctctgt gaacttattt ttctttcttg gccgtgagcc taaccatgat cttgagggac ccaacatttg taggggcact aatccagccc ttaaatcccc cagcttccaa acttgaggcc caccatctcc	3960 4020 4080 4140
	4176

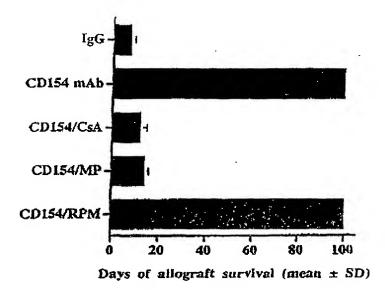


Figure 10

3	Condition	CD154 Expression	Mean
-	•	196	11
	anti-CD3	28%	78
	+ CSA	5%	22
	+ RPM	21%	54
	+ MP	3%	13
	→ MMF	23%	63
-	10	0 10 ¹ 10 ² 10	3

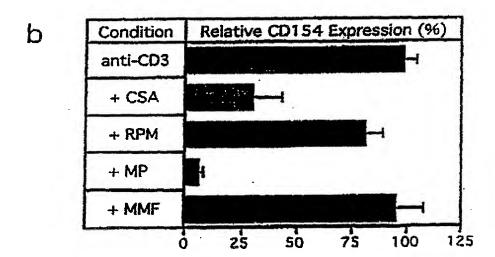


Figure 11

Condition	CD154 Expression	Mean
WT	42%	74
p50 KO	15%	32
WT + LC	22%	40
WT + MG	18%	33

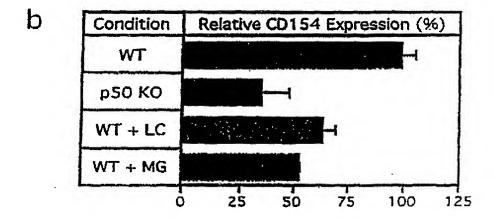


Figure 12

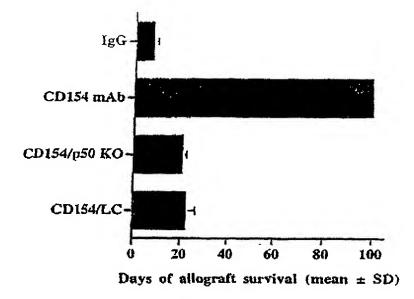


Figure 13

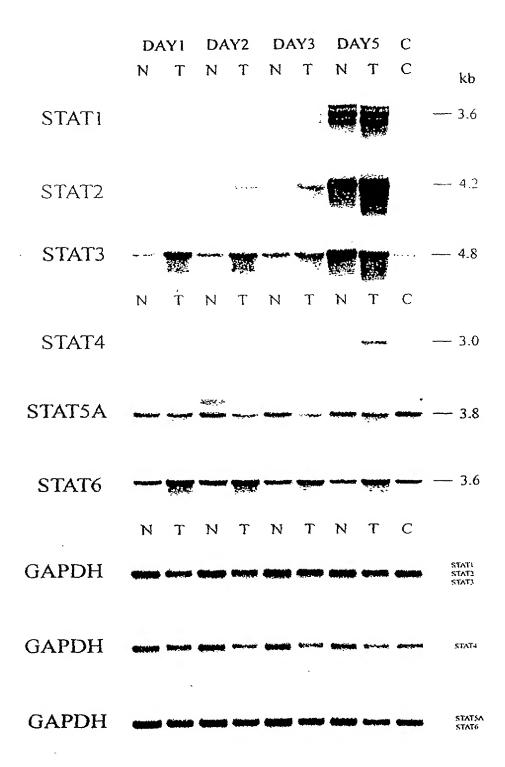
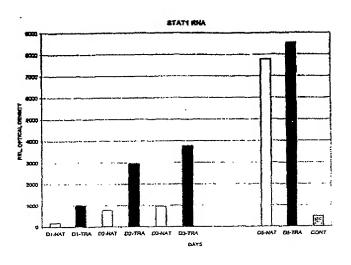
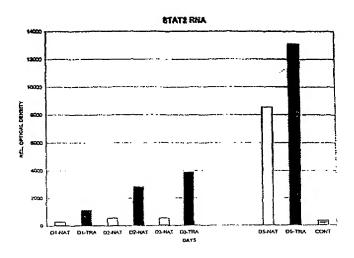
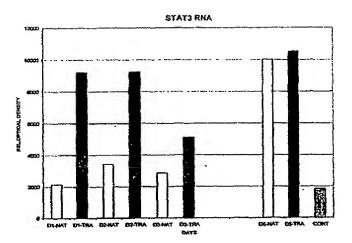
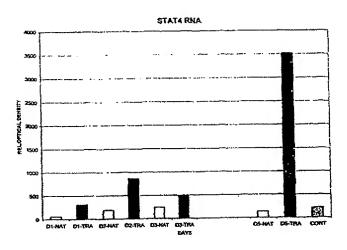


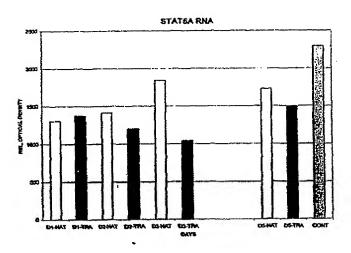
Figure 14A











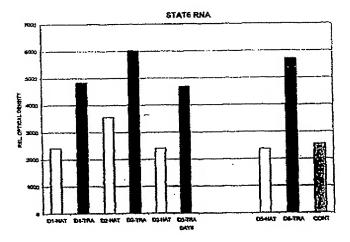


Figure 14B

DAYI DAY2 DAY3 DAY5 C NTNTNT NTC kb SOCS1 -1.5SOCS3 -3.0NTNTNTNTC -- 4.7 SOCS5 CIS - - 2.3 NTNTNTC GAPDH ------GAPDH - CHARLE STATE SOC\$3 SOCSS GAPDH -----CIS

Figure 15

42/48

DAYI DAY2 DAY3 DAY5 C N T N T N T N T C

STAT4

SOCS3

Figure 16

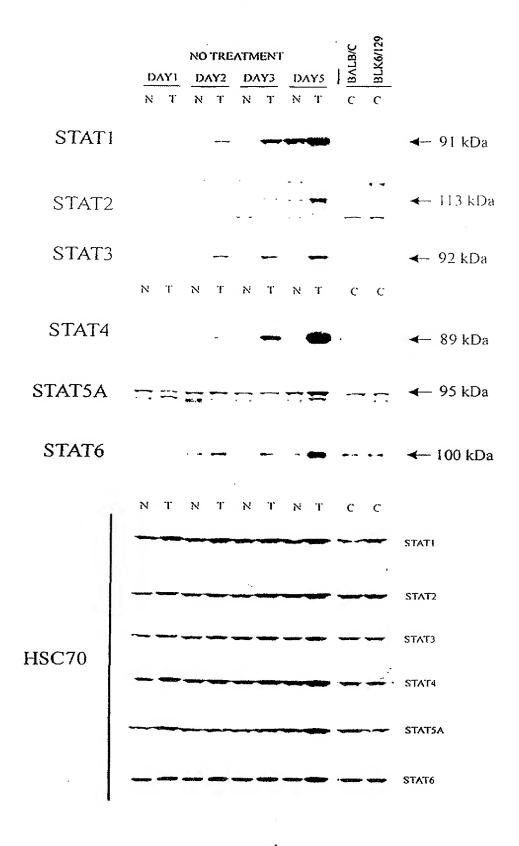


Figure 17

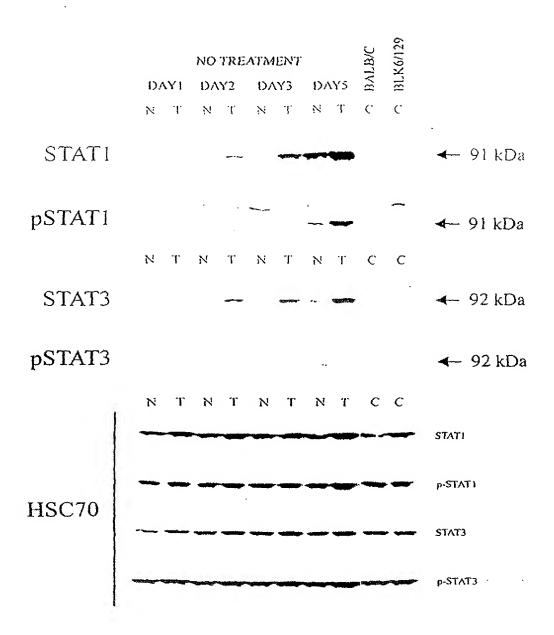


Figure 18

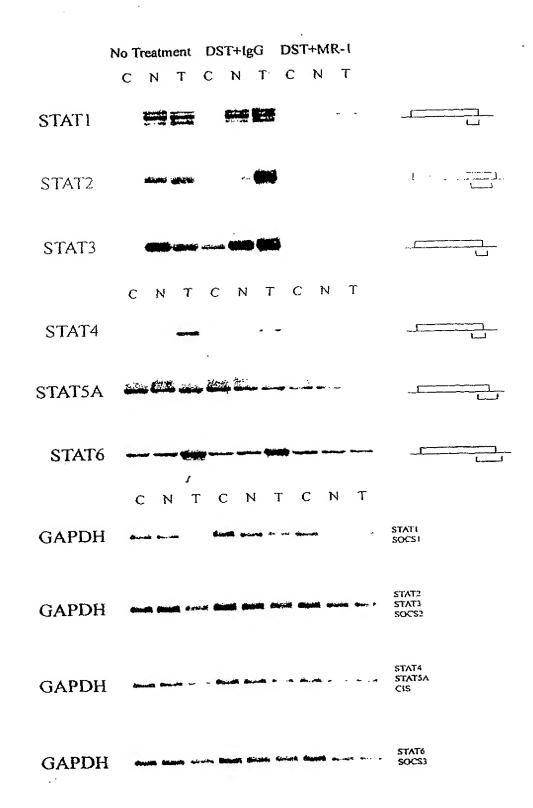


Figure 19

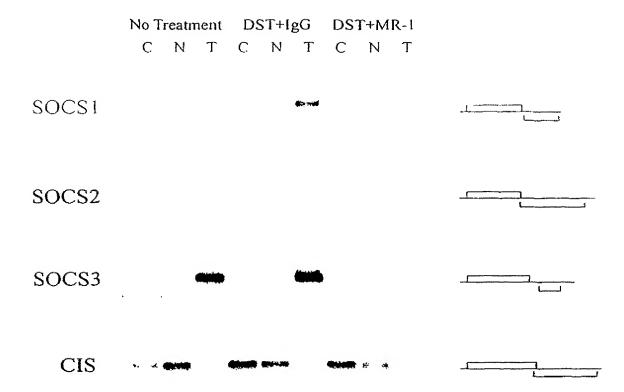


Figure 20

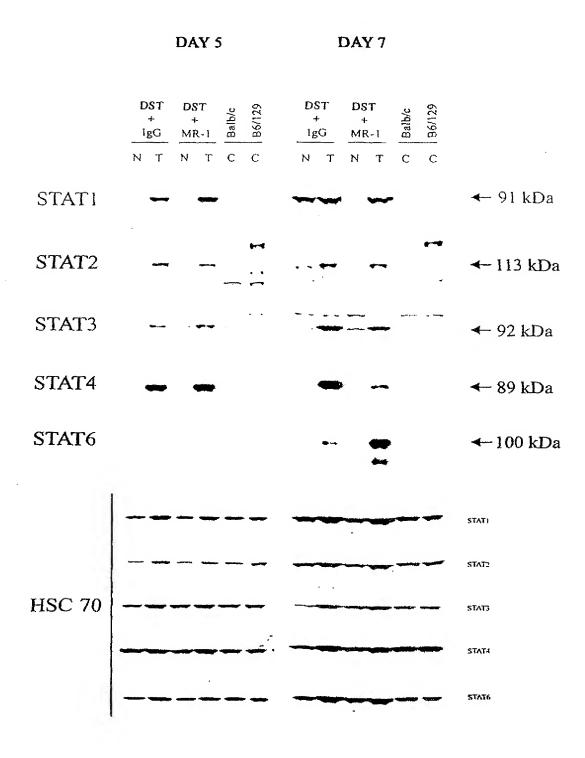


Figure 21

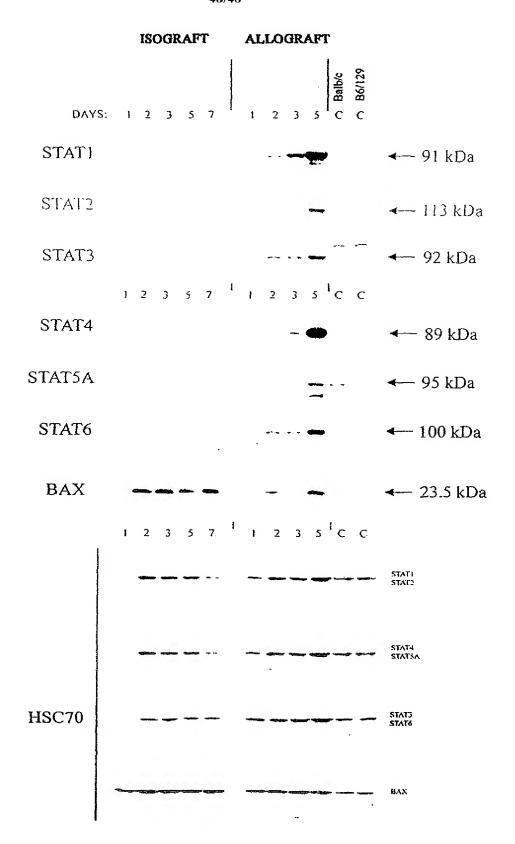


Figure 22

SEQUENCE LISTING

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		140> 141>														
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												agt Ser 25				280
												caa Gln				328
												ttt Phe				376
												ttg Leu				424
												aat Asn				472

					cca											520
Asn	Phe	Gln 95	Glu	Asp	Pro	Ile	Gln 100	Met	Ser	Met	Ile	Ile 105	Tyr	Ser	Суѕ	
					aaa Lys											568
gct Ala 125	cag Gln	tcg Ser	ggg Gly	aat Asn	att Ile 130	cag Gln	agc Ser	aca Thr	gtg Val	atg Met 135	tta Leu	gac Asp	aaa Lys	cag Gln	aaa Lys 140	616
					gtc Val											664
					agc Ser											712
Lys	Cys	Lys 175	Thr	Leu	cag Gln	Asn	Arg 180	Glu	His	Glu	Thr	Asn 185	Gly	Val	Ala	760
ГÀЗ	Ser 190	Asp	Gln	Lys	caa Gln	Glu 195	Gln	Leu	Leu	Leu	Lys 200	Lys	Met	Tyr	Leu	. 808
Met 205	Leu	Asp	Asn	Lys	aga Arg 210	Lys	Glu	Val	Val	His 215	Lys	Ile	Ile	Glu	Leu 220	856
Leu	Asn	Val	Thr	Glu 225	ctt Leu	Thr	Gln	Asn	Ala 230	Leu	Ile	Asn	Asp	Glu 235	Leu	904
gtg Val	gag Glu	tgg Trp	aag Lys 240	cgg Arg	aga Arg	cag Gln	cag Gln	agc Ser 245	gcc Ala	tgt Cys	att Ile	ely aaa	999 Gly 250	ccg Pro	ccc Pro	952
					cag Gln											1000
agt Ser	ctg Leu 270	cag Gln	caa Gln	gtt Val	cgg Arg	cag Gln 275	cag Gln	ctt Leu	aaa Lys	aag Lys	ttg Leu 280	gag Glu	gaa Glu	ttg Leu	gaa Glu	1048
					gaa Glu 290											1096
tta Leu	tgg Trp	gac Asp	cgc Arg	acc Thr 305	ttc Phe	agt Ser	ctt Leu	ttc Phe	cag Gln 310	cag Gln	ctc Leu	att Ile	cag Gln	agc Ser 315	tcg Ser	1144

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		g aag au Lys 35													1240
Val L		g caa au Gln													1288
gat a Asp L 365	aa ga ys As	ıt gtg sp Val	aat Asn	gag Glu 370	aga Arg	aat Asn	aca Thr	gta Val	aaa Lys 375	gga Gly	ttt Phe	agg Arg	aag Lys	ttc Phe 380	1336
		g ggc													1384
		t ctg r Leu 400													1432
		t ggc .a Gly .5													1480
gag ct Glu Lo 4:															1528
gta at Val II 445	tt ga le As	c ctc p Leu	gag Glu	acg Thr 450	acc Thr	tct Ser	ctg Leu	ccc Pro	gtt Val 455	gtg Val	gtg Val	atc Ile	tcc Ser	aac Asn 460	1576
gtc ag Val Se	gc ca er Gl	g ctc n Leu	ccg Pro 465	agc Ser	ggt Gly	tgg Trp	gcc Ala	tcc Ser 470	atc Ile	ctt Leu	tgg Trp	tac Tyr	aac Asn 475	atg Met	1624
		g gaa a Glu 480													1672
gca co Ala Ai		p Ala													1720
gtc ac Val Ti 5:															1768
aag ct Lys Le 525															1816

agg tt Arg Pl															1864
tgg at Trp I															1912
tgg aa Trp As	sn.														1960
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gag ag Glu Se 605															2056
cag as Gln As															2104
aaa ga Lys Gl															2152
gtc at Val Me	et :														2200
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tat at Tyr Il															2344
ctt ca Leu Gl		Thr													2392
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<213> Homo sapiens

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Lys	Arg 210	Lys	Glu	Val	Val	His 215	Lys	Ile	Ile	Glu	Leu 220	Leu	Asn	Val	Thr
225					230					235				Trp	240
Arg	Arg	Gln	Gln	Ser 245	Ala	Cys	Ile	Gly	Gly 250	Pro	Pro	Asn	Ala	Cys 255	Leu
Asp	Gln	Leu	Gln 260	Asn	Trp	Phe	Thr	Ile 265	Val	Ala	Glu	Ser	Leu 270	Gln	Gln
Val	Arg	Gln 275	Gln	Leu	Lys	Lys	Leu 280	Glu	Glu	Leu	Glu	Gln 285	Lys	Tyr	Thr
Tyr	Glu 290	Hìs	Asp	Pro	Ile	Thr 295	Lys	Asn	Lys	Gln	Val 300	Leu	Trp	Asp	Arg
Thr 305	Phe	Ser	Leu	Phe	Gln 310	Gln	Leu	Ile	Gln	Ser 315	Ser	Phe	Val	Val	Glu 320
Arg	Gln	Pro	Cys	Met 325	Pro	Thr	His	Pro	Gln 330	Arg	Pro	Leu	Val	Leu 335	Lys
Thr	Gly	Val	Gln 340	Phe	Thr	Val	Lys	Leu 345	Arg	Leu	Leu	Val	Lys 350	Leu	Gln
Glu	Leu	Asn 355	Tyr	Asn	Leu	Ъуs	Val 360	Lys	Val	Leu	Phe	Asp 365	Lys	Asp	Val
Asn	Glu 370	Arg	Asn	Thr	Val	Lys 375	Gly	Phe	Arg	ГÀЗ	Phe 380	Asn	Ile	Leu	Gly
Thr 385	His	Thr	Lys	Val	Met 390	Asn	Met	Glu	Glu	Ser 395	Thr	Asn	Gly	Ser	Leu 400
Ala	Ala	Glu	Phe	Arg 405	His	Leu	Gln	Leu	Lys 410	Glu	Gln	Ьys	Asn	Ala 415	Gly
Thr	Arg	Thr	Asn 420	Glu	Gly	Pro	Leu	Ile 425	Val	Thr	Glu	Glu	Leu 430	His	Ser
Leu	Ser	Phe 435	Glu	Thr	Gln	Leu	Cys 440	Gln	Pro	Gly	Leu	Val 445	Ile	Asp	Leu
Glu	Thr 450	Thr	Ser	Leu	Pro	Val 455	Val	Val	Ile	Ser	Asn 460	Val	Ser	Gln	Leu
465		_	_		470			_	_	475				Ala	480
				485					490					Trp 495	
			500				***	505					510	Lys	
_		515					520					525		Leu	
	530					535				,	540	_		Cys	
G1u 545	Asn	lle	Asn	Asp	цуя 550	Asn	Pne	Pro	Pne	Trp 555	Leu	Trp	TTE	Glu	Ser 560
	Leu	Glu	Leu	Ile 565		Lys	His	Leu	Leu 570		Leu	Trp	Asn	Asp 575	
Cys	Ile	Met	Gly 580	Phe	Ile	Ser	Lys	Glu 585		Glu	Arg	Ala	Leu 590	Leu	Lys
Asp	Gln	Gln 595	Pro	Gly	Thr	Phe	Leu 600	Leu	Arg	Phe	Ser	Glu 605	Ser	Ser	Arg
Glu	Gly 610	Ala	Ile	Thr	Phe	Thr 615	Trp	Val	Glu	Arg	Ser 620	Gln	Asn	Gly	Gly
Glu	Pro	Asp	Phe	His		Val	Glu	Pro	Tyr		Lys	rys	Glu	Leu	
625	77~7	ጠጉ~	Dha	5~~	630	тл~	T7~	71	λ ~~	635	T.T.***	≒7- 7	Mat	ልግ 🗢	640
				645					650					Ala 655 Ile	
σ±u	L. P.		660		بالله تعدي			665	~ J **	سابت لل	-1.	I	670		بيسي

Lys Asp His Ala Phe Gly Lys Tyr Tyr Ser Arg Pro Lys Glu Ala Pro 680 Glu Pro Met Glu Leu Asp Gly Pro Lys Gly Thr Gly Tyr Ile Lys Thr 695 700 Glu Leu Ile Ser Val Ser Glu Val His Pro Ser Arg Leu Gln Thr Thr 710 715 Asp Asn Leu Leu Pro Met Ser Pro Glu Glu Phe Asp Glu Val Ser Arg 725 730 Ile Val Gly Ser Val Glu Phe Asp Ser Met Met Asn Thr Val 745 <210> 3 <211> 3291 <212> DNA <213> Homo sapiens <220> <221> CDS <222> (58)...(2613) <400> 3 aagtegegae cagagecatt ggagggegeg gggaetgeaa eeetaateag ageecaa atg Met gcg cag tgg gaa atg ctg cag aat ctt gac agc ccc ttt cag gat cag 108 Ala Gln Trp Glu Met Leu Gln Asn Leu Asp Ser Pro Phe Gln Asp Gln ctg cac cag ctt tac tcg cac agc ctc ctg cct gtg gac att cga cag 156 Leu His Gln Leu Tyr Ser His Ser Leu Leu Pro Val Asp Ile Arg Gln 20 tac ttg get gte tgg att gaa gac cag aac tgg cag gaa get gea ett 204 Tyr Leu Ala Val Trp Ile Glu Asp Gln Asn Trp Gln Glu Ala Ala Leu 35 ggg agt gat gat tee aag get ace atg eta tte tte cae tte ttq qat 252 Gly Ser Asp Asp Ser Lys Ala Thr Met Leu Phe Phe His Phe Leu Asp 50 55 cag ctg aac tat gag tgt ggc cgt tgc agc cag gac cca gag tcc ttg 300 Gln Leu Asn Tyr Glu Cys Gly Arg Cys Ser Gln Asp Pro Glu Ser Leu 70 ttg ctg cag cac aat ttg cgg aaa ttc tgc cgg gac att cag ccc ttt 348 Leu Leu Gln His Asn Leu Arg Lys Phe Cys Arg Asp Ile Gln Pro Phe tcc cag gat cct acc cag ttg gct gag atg atc ttt aac ctc ctt ctg 396 Ser Gln Asp Pro Thr Gln Leu Ala Glu Met Ile Phe Asn Leu Leu 100 1.05 gaa gaa aaa aga att ttg atc cag gct cag agg gcc caa ttg qaa caa 444 Glu Glu Lys Arg Ile Leu Ile Gln Ala Gln Arg Ala Gln Leu Glu Gln 115 120

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gaa Glu	tcc Ser	cgg Arg	atc Ile	ctg Leu 150	gat Asp	tta Leu	agg Arg	gct Ala	atg Met 155	atg Met	gag Glu	aag Lys	ctg Leu	gta Val 160	aaa Lys	540
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aaa Lys	gag Glu 195	cag Gln	aag Lys	att Ile	ctg Leu	cag Gln 200	gaa Glu	act Thr	ctc Leu	aat Asn	gaa Glu 205	ctg Leu	gac Asp	aaa Lys	agg Arg	684
aga Arg 210	aag Lys	gag Glu	gtg Val	ctg Leu	gat Asp 215	gcc Ala	tcc Ser	aaa Lys	gca Ala	ctg Leu 220	cta Leu	Gly ggc	cga Arg	tta Leu	act Thr 225	732
acc Thr	cta Leu	atc Ile	gag Glu	cta Leu 230	ctg Leu	ctg Leu	cca Pro	aag Lys	ttø Le 235	gag Glu	gag Glu	tgg Trp	aag Lys	gcc Ala 240	cag Gln	780
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tta Leu 370	Gln	ggc Gly	ttc Phe	cgg Arg	aag Lys 375	ttc Phe	aac Asn	att Ile	ctg Leu	act Thr 380	tca Ser	aac Asn	cag Gln	aaa Lys	act Thr 385	1212
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tac Tyr	ctg Leu	act Thr	ctg Leu 405	gtg Val	gag Glu	caa Gln	cgt Arg	tca Ser 410	ggt Gly	ggt Gly	tca Ser	gga Gly	aag Lys 415	ggc Gly	agc Ser	1308
aat Asn	aag Lys	999 Gly 420	cca Pro	cta Leu	ggt Gly	gtg Val	aca Thr 425	gag Glu	gaa Glu	ctg Leu	cac His	atc Ile 430	atc Ile	agc Ser	ttc Phe	1356
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acc Thr 450	ctc Leu	cct Pro	gtg Val	gtg Val	att Ile 455	att Ile	tcc Ser	aac Asn	atg Met	aac Asn 460	cag Gln	ctc Leu	tca Ser	att Ile	gcc Ala 465	1452
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tca Ser	gac Asp 515	cag Gln	ctg Leu	agc Ser	atg Met	ctg Leu 520	aga Arg	aac Asn	aag Lys	ctg Leu	ttc Phe 525	elà aaa	cag Gln	aac Asn	tgt Cys	1644
agg Arg 530	act Thr	gag Glu	gat Asp	cca Pro	tta Leu 535	ttg Leu	tcc Ser	tgg Trp	gct Ala	gac Asp 540	ttc Phe	act Thr	aag Lys	cga Arg	gag Glu 545	1692
agc Ser	cct Pro	cct Pro	ggc Gly	aag Lys 550	tta Leu	cca Pro	ttc Phe	tgg Trp	aca Thr 555	tgg Trp	ctg Leu	gac Asp	aaa Lys	att Ile 560	ctg Leu	1740
gag Glu	ttg Leu	gta Val	cat His 565	gac Asp	cac His	ctg Leu	aag Lys	gat Asp 570	ctc Leu	tgg Trp	aat Asn	gat Asp	gga Gly	cgc Arg	atc Ile	1788

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	-				ctc Leu 695											2172
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															gag Glu	2460

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aat ggt gac cca ctg ttg gct ggc cag aac acc gtg gat gag gtt tac Asn Gly Asp Pro Leu Leu Ala Gly Gln Asn Thr Val Asp Glu Val Tyr 820 825 830	2556
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_		_		_					_						
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Arg	Arg 210	Lys	Glu	Val	Leu	Asp 215	Ala	Ser	Lys	Ala	Leu 220	Leu	Gly	Arg	Leu
Thr	Thr	Leu	Ile	Glu		Leu	Leu	Pro	Lys	Leu	Glu	Glu	Trp	Lys	
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Gln	Gln	Gln	Lys	Ala 245	Cys	Ile	Arg	Ala	Pro 250	Ile	Asp	His	Gly	Leu 255	Glu
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Gln	Asp 290		Pro	Leu	Thr	Lys 295		Val	Asp	Leu	Arg 300		Ala	Gln	Val
Thr		ĭ. ם .11	T.e.13	Gl m	Δra		ĩ.eu	нiс	Ara	Zla		Ta1	Val	CI 11	ጥክኮ
305	CIU	,1.5 C- LA	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	Q.1.1.1	310		J V.	11713	W. A	315	1110	V CL L	V	G T G	320
	Pro	Cvs	Met	Pro		Thr	Pro	His	Ara		Len	Tle	Leu	Tivs	
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Phe	Thr	Val 435	Lys	Tyr	Thr	Tyr	Gln 440	Gly	Leu	Lys	Gln	Glu 445	Leu	Lys	Thr
Asp	Thr 450		Pro	Val	Val	Ile 455		Ser	Asn	Met	Asn 460		Leu	Ser	Ile
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Asn	Gin	Gin	Pne	485	ser	Asn	Pro	Pro	ьуs 490	Ala	Pro	ı,r.b	Ser	ьец 495	Leu
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545	~ 5		*** 7	TT 4	550	w=1	*	+		555		70	33	~ 7	560
Leu	GIU	цец	vaı	H1S	Asp	HIS	теп	гур	570	Leu	Trp	Asn	Asp	575	Arg
Ile	Met	Gly	Phe 580	Val	Ser	Arg	Ser	Gln 585	Glu	Arg	Arg	Leu	Leu 590	Lys	Lys
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Gly			Cys	Ser	Trp	Val 615		His	Gln	Asp	Asp 620		Lys	Val	Leu
	610 Tyr	Ser	Val	Gln			Thr	Ьуs	Glu			Gln	Ser	Leu	
625	_,		 -7		630	** *		~~	_	635		~-		_	640
Leu	Thr	GLu	Ile	11e 645	Arg	His	Tyr	Gln	Leu 650	Leu	Thr	Glu	Glu	Asn 655	Ile

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Ala Phe Gly Cys Tyr Tyr Gln Glu Lys Val Asn Leu Gln Glu Arg Arg
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Lys Tyr Leu Lys His Arg Leu Ile Val Val Ser Asn Arg Gln Val Asp
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Leu Glu Pro Leu Leu Lys Ala Gly Leu Asp Leu Gly Pro Glu Leu Glu
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Ser Val Leu Glu Ser Thr Leu Glu Pro Val Ile Glu Pro Thr Leu Cys
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Met Val Ser Gln Thr Val Pro Glu Pro Asp Gln Gly Pro Val Ser Gln
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Pro Val Pro Glu Pro Asp Leu Pro Cys Asp Leu Arg His Leu Asn Thr
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Glu Pro Met Glu Ile Phe Arg Asn Cys Val Lys Ile Glu Glu Ile Met
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Pro Asn Gly Asp Pro Leu Leu Ala Gly Gln Asn Thr Val Asp Glu Val
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                                             Met Ala Gln Trp Asn
cag cta cag cag ctt gac aca cgg tac ctg gag cag ctc cat cag ctc
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Gln Leu Gln Gln Leu Asp Thr Arg Tyr Leu Glu Gln Leu His Gln Leu
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tac agt gac agc ttc cca atg gag ctg cgg cag ttt ctg gcc cct tgg
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Tyr Ser Asp Ser Phe Pro Met Glu Leu Arg Gln Phe Leu Ala Pro Trp
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Ile Glu Ser Gln Asp Trp Ala Tyr Ala Ala Ser Lys Glu Ser His Ala
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	cgg Arg															571
	gca Ala															619
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	gga Gly															811
	aag Lys															859
	aga Arg 215															907
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	caa Gln															1003
	cta Leu															1051

		caa Gln 280														1099
		gac Asp														1147
		ctg Leu														1195
		tgc Cys														1243
		cag Gln														1291
		tat Tyr 360														1339
		gca Ala														1387
		aaa Lys														1435
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		gcc Ala														1531
		acc Thr 440														1579
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		aag Lys														1723

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		tgt Cys														2395

atg too coc ogo got tta gat toa ttg atg cag ttt gga aat aat ggt Met Ser Pro Arg Ala Leu Asp Ser Leu Met Gln Phe Gly Asn Asn Gly 730 735 740	2443
gaa ggt gct gaa ccc tca gca gga ggg cag ttt gag tcc ctc acc ttt Glu Gly Ala Glu Pro Ser Ala Gly Gly Gln Phe Glu Ser Leu Thr Phe 745 750 755	2491
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<213> Homo sapiens

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Ser Gln Leu Gln Thr Arg Gln Gln Ile Lys Lys Leu Glu Glu Leu His Gln Lys Val Ser Tyr Lys Gly Asp Pro Ile Val Gln His Arg Pro Met Leu Glu Glu Arg Ile Val Glu Leu Phe Arg Asn Leu Met Lys Ser Ala Phe Val Val Glu Arg Gln Pro Cys Met Pro Met His Pro Asp Arg Pro Leu Val Ile Lys Thr Gly Val Gln Phe Thr Thr Lys Val Arg Leu Leu Val Lys Phe Pro Glu Leu Asn Tyr Gln Leu Lys Ile Lys Val Cys Ile Asp Lys Asp Ser Gly Asp Val Ala Ala Leu Arg Gly Ser Arg Lys Phe Asn Ile Leu Gly Thr Asn Thr Lys Val Met Asn Met Glu Glu Ser Asn Asn Gly Ser Leu Ser Ala Glu Phe Lys His Leu Thr Leu Arg Glu Gln Arg Cys Gly Asn Gly Gly Arg Ala Asn Cys Asp Ala Ser Leu Ile Val Thr Glu Glu Leu His Leu Ile Thr Phe Glu Thr Glu Val Tyr His Gln Gly Leu Lys Ile Asp Leu Glu Thr His Ser Leu Ser Val Val Val Ile Ser Asn Ile Cys Gln Met Pro Asn Ala Trp Ala Ser Ile Leu Trp Tyr Asn Met Leu Thr Asn Asn Pro Lys Asn Val Asn Phe Phe Thr Lys Pro 4.85 Pro Ile Gly Thr Trp Asp Gln Val Ala Glu Val Leu Ser Trp Gln Phe Ser Ser Thr Thr Lys Arg Gly Leu Ser Ile Glu Gln Leu Thr Thr Leu Ala Glu Lys Leu Leu Gly Pro Gly Val Asn Tyr Ser Gly Cys Gln Ile Thr Trp Ala Asn Phe Cys Lys Glu Asn Met Ala Gly Lys Gly Phe Ser Tyr Trp Val Trp Leu Asp Asn Ile Ile Asp Leu Val Lys Lys Tyr Ile Leu Ala Leu Trp Asn Glu Gly Tyr Ile Met Gly Phe Ile Ser Lys Glu Arg Glu Arg Ala Ile Leu Ser Thr Lys Pro Pro Gly Thr Phe Leu Leu Arg Phe Ser Glu Ser Ser Lys Glu Gly Gly Val Thr Phe Thr Trp Val Glu Lys Asp Ile Ser Gly Lys Thr Gln Ile Gln Ser Val Glu Pro Tyr Thr Lys Gln Gln Leu Asn Asn Met Ser Phe Ala Glu Ile Ile Met Gly Tyr Lys Ile Met Asp Ala Thr Asn Ile Leu Leu Ser Pro Leu Val Tyr Leu Tyr Pro Asp Ile Pro Lys Glu Glu Ala Phe Gly Lys Tyr Cys Arg Pro Glu Ser Gln Glu His Pro Glu Ala Asp Pro Gly Ser Ala Ala Pro Tyr Leu Lys Thr Lys Phe Ile Cys Val Thr Pro Thr Thr Cys Ser Asn Thr Ile Asp Leu Pro Met Ser Pro Arg Ala Leu Asp Ser Leu Met Gln

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			340					345					350	Pro	
		355					360				_	365		Val	
	370			_	_	375					380			Lys	
Met 385	ser	TIE	GIU	GIU	390	ser	Asn	GTA	ser	ьеи 395	ser	vaı	GIU	Phe	Arg 400
His	Leu	Gln	Pro	Lys 405	Glu	Met	Lys	Ser	Ser 410	Ala	Gly	Gly	Lys	Gly 415	Asn
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465 Val	Phe	Phe	Asn	Asn 485	470 Pro	Pro	Pro	Ala	Thr 490	475 Leu	Ser	Gln	Leu	Leu 495	480 Glu
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Asp	Gln	Leu 515	His	Met	Leu	Ala	Glu 520	Lys	Leu	Thr	Val	Gln 525	Ser	Ser	Tyr	
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Ile	Leu	Arg	Asp	Tyr 645	Lys	Val	Ile	Met	Ala 650	Glu	Asn	Ile	Pro	Glu 655	Asn	
Pro	Leu	Lys	Tyr 660	Leu	Tyr	Pro	Asp	Ile 665	Pro	Lys	Asp	Lys	Ala 670	Phe	Gly	
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Arg 705	Ser	Asp	Ser	Thr	Glu 710	Pro	His	Ser	Pro	Ser 715	Asp	Leu	Leu	Pro	Met 720	
Ser	Pro	Ser	Val	Tyr 725	Ala	Val	Leu	Arg	Glu 730	Asn	Leu	Ser	Pro	Thr 735	Thr	
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	_				-			_			_		ctc		_	225
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													tgc Cys 50			321

gct Ala	agt Ser	gcc Ala 55	cta Leu	ctt Leu	tca Ser	gac Asp	act Thr 60	gtc Val	cag Gln	cac His	ctt Leu	cag Gln 65	gcc Ala	tcg Ser	gtg Val	369
	gag Glu 70															417
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caa Gln	ata Ile	ctt Leu	caa Gln	gga Gly 105	gag Glu	aaa Lys	aaa Lys	gct Ala	gtt Val 110	atg Met	gaa Glu	cag Gln	ttc Phe	cgc Arg 115	cac Hìs	513
ttg Leu	cca Pro	atg Met	cct Pro 120	ttc Phe	cac His	tgg Trp	aag Lys	cag Gln 125	gaa Glu	gaa Glu	ctc Leu	aag Lys	ttt Phe 130	aag Lys	aca Thr	561
Gly	ttg Leu	cgg Arg 135	agg Arg	ctg Leu	cag Gln	cac His	cga Arg 140	gta Val	gjå aaa	gag Glu	atc Ile	cac His 145	ctt Leu	ctc Leu	cga Arg	609
	gcc Ala 150															657
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gtg Val	ctg Leu	aag Lys	agg Arg 200	atc Ile	cag Gln	att Ile	tgg Trp	aaa Lys 205	cgg	cag Gln	cag Gln	cag Gln	ctg Leu 210	gca Ala	eli aaa	801
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gct Ala 245	ggt	ej aaa	gag Glu	ctt Leu	gag Glu 250	ccc Pro	aag Lys	acc Thr	cgg Arg	gca Ala 255	tcg Ser	ctg Leu	act Thr	gly ggc	cgg Arg 260	945
ctg Leu	gat Asp	gaa Glu	Val	ctg Leu 265	aga Arg	acc Thr	ctc Leu	gtc Val	acc Thr 270	agt Ser	tgc Cys	ttc Phe	ctg Leu	gtg Val 275	gag Glu	993

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		ctg Leu								1089
		agg Arg								1137
		cag Gln								1185
		act Thr								1233
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		ctg Leu								1425
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		gag Glu								1617
		agt Ser							tcg Ser 500	1665

		aag Lys											1713
		ggt Gly 520											1761
		cgg Arg											1809
		ctc Leu											1857
		att Ile											1905
		cca Pro											1.953
		egc Arg 600											2001
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		atg Met											2145
		cct Pro											2193
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atg cce Met Pro 725															2385
ccc caq Pro Gli															2433
cct gad Pro Asp															2481
ctg age Leu Se															2529
gac ata Asp Ile 790	e Phe														2577
ctt cto Leu Leu 805															2625
cag cco Gln Pro															2673
tcc cac Ser His										tga *	tcc	cagc	tgg		2719
agggaga tcatgco caggago ctatcat acacgco tctctga	ectg o gaaa a etcc o	caag agact cctgo acatg	aaca caaca ccaa gcctq	ca ga ag ga cc to gc ao	atggg agaat cctto cctgo	geage ceage agege	g gtg c agt c act	gccci cgggi	cct tgga tgga	atco gcca aggg	ccca aatc gaag	cct a cac : ttc a	actco tecti aggei	ctgggt tccttt tctgag	2779 2839 2899 2959 3019 3046

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 Trp
 Gly
 Leu
 Val
 Ser
 Lys
 Met
 Pro
 Pro
 Glu
 Lys
 Val
 Gln
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 Lu
 Lu

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Gln	Phe	Arg 115	His	Leu	Pro	Met	Pro 120	Phe	His	Trp	Lys	Gln 125	Glu	Glu	Leu
Lys	Phe 130	Lys	Thr	Gly	Leu	Arg 135	Arg	Leu	Gln	His	Arg 140	Val	Gly	Glu	Ile
His 145	Leu	Leu	Arg	Glu	Ala 150	Leu	Gln	Lys	Gly	Ala 155	Glu	Ala	Gly	Gln	Val 160
Ser	Leu	His	Ser	Leu 165	Ile	Glu	Thr	Pro	Ala 170	Asn	Gly	Thr	Gly	Pro 175	Ser
Glu	Ala	Leu	Ala 180	Met	Leu	Leu	Gln	Glu 185	Thr	Thr	Gly	Glu	Leu 190	Glu	Ala
Ala	Lys	Ala 195	Leu	Val	Leu	Lys	Arg 200	Ile	Gln	Ile	Trp	Lys 205	Arg	Gln	Gln
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				245					250					Ala 255	
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	290					295					300			Gly	
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	370					375					380			Lys	_
385					390					395				Leu	400
				405					410					His 415	-
			420					425				-	430	Ala	
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	450					455					460			Val	_
465					470					475				Gln	480
				485					490					Arg 495	
			500					505					510	Gly	
		515					520					525		Arg	
	530					535					540			Ser	_
Gln 545	Tyr	Val	Thr	Ser	Leu 550	Leu	Leu	Asn	Glu	Pro 555	Asp	Gly	Thr	Phe	Leu 560

Leu Arg Phe Ser Asp Ser Glu Ile Gly Gly Ile Thr Ile Ala His Val 565 570 Ile Arg Gly Gln Asp Gly Ser Pro Gln Ile Glu Asn Ile Gln Pro Phe 585 Ser Ala Lys Asp Leu Ser Ile Arg Ser Leu Gly Asp Arg Ile Arg Asp 600 Leu Ala Gln Leu Lys Asn Leu Tyr Pro Lys Lys Pro Lys Asp Glu Ala 615 Phe Arg Ser His Tyr Lys Pro Glu Gln Met Gly Lys Asp Gly Arg Gly 630 635 Tyr Val Pro Ala Thr Ile Lys Met Thr Val Glu Arg Asp Gln Pro Leu 645 650 Pro Thr Pro Glu Leu Gln Met Pro Thr Met Val Pro Ser Tyr Asp Leu 665 Gly Met Ala Pro Asp Ser Ser Met Ser Met Gln Leu Gly Pro Asp Met 680 Val Pro Gln Val Tyr Pro Pro His Ser His Ser Ile Pro Pro Tyr Gln 700 695 Gly Leu Ser Pro Glu Glu Ser Val Asn Val Leu Ser Ala Phe Gln Glu 710 715 Pro His Leu Gln Met Pro Pro Ser Leu Gly Gln Met Ser Leu Pro Phe 725 730 Asp Gln Pro His Pro Gln Gly Leu Leu Pro Cys Gln Pro Gln Glu His 745 Ala Val Ser Ser Pro Asp Pro Leu Leu Cys Ser Asp Val Thr Met Val 760 Glu Asp Ser Cys Leu Ser Gln Pro Val Thr Ala Phe Pro Gln Gly Thr 775 780 Trp Ile Gly Glu Asp Ile Phe Pro Pro Leu Deu Pro Pro Thr Glu Gln 790 795 Asp Leu Thr Lys Leu Leu Glu Gly Gln Gly Glu Ser Gly Gly Gly 805 810 Ser Leu Gly Ala Gln Pro Leu Gln Pro Ser His Tyr Gly Gln Ser 820 825 Gly Ile Ser Met Ser His Met Asp Leu Arg Ala Asn Pro Ser Trp 835 840 <210> 11 <211> 908 <212> DNA <213> Homo sapiens <220> <221> CDS <222> (14)...(649) <400> 11 ccccttctgt agg atg gta gca cac aac cag gtg gca gcc gac aat gca 49 Met Val Ala His Asn Gln Val Ala Ala Asp Asn Ala gto too aca goa goa gag coo cga cgg cga cca gaa cot too too tot 97 Val Ser Thr Ala Ala Glu Pro Arg Arg Pro Glu Pro Ser Ser Ser 15 20 tee tee tee teg eee geg gee eee geg ege eeg egg eeg tge eee geg 145

Ser Ser Ser Pro Ala Ala Pro Ala Arg Pro Arg Pro Cys Pro Ala

35

30

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cgg Arg	aac Asn 110	tgc Cys	ttt Phe	ttc Phe	gcc Ala	ctt Leu 115	agc Ser	gtg Val	aag Lys	atg Met	gcc Ala 120	tcg Ser	gga Gly	ccc Pro	acg Thr	385
agc Ser 125	atc Ile	cgc Arg	gtg Val	cac His	ttt Phe 130	cag Gln	gcc Ala	gly	cgc Arg	ttt Phe 135	cac His	ctg Leu	gat Asp	ggc Gly	agc Ser 140	433
cgc Arg	gag Glu	agc Ser	ttc Phe	gac Asp 145	tgc Cys	ctc Leu	ttc Phe	gag Glu	ctg Leu 150	ctg Leu	gag Glu	cac His	tac Tyr	gtg Val 155	gcg Ala	481
			ege Arg 160													529
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ccto	iggtt igccg	gg a	raaas	gegç ette	ga to et ca	ggt:	gtago ettga	. 336 3 336	gagg gggt	gege cete	ctc	cege	cct (tgg i	egget	ccccgg tggaga ccctct	739 799 859 908

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Pro	Ala	Ala 35	Pro	Ala	Arg	Pro	Arg 40	Pro	Суѕ	Pro	Ala	Val 45	Pro	Ala	Pro	
Ala	Pro 50	Gly	Asp	Thr	His	Phe 55	Arg	Thr	Phe	Arg	Ser 60	His	Ala	Asp	Tyr	
Arg 65	Arg	Ile	Thr	Arg	Ala 70	Ser	Ala	Leu	Leu	Asp 75	Ala	Cys	Gly	Phe	Tyr 80	
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Pro	Val	Gly	Thr 100	Phe	Leu	Val	Arg	Asp 105	Ser	Arg	Gln	Arg	Asn 110	Cys	Phe	
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His	Phe 130	Gln	Ala	Gly	Arg	<i>P</i> he 135	His	Leu	Asp	Gly	Ser 140	Arg	Glu	Ser	Phe	
Asp 145	Сув	Leu	Phe	Glu	Leu 150	Leu	Glu	His	Tyr	Val 155	Ala	Ala	Pro	Arg	Arg 160	
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Phe	Gln 210	Ile														
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			1. 1. ±		_ •											_ 0
gcc Ala 65	Gly	acc	Phe	Leu	Ile 70	cgc Arg	gac Asp	agc Ser	ccg Ser	gac Asp 75	cag Gln	cgc Arg	cac His	rtc Phe	Phe 80	240

acg Thr	ct <i>c</i> Leu	agc Ser	gtc Val	aag Lys 85	acc Thr	cag Gln	tct Ser	G1Y 999	acc Thr 90	aag Lys	aac Asn	ctg Leu	cgc Arg	atc Ile 95	cag Gln	288
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ccg Pro	ccc Pro 130	cct Pro	gga Gly	gcc Ala	ccc Pro	tcc Ser 135	ttc Phe	ccc	tcg Ser	cca Pro	cct Pro 140	act Thr	gaa Glu	ecc Pro	tcc Ser	432
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 Arg
 Pro
 Leu

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 Leu
 Lys
 Thr
 Phe
 Ser
 Lys
 Ser
 Glu
 Tyr
 Gln

 Leu
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 Ala
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 Arg
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 Ala
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 Arg
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 Leu
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 Ser
 Ala
 Glu
 Phe

 For
 Frame
 Frame
 Arg
 Arg

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Cys Glu Gly Gly Ser Phe Ser Leu Gln Ser Asp Pro Arg Ser Thr Gln
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                                105
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Pro Pro Pro Gly Ala Pro Ser Phe Pro Ser Pro Pro Thr Glu Pro Ser
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Pro Arg Arg Ala Tyr Tyr Ile Tyr Ser Gly Glu Lys Ile Pro Leu
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Val Leu Ser Arg Pro Leu Ser Ser Asn Val Ala Thr Leu Gln His Leu
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Cys Arg Lys Thr Val Asn Gly His Leu Asp Ser Tyr Glu Lys Val Thr
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Gly Met Ala Cys Leu Thr Met Thr Glu Met Glu Gly Thr Ser Thr Ser
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Ser Ile Tyr Gln Asn Gly Asp Ile Ser Gly Asn Ala Asn Ser Met Lys
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caa ata gat cca gtt ctt cag gtg tat ctt tac cat tcc ctt ggg aaa
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Gln Ile Asp Pro Val Leu Gln Val Tyr Leu Tyr His Ser Leu Gly Lys
    35
tet gag gea gat tat etg ace ttt eea tet ggg gag tat gtt gea gaa
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Ser Glu Ala Asp Tyr Leu Thr Phe Pro Ser Gly Glu Tyr Val Ala Glu
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			gtc Val													816
			aga Arg													864
			cgg Arg													912
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			gcc Ala 245													1248
			ctg Leu										-			1296
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	_		-					• • • • • • • • • • • • • • • • • • • •				tta Leu		1584
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	tgt Cys 675															2544
	aat Asn															2592
	ttg Leu															2640
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gaa Glu 930	tat Tyr	tta Leu	cca Pro	tat Tyr	gga Gly 935	agt Ser	tta Leu	cga Arg	gac Asp	tat Tyr 940	ctt Leu	caa Gln	aaa Lys	cat His	aaa Lys 945	3312
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Ile	Asp 370	Gly	Tyr	Tyr	Arg	Leu 375	Thr	Ala	qaA	Ala	His 380	His	Tyr	Leu	Cys
Lys 385	Glu	Val	Ala	Pro	Pro 390	Ala	Val	Leu	Glu	Asn 395	Ile	Gln	Ser	Asn	Cys 400
His	Gly	Pro	Ile	Ser 405	Met	Asp	Phe	Ala	Ile 410	Ser	Lys	Leu	Lys	Lys 415	Ala
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-	450		***			455	-				460		Tyr		
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545					550				_	555			Lys		560
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	770					775	-	-			780		Asn		
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Ile	Ile	Arg	Asp	Leu 805	Asn	Ser	Leu	Phe	Thr 810	Pro	Asp	Tyr	Glu	Leu 815	Leu

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Ser Gly Ala Phe Glu Asp Arg Asp Pro Thr Gln Phe Glu Glu Arg His
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Leu Lys Phe Leu Gln Gln Leu Gly Lys Gly Asn Phe Gly Ser Val Glu
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Met Cys Arg Tyr Asp Pro Leu Gln Asp Asn Thr Gly Glu Val Val Ala
                  870
                                    875
Val Lys Lys Leu Gln His Ser Thr Glu Glu His Leu Arg Asp Phe Glu
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                                890
Arg Glu Ile Glu Ile Leu Lys Ser Leu Gln His Asp Asn Ile Val Lys
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Tyr Lys Gly Val Cys Tyr Ser Ala Gly Arg Arg Asn Leu Lys Leu Ile
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Met Glu Tyr Leu Pro Tyr Gly Ser Leu Arg Asp Tyr Leu Gln Lys His
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Lys Glu Arg Ile Asp His Ile Lys Leu Leu Gln Tyr Thr Ser Gln Ile
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Cys Lys Gly Met Glu Tyr Leu Gly Thr Lys Arg Tyr Ile His Arg Asp
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Leu Ala Thr Arg Asn Ile Leu Val Glu Asn Glu Asn Arg Val Lys Ile
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Ser Leu Thr Glu Ser Lys Phe Ser Val Ala Ser Asp Val Trp Ser Phe
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Gly Val Val Leu Tyr Glu Leu Phe Thr Tyr Ile Glu Lys Ser Lys Ser
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์ อีก

120

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agt gag tca tcg Ser Glu Ser Ser 50	Leu Thr Ala				492
aaa gtt ggt ato Lys Val Gly Ile 65					540
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														gcg Ala		1548
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			ttc Phe													1884
			999 939													1932
			tct Ser													1980
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			cac His													2124
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			aaa Lys													2268
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				tgg Trp 710						2460
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				agt Ser						2604
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				tgc Cys						2844
				acc Thr						2892
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Tyr	Val 690	Glu	His	Gly	Pro	Leu 695	Asp	Val	Trp	Leu	Arg 700	Arg	Glu	Arg	Gly
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## (19) World Intellectual Property Organization International Bureau





(43) International Publication Date 25 October 2001 (25.10.2001)

PCT

# (10) International Publication Number WO 01/079555 A3

(51) International Patent Classification⁷: C12Q 1/68, G01N 33/68

(21) International Application Number: PCT/US01/12131

(22) International Filing Date: 13 April 2001 (13.04.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data: 09/549,654 14 April 2000 (14.04.2000) US

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(72) Inventors: HANCOCK, Wayne, William; 301 North Street, Medfield, MA 02052 (US). OZKAYNAK, Engin; 44 Purdue Drive, Milford, MA 01757 (US).

(74) Agents: CORUZZI, Laura, A. et al.; Pennie & Edmonds LLP, 1155 Avenue of the Americas, New York, NY 10036 (US). (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

#### Published:

with international search report

(88) Date of publication of the international search report: 27 February 2003

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: ROLES OF JAK/STAT FAMILY MEMBERS IN TOLERANCE INDUCTION

(57) Abstract: The present invention relates to methods and compositions for reducing immune rejection, for example, transplant or autoimmune disorder-related immune rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can reduce immune rejection. The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is suspected of, being affected by an autoimmune disorder). The present invention is further based, in part, on the discovery, demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue. In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present, e.g., present in an affected tissue.

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According to International Palent Classification (IPC) or to both national classification and IPC						
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Minimum do IPC 7	cumentation searched (classification system followed by classification C12Q	n symbols)				
Documenta	ion searched other than minimum documentation to the extent that su	ech documents are included in the heios se	arcneo			
Electronic d	ata base consulted during the international search (name of data bas	e and, where practical, search terms used	)			
EPO-In	ternal, WPI Data, PAJ, SEQUENCE SEAR	CH, MEDLINE, BIOSIS				
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		,				
	-	/				
X Further documents are listed in the continuation of box C. X Patent family members are listed in annex.						
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consid	nt defining the general state of the an which is not ered to be of particular relevance	cited to understand the principle or the invention				
*E* earlier document but published on or after the international filing date  *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to						
*L* document which may throw doubts on priority_ctalm(s) or involve an inventive step when the document is taken alone which is cited to establish the publication date of another citation or other special reason (as specified)  "Y* document of particular relevance; the claimed invention control to provide the provide step when the						
Cannot be considered to involve an inventive step when the  "O" document referring to an oral disclosure, use, exhibition or other means  acomptine combined with one or more other such document is combined with one or more other such document, such combination being obvious to a person skilled						
"P" document published prior to the International filling date but later than the priority date claimed "&" document member of the same patent family						
Date of the actual completion of the international search  Date of mailing of the international search report						
14 October 2002 25/10/2002						
Name and n	nailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2	Authorized officer				
	NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016  Hagenmaier, S					

Form PCT/ISA/210 (second sheet) (July 1992)

Int onal Application No PCT/US 01/12131

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C.(Continua Category®	ation) DOCUMENTS CONSIDERED TO BE RELEVANT  Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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national application No. PCT/US 01/12131

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	rnational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1.	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. X	Claims Nos.: 57–67 because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:  see FURTHER INFORMATION sheet PCT/ISA/210
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inte	mational Searching Authority found multiple inventions in this international application, as follows:
	see additional sheet
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2, X	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	on Protest  The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet (1)) (July 1998)

## FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-56 (all partially)

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of Stat4 mRNA or Stat4 protein, present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising: determining the amount of Stat4 mRNA or Stat4 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of Stat4 mRNA or Stat4 protein in T-cell samples.

2. Claims: 1-56 (all partially)

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of Stat6 mRNA or Stat6 protein present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising: determining the amount of Stat6 mRNA or Stat6 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of Stat6 mRNA or Stat6 protein in T-cell samples.

3. Claims: 1-20,23-36, 39-46, 49-54 (all partially)

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of SOCS1 mRNA or SOCS1 protein present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising: determining the amount of SOCS1 mRNA or SOCS1 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of SOCS1 mRNA or SOCS1 protein in T-cell samples.

4. Claims: 1-20,23-36, 39-46, 49-54 (all partially)

## FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of SOCS3 mRNA or SOCS3 protein, present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising determining the amount SOCS3 mRNA or SOCS3 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of SOCS3 mRNA or SOCS3 protein in T-cell samples.

page 2 of 2

## FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 57-67

Although claims 57-67 are directed to a method of treatment of the human/animal body, a search could have been carried out based on the compound and its alleged effect. Nevertheless, present claims 57-67 only relate to a compound defined by reference to a desirable characteristic or property, namely a compound able to reduce the level of STAT4 mRNA and/or maintaining or increasing the level of STAT6 mRNA. The claims cover all compounds having this characteristic or property, whereas the application does not provide any support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT for such compounds. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claims also lack clarity (Article 6 PCT). An attempt is made to define the compound by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, no search has been carried out for these claims.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

information on patent family members

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